

**Brief Report**

DOI: 10.5582/ddt.8.76

**Design, synthesis and biological evaluation of 4-chromanone derivatives as  $I_{Kr}$  inhibitors**

Rong Wang, Zhenzhen Liu, Lupei Du\*, Minyong Li\*

*Department of Medicinal Chemistry, Key Laboratory of Chemical Biology (MOE), School of Pharmacy, Shandong University, Ji'nan, Shandong, China.*

**Summary** Cardiac arrhythmia is a major cause of death in the world. Among many delayed rectifier potassium currents, the rapid delayed rectifier K current ( $I_{Kr}$ ) plays an important role in the repolarization of cardiac tissue. The inhibition of  $I_{Kr}$  can delay repolarization and lead to an increase in the QT interval of the electrocardiogram, which is the treatment mechanism of Class III antiarrhythmic agents. Therefore,  $I_{Kr}$  can be considered as the drug target for the treatment of cardiac arrhythmia. In the current study, a series of 4-chromanone compounds (WR1-WR12) were well designed and synthesized as  $I_{Kr}$  inhibitors. The results disclosed that two compounds displayed potent inhibitory activities against  $I_{Kr}$ . Moreover, our structure-activity relationship results might provide necessary information for the rational design of inhibitors for  $I_{Kr}$ .

**Keywords:**  $I_{Kr}$  inhibitors, 4-chromanone, arrhythmia

**1. Introduction**

Potassium ion ( $K^+$ ) channels consist of a ubiquitous family of membrane proteins that play critical roles in a wide variety of physiological processes, such as the regulation of neuronal excitability, cell proliferation, muscle contraction, and insulin secretion (1).  $K^+$  channels have long been attractive targets for the rational drug design, due to their pivotal functions in biological systems (2). So far, various small-molecule compounds and toxins have been discovered as  $K^+$  channel modulators (3).

Multiple delayed rectifier potassium currents play an important role in the repolarization and termination of the cardiac action potential. Inhibition of these potassium currents prolongs action potential duration (4), delays repolarization, and produces an antiarrhythmic effect (5). As the rapid component of cardiac delayed rectifier potassium current, the  $I_{Kr}$  potassium channel is mainly encoded by hERG (human ether-a-go-go-related gene) (6,7), and its

electrophysiological properties can be regulated by its auxiliary subunit KCNE1 and KCNE2 (8). Besides in heart, it was reported that the expressing level of hERG in cancer cells was greatly increased (9). As a result,  $I_{Kr}$  encoded by hERG channel may be a potential cancer therapeutic target (10).

$I_{Kr}$  is highly sensitive to blockade by many structurally diverse compounds (11), such as astemizole, imipramine, and dofetilide (12). However, there is still an urgent demand on  $I_{Kr}$  inhibitors for examining the mechanism of inhibition of  $I_{Kr}$ . In the current research, we designed and synthesized a series of 4-chromanone compounds, and evaluated their inhibitory activity against  $I_{Kr}$  using a radio-ligand based assay. The experimental results revealed that several compounds exhibited respectable activity against  $I_{Kr}$ . Moreover, analysis of the structure-activity relationship on these compounds could contribute to designing new  $I_{Kr}$  blocker for preventing arrhythmia and/or cancer therapy.

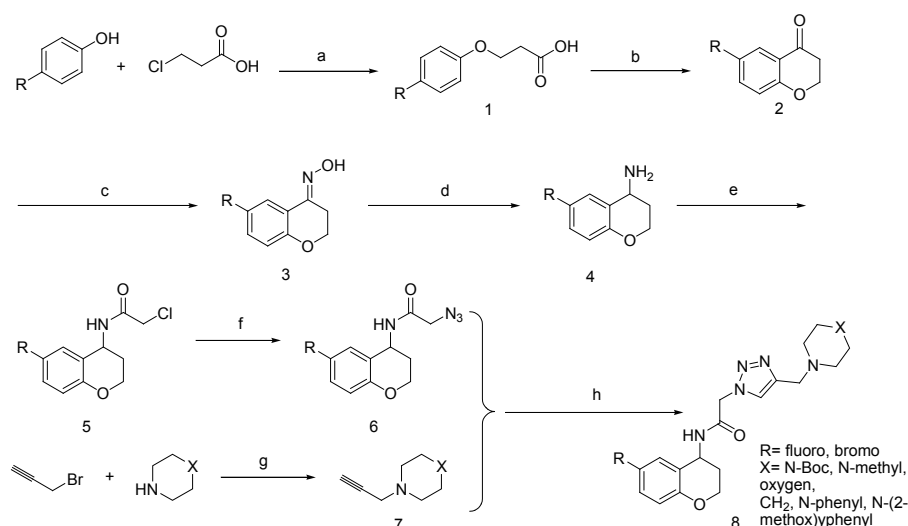
**2. Materials and Methods****2.1. Chemicals**

In summary, a series of 4-chromanone derivatives were well designed and synthesized. The synthetic route is outlined in Scheme 1. In this case, 4-substituted phenol

\*Address correspondence to:

Dr. Minyong Li and Dr. Lupei Du, Department of Medicinal Chemistry, Key Laboratory of Chemical Biology (MOE), School of Pharmacy, Shandong University, Jinan, Shandong 250012, China.

E-mail: mli@sdu.edu.cn (Li MY); dulupei@sdu.edu.cn (Du LP)



**Scheme 1. Synthetic route of WR1-WR12.** Reagents and conditions: (a) KOH, H<sub>2</sub>O, reflux 4 h; (b) PPA, 60°C, 4 h; (c) Hydroxylamine hydrochloride, Na<sub>2</sub>CO<sub>3</sub>, EtOH, H<sub>2</sub>O, 0°C → r.t., 12 h; (d) CH<sub>3</sub>COOH, Zn, 0°C → r.t., 24 h; (e) Chloroacetyl chloride, K<sub>2</sub>CO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 0°C → r.t., 4 h; (f) NaN<sub>3</sub>, KI, CH<sub>3</sub>CN, reflux 6 h; (g) K<sub>2</sub>CO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, 0°C → r.t., 4 h; (h) Sodium ascorbate, CuSO<sub>4</sub>, MeOH, 0°C → r.t., 24 h.

and 3-chloropropionic acid gave compound 1, which was cyclized to form compound 2 by polyphosphoric acid (PPA). Compound 2 is then conveniently converted to compound 3 under oximation reaction. Hydrogenation of compound 3 with CH<sub>3</sub>COOH/Zn gave compound 4, subsequently provided the key intermediate compound 6 *via* acylation with chloroacetyl chloride and substitution with sodium azide. Finally, reaction of compound 6 with the corresponding alkyne presented compound WR1-WR12 *via* click reaction, which was catalyzed by CuSO<sub>4</sub> and sodium ascorbate.

## 2.2. *I<sub>Kr</sub>* inhibition assay

The inhibitory activities of these 12 compounds against *I<sub>Kr</sub>* were evaluated by testing their affinities with hERG in the presence of 9 nM [<sup>3</sup>H] dofetilide. Astemizole (Cat. No. #A2861-10MG; Sigma-Aldrich, St. Louis, MO, USA) and atropin was selected as positive and negative controls, respectively. The affinity with hERG potassium channel was accessed in the presence of 9 nM [<sup>3</sup>H]-dofetilide. Their binding abilities with the hERG were exhibited and compared with the positive and negative controls.

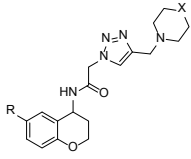
In brief, each compound was dissolved in DMSO as a stock solution (1 mM), which was diluted with binding buffers (10 folds, 6 points) when applied to the binding assays. Cell membranes were prepared as the instruction (GenScript USA Inc.). First, each well of Uni-filter 96 GF/C microplate was incubated with 100 μL 0.5% PEI (Polyethyleneimine, Sigma-Aldrich, dissolved in milli-Q water) at 4°C for 30-60 min. PEI was then discarded, and plates were washed with 2 mL/well wash buffer (50 mM Tris-HCl, pH 7.4; filtered and stored at 4°C). The reaction mixtures,

including membrane (10 μg/well), each compound and [<sup>3</sup>H]-dofetilide ligand (9 nM), were prepared in 24-well plates in a final volume of 100 μL (binding buffer: 10 mM HEPES, 130 mM NaCl, 60 mM KCl, 0.8 mM MgCl<sub>2</sub>, 1 mM NaEGTA, 10 mM glucose, 0.1% BSA, pH7.4; filtered and stored at 4°C) and incubated at 25°C for 2 h with a shaking speed of 530 RPM. The reaction system was transferred into the filter plates and filtered with a Millipore vacuum manifold. The wells were washed with 2 mL/well cold wash buffer and dried at room temperature for 120 min. The bottom of the plates was sealed with Bottom seal™ (opaque) (Perkin Elmer) and 50 μL MicroScint 20™ (Perkin Elmer) was added to each well. Finally, the plates were sealed with Topseal A (Perkin Elmer) and counted on TopCount NXT for 1 min/well. IC<sub>50</sub> values were calculated by GraphPad Prism 4 using the Cheng-Prusoff equation.

The binding data were converted to % displacement according to the below equation: % displacement = 100 × (1 - (sample CPM/Total binding CPM)) (in which total binding CPM values were obtained by testing binding of [<sup>3</sup>H]-dofetilide to the target channel without competitors).

## 3. Results and Discussion

In order to study the influence of different substituents in the phenyl ring and piperazine ring in 4-chromanone-based compounds on the inhibitory activity, we design and synthesize twelve compounds (WR1-WR12) with different R and X group to examine the importance of phenyl ring and piperazine ring, respectively. All inhibition results are presented in Table 1. No compound exhibits a comparable replacement percentage (56% at 10 μM) with astemizole; however, several compounds could still prevent the binding

**Table 1. Structures and inhibitory activities against  $I_{Kr}$  of compounds.**


Numbers	R	X	IC <sub>50</sub> /( $\mu$ M)	% Displacement at 10 $\mu$ M
Astemizole			0.0084	56.61 $\pm$ 4.29
Atropine				7.41 $\pm$ 3.53
WR1	F	N-phenyl	NA	29.08 $\pm$ 0.27
WR2	F	N-(2-methoxy)phenyl	NA	24.69 $\pm$ 0.27
WR3	F	O	NA	2.26 $\pm$ 0.89
WR4	F	C	NA	31.66 $\pm$ 1.29
WR5	F	N-methyl	NA	3.22 $\pm$ 1.27
WR6	F	N-Boc	NA	6.20 $\pm$ 2.36
WR7	Br	N-phenyl	NA	34.98 $\pm$ 2.33
WR8	Br	N-(2-methoxy)phenyl	9.16	25.79 $\pm$ 2.81
WR9	Br	O	NA	23.49 $\pm$ 0.06
WR10	Br	C	NA	17.95 $\pm$ 5.86
WR11	Br	N-methyl	NA	11.08 $\pm$ 9.16
WR12	Br	N-Boc	1.07	25.85 $\pm$ 2.88

NA: No significant dose response curve fitted.

of the ligand to the channel with > 20% replacement percentage when their concentrations are 10  $\mu$ M (Table 1). We then calculated the inhibitory IC<sub>50</sub> values for providing the exact value of inhibitory activity against  $I_{Kr}$ . As a result, two compounds with bromine rather than fluorine displayed potent inhibitory activities against  $I_{Kr}$ , which proposes that R group should be low electronegativity. While compare the X group we can find that phenyl and 2-methoxyphenyl are superior to other substituents to a piperazine ring, such as O, N-methyl, *etc.*, which suggests that phenylpiperazine is highly fitted with the binding pocket of hERG.

On the basis of the result of  $I_{Kr}$  inhibition assay, we harvested several compounds with  $I_{Kr}$  inhibitory activities. In our opinions, these compounds are a new series of  $I_{Kr}$  inhibitors with novel chemical structures, so that can serve as lead compounds for the development of new  $I_{Kr}$  inhibitors.

#### 4. Conclusion

In conclusion, a series of 4-chromanone derivatives were well designed and synthesized in the current study. After biological evaluation, two compounds exhibited moderate inhibition against  $I_{Kr}$ . It should be noted that these compounds are new  $I_{Kr}$  inhibitors with novel structure, so that can serve as lead compounds for further development. SAR analysis revealed that both the R group and the X group might play an important role in anti- $I_{Kr}$  activity, and substituents of a phenyl ring should have a high influence on the activity. The proof-of-concept in this study may provide essential information for the future design of inhibitors for  $I_{Kr}$ . In our follow-up study, we will continue to manipulate the

chemical structures of these compounds for improving the activity against  $I_{Kr}$ , as well as to take an attempt to employ other methods to test the inhibition of compound against  $I_{Kr}$  and to explore their possibility of preventing cardiac arrhythmia.

#### Acknowledgment

The present project was supported by grants from the National Natural Science Foundation of China (No. 30901836), the Doctoral Fund of Shandong Province BS2012YY008), the Shandong Natural Science Foundation (No. JQ201019), the Scientific Research Foundation for the Returned Overseas Chinese Scholars and the Independent Innovation Foundation of Shandong University, IIFSDU (No. 2009TB021 and 2012JC002).

#### References

- Ye D. Current strategies for the discovery of K<sup>+</sup> channel modulators. *Curr Top Med Chem.* 2009; 9:348-361.
- Hu B, Zhu XL, Fan QX, Li HX, & Zou CW. Experimental study on inhibition of rat ventricular Ik1 by RNA interference targeting the KCNJ2 gene. *Biosci Trends.* 2012; 6:26-32.
- Sanguinetti MC & Tristani-Firouzi M. hERG potassium channels and cardiac arrhythmia. *Nature.* 2006; 440:463-469.
- Vizzardi E. Efficacy of ranolazine in a patient with idiopathic dilated cardiomyopathy and electrical storm. *Drug Discov Ther.* 2013; 7:43-45.
- Lloyd J. Design and synthesis of 4-substituted benzamides as potent, selective, and orally bioavailable I(Ks) blockers. *J Med Chem.* 2001; 44:3764-3767.
- Sanguinetti M. A mechanistic link between an inherited

- and an acquired cardiac arrhythmia: HERG encodes the IKr potassium channel. *Cell*. 1995; 81:299-307.
7. Warmke JW, Ganetzky B. A family of potassium channel genes related to eag in *Drosophila* and mammals. *Proc Natl Acad Sci U S A*. 1994; 91:3438-3442.
  8. Du LP, Tsai KC, Li MY, You QD, Xia L. The pharmacophore hypotheses of I(Kr) potassium channel blockers: Novel class III antiarrhythmic agents. *Bioorg Med Chem Lett*. 2004; 14:4771-4777.
  9. Pardo L, Contreras-Jurado C, Zientkowska M, Alves F, Stühmer W. Role of voltage-gated potassium channels in cancer. *J Membr Biol*. 2005; 205:115-124.
  10. Witchel HJ. The hERG potassium channel as a therapeutic target. *Expert Opin Ther Targets*. 2007; 11:321-336.
  11. Du L, Li M, You Q, Xia L. A novel structure-based virtual screening model for the hERG channel blockers. *Biochem Biophys Res*. 2007; 355:889-894.
  12. Wulff H, Castle NA, Pardo LA. Voltage-gated potassium channels as therapeutic targets. *Nat Rev Drug Discov*. 2009; 8:982-1001.
  13. Zheng H. Design, synthesis, and evaluation of novel bifunctional iron-chelators as potential agents for neuroprotection in Alzheimer's, Parkinson's, and other neurodegenerative diseases. *Bioorg Med Chem*. 2005; 13:773-783.
  14. Goffin E. N-Aryl-N'-(chroman-4-yl)ureas and thioureas display *in vitro* anticancer activity and selectivity on apoptosis-resistant glioblastoma cells: screening, synthesis of simplified derivatives, and structure-activity relationship analysis. *Eur J Med Chem*. 2012; 54:834-844.
  15. Sarges R, Bordner J, Dominy BW, Peterson MJ, Whipple EB. Synthesis, absolute configuration and conformation of the aldose reductase inhibitor sorbinil. *J Med Chem*. 1985; 28:1716-1720.
  16. Patonay T, Vasas A, Kiss-Szikszai A, Silva AM, Cavaleiro JA. Efficient synthesis of chromones with alkenyl functionalities by the heck reaction. *Aust J Chem*. 2010; 63:1582-1593.
  17. Reddy M, Krupadanam G, Srimannarayana G. A Facile synthesis of 3,4-dihydro-1,5-benzodioxepin-2-ones. *Org Prep Proced Int*. 1989; 21:221-223.
  18. Huang K. Highly enantioselective borane reduction of heteroaryl and heterocyclic ketoxime ethers catalyzed by novel spiroborate ester derived from diphenylvalinol: Application to the synthesis of nicotine analogues. *J Org Chem*. 2008; 73:4017-4026.
  19. Baraldi PG. N6-[(Hetero)aryl]/(cyclo)alkyl-carbamoyl-methoxy-phenyl]-2-(chloro)-5'-N-ethylcarboxamido-adenosines: The first example of adenosine-related structures with potent agonist activity at the human A2B adenosine receptor. *Bioorg Med Chem*. 2007; 15:2514-2527.
  20. Li HJ. Mechanism of the intramolecular Claisen condensation reaction catalyzed by MenB, a crotonase superfamily member. *Biochemistry*. 2011; 50:9532-9544.

(Received March 11, 2014; Revised March 25, 2014; Accepted March 27, 2014)

## Appendix

### Chemistry: general procedures

All materials were purchased from commercial suppliers and used without further purification. Twice-distilled water was used throughout all experiments. Solvents were distilled prior to use, and all the reactions were monitored by thin-layer chromatography on 0.25 mm silica gel plates (60GF-254) and visualized with UV light or ninhydrin. Mass spectra were performed by the analytical and the mass spectrometry facilities in Drug Analysis Center at Shandong University on Agilent Technologies 1100 infinity HPLC, Applied Biosystems API4000. <sup>1</sup>H-NMR spectra were recorded on the Bruker 300 MHz NMR and 600 MHz NMR spectrometers. Chemical shifts are reported in delta ( $\delta$ ) units, parts per million (ppm) downfield from trimethylsilane. Melting points were determined on an electrothermal melting point apparatus (uncorrected).

#### 1.1. Phenyl-4-propynylpiperazine (7a)

Propargyl bromide (1.19 g, 10 mmol) was added slowly to a mixture of 1-phenylpiperazine (1.95 g, 12 mmol) and K<sub>2</sub>CO<sub>3</sub> (2.76 g, 20 mmol) in acetone. The mixture was stirred for 16 h at room temperature, filtered, and the precipitation was washed with acetone. The combined filtrate was evaporated under vacuum. Water was added to the residue, and the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> (40 mL). The combined organic layer was washed with 5% NaHCO<sub>3</sub> (3 × 50 mL), saturated brine (2 × 50 mL), and then dried over MgSO<sub>4</sub>. The solution was filtered and evaporated. The crude product was then purified by column chromatography to obtain a white solid, yield 80%, m.p: 46-48°C. ESI-MS calcd for C<sub>13</sub>H<sub>16</sub>N<sub>2</sub> (M + H<sup>+</sup>): 201.1; found: 201.4. <sup>1</sup>H-NMR (600 MHz, DMSO-d<sub>6</sub>):  $\delta$  = 7.19 (dd, *J*<sub>1</sub> = 9.0, 7.8 Hz, 2H), 6.92 (d, *J* = 7.8 Hz, 2H), 6.76 (t, *J* = 7.2 Hz, 1H), 3.36 (s, 2H), 3.22 (s, 1H), 3.15 (s, 4H), 2.62 (s, 4H).

#### 1.2. 1-(2-Methoxyphenyl)-4-propynylpiperazine (7b)

Compound 7b was synthesized following the procedure described in 1.1. White solid, yield 76%, m.p: 75-77°C. ESI-MS calcd for C<sub>14</sub>H<sub>18</sub>N<sub>2</sub>O (M + H<sup>+</sup>): 231.1; found: 231.2. <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.01 (m, 1H), 6.96 (d, *J* = 1.2 Hz, 1H), 6.92 (t, *J* = 7.2 Hz, 1H), 6.86 (d, *J* = 7.8 Hz, 1H), 3.87 (s, 3H), 3.36 (d, *J* = 1.8 Hz, 4H), 2.79 (s, 4H), 2.27 (s, 1H).

#### 1.3. N-propargylmorpholine (7c)

Compound 7c was synthesized following the procedure described in 1.1. White solid, yield 51%, m.p: 161-163°C. ESI-MS calcd for C<sub>7</sub>H<sub>11</sub>NO (M + H<sup>+</sup>): 126.1; found: 126.0. <sup>1</sup>H-NMR (600 MHz, D<sub>2</sub>O)  $\delta$ : 3.95-3.90 (m,

4H), 3.77 (s, 2H), 3.43 (s, 2H), 3.13 (s, 2H), 2.95 (t,  $J = 2.4$  Hz, 1H).

#### 1.4. 1-(Prop-2-ynyl)piperidine (7d)

Compound 7d was synthesized following the procedure described in 1.1. White solid, yield 46%, m.p: 178-180°C. ESI-MS calcd for  $C_8H_{13}N$  ( $M + H^+$ ): 124.1; found: 124.3.  $^1H$ -NMR (600 MHz,  $D_2O$ )  $\delta$ : 3.79 (s, 2H), 3.45-3.43 (m, 2H), 2.90-2.83 (m, 3H), 1.80-1.78 (m, 2H), 1.65-1.63 (m, 1H), 1.57-1.50 (m, 2H), 1.29-1.26 (m, 1H).

#### 1.5. 1-Methyl-4-propargylpiperazine (7e)

Compound 7e was synthesized following the procedure described in 1.1. White solid, yield 72%, m.p: 210°C. ESI-MS calcd for  $C_8H_{14}N_2$  ( $M + H^+$ ): 139.1; found: 139.1.  $^1H$ -NMR (600 MHz,  $D_2O$ )  $\delta$ : 3.96 (s, 2H), 3.69 (s, 4H), 3.35 (s, 4H), 2.98 (d,  $J = 2.4$  Hz, 1H), 2.85 (s, 3H).

#### 1.6. Tert-butyl 4-propargylpiperazine-1-carboxylate (7f) (13)

A solution of di-*tert*-butyl dicarbonate (4.34 g, 20 mmol) in  $CH_2Cl_2$  (25 mL) was slowly added to a stirring solution of piperazine (3.46 g, 40 mmol) in  $CH_2Cl_2$  (50 mL) at 0°C. The mixture was then stirred for 24 h at room temperature, and the solvent removed in vacuum. The crude solid was redissolved in diethyl ether (100 mL) with warming, and the white precipitate was filtered to. The product was extracted from the mother liquor with 1M citric acid solution ( $3 \times 50$  mL), and the aqueous layer was washed with  $CH_2Cl_2$  ( $3 \times 50$  mL), basified with  $Na_2CO_3$  (pH 11), and extracted with  $CH_2Cl_2$  ( $3 \times 50$  mL). The combined organic layer was dried over  $MgSO_4$  and evaporated in vacuum to give *tert*-butyl 1-piperazinecarboxylate as a waxy white solid (2.82 g, yield 75.6%). Propargyl bromide (2.38 g, 20 mmol) was added slowly to a mixture of *tert*-butyl 1-piperazinecarboxylate (4.4 mg, 24 mmol) and  $K_2CO_3$  (3.32 g, 24 mmol) in acetone. The mixture was stirred for 12 h at room temperature.  $CH_2Cl_2$  (40 mL) was then added, and the solution obtained was washed with 5%  $NaHCO_3$  ( $3 \times 50$  mL), saturated brine ( $2 \times 50$  mL), and then dried over  $MgSO_4$ . The solution was filtrated and evaporated to dryness. The residue was crystallized from ethanol and gave *tert*-butyl 4-propargylpiperazine-1-carboxylate 7f (4.2 g, yield 78%), m.p: 98-100°C. ESI-MS calcd for  $C_{12}H_{20}N_2O_2$  ( $M + H^+$ ): 224.1; found: 225.1.  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$ : 3.41(s, 4H), 3.26 (s, 2H), 2.46 (s, 4H), 2.22 (s, 1H), 1.42 (s, 9H).

#### 2.1. 3-(4-Fluorophenoxy)propanoic acid (1a) (14)

A mixture of potassium hydroxide (12.34 g, 220 mmol), 4-fluorophenol (11.21 g, 100 mmol), 3-chloropropionic

acid (10.85 g, 100 mmol) and ethanol (2 mL) in water (40 mL) were refluxed 6 h. After cooling, the solution was acidified with concentrated hydrochloric acid to pH = 2 and extracted with ethyl acetate. The organic layer was washed with saturated aqueous solution of sodium bicarbonate. The aqueous phase was then acidified with concentrated hydrochloric acid and extracted with ethyl acetate. The final organic layer was dried over magnesium sulfate, filtrated, and evaporated under vacuum. The title product was recrystallized in ethanol. The resulting precipitate was collected by filtration, washed with hexane, and dried (3.87g, yield 21%), m.p: 84-86°C (Lit (15) m.p: 84-86°C). ESI-MS calcd for  $C_9H_9FO_3$  ( $M + H^+$ ): 185.1; found: 185.3.

#### 2.2. 3-(4-Bromophenoxy)propanoic acid (1b)

Compound 1b was synthesized following the procedure described in 2.1. White solid, yield 24%, and m.p: 148-150°C (Lit (16) m.p: 145-147°C). ESI-MS calcd for  $C_9H_9BrO_3$  ( $M + H^+$ ): 243.9; found: 243.8.

#### 3.1. 6-Fluorochroman-4-one (2a)

A mixture of 3-(4-fluorophenoxy)propanoic acid (3.68 g, 20 mmol) and polyphosphoric acid (PPA) (100 g) was placed into a three-neck round-bottom flask. The mixture was heated and stirred with a blade stirrer at 60°C for 4 h. One hundred mL ice water was added to the mixture after the temperature of the mixture was equal to room temperature, and then the aqueous layer was extracted with ethyl acetate for three times. The combined organic layer was dried overnight with anhydrous magnesium sulphate, filtrated, and evaporated under vacuum. The title product was recrystallized in ethanol. The resulting precipitate was collected by filtration, washed with hexane, and dried (2.76 g, yield 83%), and m.p: 118-120°C (Lit (15) m.p: 113-116°C). ESI-MS calcd for  $C_9H_7FO_2$  ( $M + H^+$ ): 167.0; found: 167.0.

#### 3.2. 6-Bromochroman-4-one (2b)

Compound 2b was synthesized following the procedure described in 3.1. White solid, yield 78%, and m.p: 77-78°C (Lit (17) m.p: 77°C). ESI-MS calcd for  $C_9H_7BrO_2$  ( $M + H^+$ ): 227.0; found: 227.1.

#### 4.1. (E)-6-fluorochroman-4-one oxime (3a) (18)

To a suspension of hydroxylamine hydrochloride (1.39g, 20 mmol) in ethanol (50 mL), 6-fluorochroman-4-one (1.66g, 10 mmol) was added. Then, a solution of  $Na_2CO_3$  (1.05 g, 20 mmol) in water (10 mL) was added dropwise. The resulting solution was stirred overnight. Most of the ethanol was evaporated, and then 60 mL water was added. The aqueous phase was extracted

with ethyl acetate (3 × 50 mL), and the combined organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>. The solvents were removed under vacuum, and the residue was recrystallized. The resulting precipitate was collected by filtration, washed with hexane, and dried (1.54 g, yield 85%), m.p: 108-110°C. ESI-MS calcd for C<sub>9</sub>H<sub>8</sub>FNO<sub>2</sub> (M + H<sup>+</sup>): 182.1; found: 182.0. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ: 8.25 (s, 1H), 7.51 (dd, *J* = 9.3 Hz, *J* = 3 Hz, 1H), 7.01 (ddd, *J* = 10.8 Hz, *J* = 7.8 Hz, *J* = 3.0 Hz, 1H), 6.89 (dd, *J* = 9 Hz, *J* = 4.5 Hz, 1H), 4.25 (t, *J* = 6.3 Hz, 2H), 3.00 (t, *J* = 6.0 Hz, 2H).

#### 4.2. (E)-6-bromochroman-4-one oxime (3b)

Compound 3b was synthesized following the procedure described in 4.1. White solid, yield 91%, m.p: 139-141°C. ESI-MS calcd for C<sub>9</sub>H<sub>8</sub>BrNO<sub>2</sub> (M + H<sup>+</sup>): 241.9; found: 241.7. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ: 8.46 (s, 1H), 7.95 (d, *J* = 2.4 Hz, 1H), 7.36 (dd, *J* = 9.0 Hz, *J* = 2.4 Hz, 1H), 6.81 (d, *J* = 8.7 Hz, 1H), 4.26 (t, *J* = 6.3 Hz, 2H), 3.00 (t, *J* = 6.3 Hz, 2H).

#### 5.1. 2-Chloro-N-(6-fluorochroman-4-yl)acetamide (5a) (19,20)

To a suspension of (E)-6-fluorochroman-4-one oxime (1.81 g, 10 mmol) in CH<sub>3</sub>COOH (20 mL) was added the Zn (dust, 3.27 g, 50 mmol). The resulting solution was stirred 24 h. The Zn dust was filter off and washed by ethanol (20 mL) three times. The filtrate was collected and evaporated under vacuum. Then the product without purification was transferred to a mixture of K<sub>2</sub>CO<sub>3</sub> (2.76 g, 20 mmol) and CH<sub>2</sub>Cl<sub>2</sub> (30 mL), and chloroacetyl chloride (2.26 g, 20 mmol) was added in the mixture at room temperature. The reaction mixture was stirred for 4 h, subsequently slowly poured into 100 mL of ice water. The aqueous solution was extracted with CH<sub>2</sub>Cl<sub>2</sub> (40 mL × 3), the organic phase was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent was evaporated to furnish a solid residue which was purified by crystallization from ethanol. The resulting precipitate was collected by filtration, washed with hexane, and dried (1.27 g, yield 52%), m.p: 129-131°C. ESI-MS calcd for C<sub>11</sub>H<sub>11</sub>ClFNO<sub>2</sub> (M + H<sup>+</sup>): 244.1; found: 244.2. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ: 6.95-6.89 (m, 2H), 6.83-6.80 (m, 2H), 5.19 (dt, *J* = 13.8 Hz, *J* = 6.3 Hz, 1H), 4.29-4.22 (m, 1H), 4.19-4.15 (m, 1H), 4.12 (d, *J* = 2.4 Hz, 2H), 2.29-2.19 (m, 1H), 2.90-2.00 (m, 1H).

#### 5.2. 2-Chloro-N-(6-bromochroman-4-yl)acetamide (5b)

Compound 5b was synthesized following the procedure described in 5.1. White solid, yield 50%, m.p: 176-178°C. ESI-MS calcd for C<sub>11</sub>H<sub>11</sub>BrClNO<sub>2</sub> (M + H<sup>+</sup>): 304.0; found: 304.2. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ: 7.31-7.26 (m, 2H), 6.76 (d, *J* = 6.0 Hz, 2H), 5.18 (dt, *J* = 13.5 Hz, *J* = 6.3 Hz, 1H), 4.31-4.24 (m, 1H), 4.20-

4.15(m, 1H), 4.13 (d, *J* = 4.2 Hz, 2H), 2.29-2.18 (m, 1H), 2.10-2.00 (m, 1H).

#### 6.1. N-(7-fluorochroman-4-yl)-2-(4-((4-phenylpiperazin-1-yl)methyl)-1H-1,2,3-triazol-1-yl)acetamide (WR1)

Compound 5a (0.24 g, 1.0 mmol), KI (0.17 g, 1 mmol), sodium azide (2.2 mmol), 15 mL acetonitrile and 10 mL H<sub>2</sub>O were added to a round bottom flask. After stirring overnight at 80°C, the reaction mixture was cooled to room temperature and most of the acetonitrile was evaporated and then 60 mL water was added. The product (6a) was extracted by CH<sub>2</sub>Cl<sub>2</sub> and removed the solvent under reduced pressure. Without purification, 6a was dissolved into 20 mL methanol and 2 mL water, and compound 7a (0.2 g, 1.0 mmol), an aqueous sodium ascorbate solution (0.1 g, 0.5 mmol, dissolved in 2 mL water) and an aqueous solution of copper(II) sulfate pentahydrate (0.1 mL, 0.05 mmol) was added to this stirred solution. After another 24 h of stirring, the solvent was removed under vacuum, and the crude product was purified by flash chromatography (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 10:1) to afford the product WR1 (0.2 g, yield 52%), and m.p: 97-99°C. ESI-HRMS calcd for C<sub>24</sub>H<sub>27</sub>FN<sub>6</sub>O<sub>2</sub> (M + H<sup>+</sup>): 451.2258; found: 451.2277. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ: 7.75 (s, 1H), 7.03-6.98 (m, 2H), 6.93 (d, *J* = 4.2 Hz, 2H), 6.89-6.83 (m, 2H), 6.80-6.72 (m, 2H), 6.70 (d, *J* = 7.8 Hz, 1H), 5.17-5.11(m, 3H), 4.23-4.16 (m, 1H), 4.11-4.04 (m, 1H), 3.78 (s, 2H), 3.09 (br.s, 4H), 2.75 (br.s, 4H), 2.24-2.14 (m, 1H), 2.03-1.93 (m, 1H).

#### 6.2. N-(6-fluorochroman-4-yl)-2-(4-((4-(2-methoxyphenyl)piperazin-1-yl)methyl)-1H-1,2,3-triazol-1-yl)acetamide (WR2)

Compound WR2 was synthesized as white solid following the procedure described in 6.1, yield 41%, m.p: 152-154°C. ESI-HRMS calcd for C<sub>25</sub>H<sub>29</sub>FN<sub>6</sub>O<sub>3</sub> (M + H<sup>+</sup>): 481.2363; found: 481.2389. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ: 7.80 (s, 1H), 7.04-6.67 (m, 1H), 6.93 (d, *J* = 4.2 Hz, 2H), 6.90-6.82 (m, 2H), 6.80 (dt, *J* = 8.1 Hz, *J* = 3 Hz, 2H), 6.59 (d, *J* = 8.1 Hz, 1H) 5.18-5.12 (m, 3H), 4.23-4.16 (m, 1H), 4.11-4.04 (m, 1H), 3.86 (s, 3H), 3.82 (s, 2H), 3.12 (br.s, 4H), 2.78 (br.s, 4H), 2.25-2.14 (m, 1H), 2.04-1.93 (m, 1H).

#### 6.3. N-(6-fluorochroman-4-yl)-2-(4-(morpholinomethyl)-1H-1,2,3-triazol-1-yl)acetamide (WR3)

Compound WR3 was synthesized following the procedure described in 6.1. White solid, yield 38%, m.p: 152-154°C. ESI-HRMS calcd for C<sub>18</sub>H<sub>22</sub>FN<sub>5</sub>O<sub>3</sub> (M + H<sup>+</sup>): 376.1785; found: 376.1787. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ: 9.02 (d, *J* = 8.1 Hz, 1H), 8.37 (s, 1H), 7.07-7.00 (m, 2H), 6.85 (dd, *J* = 8.7 Hz, *J* = 4.8 Hz, 1H), 5.36 (dt, *J* = 25.2 Hz, *J* = 6.3 Hz, 2H), 5.04 (dt, *J* = 13.2 Hz,

$J = 6.3$  Hz, 1H), 4.49 (s, 2H), 4.26-4.15 (m, 2H), 3.98-3.95 (m, 2H), 3.77-3.70 (m, 2H), 3.38-3.30 (m, 2H), 3.11-3.02 (m, 2H), 2.11-2.03 (m, 1H), 1.96-1.86 (m, 1H).

6.4. *N*-(6-fluorochroman-4-yl)-2-(4-(piperidin-1-ylmethyl)-1*H*-1,2,3-triazol-1-yl)acetamide (WR4)

Compound WR4 was synthesized following the procedure described in 6.1. White solid, yield 46%, m.p.: 138-140°C. ESI-HRMS calcd for  $C_{19}H_{24}FN_5O_2$  ( $M + H^+$ ): 374.1992; found: 374.1994.  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$ : 8.90 (d,  $J = 7.8$  Hz, 1H), 7.96 (s, 1H), 7.07-7.00 (m, 2H), 6.85 (dd,  $J = 8.7$  Hz,  $J = 5.1$  Hz, 1H), 5.20 (dt,  $J = 26.7$  Hz,  $J = 16.2$  Hz, 2H), 5.04 (dt,  $J = 13.2$  Hz,  $J = 6.3$  Hz, 1H), 4.27-4.14 (m, 2H), 3.55 (s, 2H), 2.50 (br.s, 4H), 2.12-2.02 (m, 1H), 1.95-1.85 (m, 1H), 1.52-1.45 (m, 4H), 1.38-1.36 (m, 2H).

6.5. *N*-(6-fluorochroman-4-yl)-2-(4-((4-methylpiperazin-1-yl)methyl)-1*H*-1,2,3-triazol-1-yl)acetamide (WR5)

Compound WR5 was synthesized following the procedure described in 6.1. White solid, yield 39%, and m.p.: 223-225°C. ESI-HRMS calcd for  $C_{19}H_{25}FN_6O_2$  ( $M + H^+$ ): 389.2101; found: 389.2113.  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$ : 9.06 (d,  $J = 8.1$  Hz, 1H), 8.36 (s, 1H), 7.07-7.00 (m, 2H), 6.85 (dd,  $J = 8.7$  Hz,  $J = 4.8$  Hz, 1H), 5.33 (dt,  $J = 24.6$  Hz,  $J = 15.0$  Hz, 2H), 5.04 (dt,  $J = 13.5$  Hz,  $J = 6.3$  Hz, 1H), 4.48 (s, 2H), 4.28-4.17 (m, 2H), 3.80-3.77 (m, 4H), 3.62-3.58 (m, 4H), 2.81 (s, 3H), 2.13-2.03 (m, 1H), 1.97-1.87 (m, 1H).

6.6. *Tert*-butyl 4-((1-(2-((6-fluorochroman-4-yl)amino)-2-oxoethyl)-1*H*-1,2,3-triazol-4-yl)methyl)piperazine-1-carboxylate (WR6)

Compound WR6 was synthesized following the procedure described in 6.1. White solid, yield 44%, m.p.: 224-226°C. ESI-HRMS calcd for  $C_{23}H_{31}FN_6O_4$  ( $M + H^+$ ): 475.2469; found: 475.2473.  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$ : 9.06 (d,  $J = 7.8$  Hz, 1H), 8.38 (d,  $J = 4.2$  Hz, 1H), 7.07-7.00 (m, 2H), 6.85 (dd,  $J = 8.7$  Hz,  $J = 4.8$  Hz, 1H), 5.34 (dt,  $J = 26.4$  Hz,  $J = 15.9$  Hz, 2H), 5.04 (dt,  $J = 13.2$  Hz,  $J = 6.3$  Hz, 1H), 4.49 (s, 2H), 4.26-4.17 (m, 2H), 3.51-3.01 (m, 8H), 2.13-2.03 (m, 1H), 1.95-1.88 (m, 1H), 1.47 (s, 9H).

6.7. *N*-(6-bromochroman-4-yl)-2-(4-((4-phenylpiperazin-1-yl)methyl)-1*H*-1,2,3-triazol-1-yl)acetamide (WR7)

Compound WR7 was synthesized following the procedure described in 6.1. White solid, yield 40%, and m.p.: 148-150°C. ESI-HRMS calcd for  $C_{24}H_{27}BrN_6O_2$  ( $M + H^+$ ): 511.1457, 513.1437; found, 511.1448, 513.1430.  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$ : 8.89 (d,  $J = 8.1$  Hz, 1H), 8.01 (s, 1H), 7.36-7.32 (m, 2H), 7.27-7.17 (m, 2H), 6.94

(d,  $J = 8.1$  Hz, 2H), 6.80-6.74 (m, 2H), 5.23 (dt,  $J = 31.2$  Hz,  $J = 15.9$  Hz, 2H), 5.04 (dt,  $J = 12.9$  Hz,  $J = 5.7$  Hz, 1H), 4.30-4.15 (m, 2H), 3.64 (s, 2H), 3.14-3.11 (m, 4H), 2.58-2.55 (m, 4H), 2.13-2.03 (m, 1H), 1.98-1.88 (m, 1H).

6.8. *N*-(6-bromochroman-4-yl)-2-(4-((4-(2-methoxyphenyl)piperazin-1-yl)methyl)-1*H*-1,2,3-triazol-1-yl)acetamide (WR8)

Compound WR8 was synthesized following the procedure described in 6.1. White solid, yield 42%, m.p.: 106-108°C. ESI-HRMS calcd for  $C_{25}H_{29}BrN_6O_3$  ( $M + H^+$ ): 541.1485, 543.1464; found, 541.1540, 543.1527.  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$ : 8.89 (d,  $J = 8.1$  Hz, 1H), 8.03 (s, 1H), 7.35-7.32 (m, 2H), 6.94-6.85 (m, 4H), 6.82-6.77 (m, 1H), 5.23 (dt,  $J = 31.2$  Hz,  $J = 16.2$  Hz, 2H), 5.04 (dt,  $J = 13.5$  Hz,  $J = 6.3$  Hz, 1H), 4.26-4.16 (m, 2H), 3.76 (s, 3H), 3.64 (s, 2H), 2.95 (br.s, 4H), 2.56 (br.s, 4H), 2.10-2.03 (m, 1H), 1.95-1.88 (m, 1H).

6.9. *N*-(6-bromochroman-4-yl)-2-(4-(morpholinomethyl)-1*H*-1,2,3-triazol-1-yl)acetamide (WR9)

Compound WR9 was synthesized following the procedure described in 6.1. White solid, yield 39%, m.p.: 166-168°C. ESI-HRMS calcd for  $C_{18}H_{22}BrN_5O_3$  ( $M + H^+$ ): 436.0984, 438.0964; found, 436.0999, 438.0980.  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$ : 9.01 (d,  $J = 7.8$  Hz, 1H), 8.36 (s, 1H), 7.36-7.32 (m, 2H), 6.81-6.77 (m, 1H), 5.34 (dt,  $J = 31.5$  Hz,  $J = 16.2$  Hz, 2H), 5.04 (dt,  $J = 13.5$  Hz,  $J = 6.3$  Hz, 1H), 4.49 (s, 2H), 4.30-4.17 (m, 2H), 4.07-3.95 (m, 2H), 3.76-3.69 (m, 2H), 3.37-3.30 (m, 2H), 3.12 (s, 2H), 2.13-2.03 (m, 1H), 1.97-1.87 (m, 1H).

6.10. *N*-(6-bromochroman-4-yl)-2-(4-(piperidin-1-ylmethyl)-1*H*-1,2,3-triazol-1-yl)acetamide (WR10)

Compound WR10 was synthesized following the procedure described in 6.1. White solid, yield 43%, m.p.: 215-216°C. ESI-HRMS calcd for  $C_{19}H_{24}BrN_5O_2$  ( $M + H^+$ ): 434.1192, 436.1171; found, 434.1203, 436.1185.  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$ : 7.71 (s, 1H), 7.26-7.22 (m, 1H), 7.18 (d,  $J = 2.1$  Hz, 1H), 6.72-6.67 (m, 2H), 5.15-5.10 (m, 3H), 4.24-4.17 (m, 1H), 4.11-4.03 (m, 1H), 3.64 (s, 2H), 2.44 (br.s, 4H), 2.22-2.12 (m, 1H), 2.06-1.93 (m, 1H), 1.61-1.54 (m, 4H), 1.46-1.42 (m, 2H).

6.11. *N*-(6-bromochroman-4-yl)-2-(4-((4-methylpiperazin-1-yl)methyl)-1*H*-1,2,3-triazol-1-yl)acetamide (WR11)

Compound WR11 was synthesized following the procedure described in 6.1. White solid, yield 34%, m.p.: 202-204°C. ESI-HRMS calcd for  $C_{19}H_{25}BrN_6O_2$  ( $M + H^+$ ): 449.1301, 451.1280; found, 449.1293, 451.1270.

<sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ: 9.03 (d, *J* = 7.8 Hz, 1H), 8.32 (s, 1H), 7.36-7.32 (m, 2H), 6.81-6.78 (m, 1H), 5.34 (dt, *J* = 30.9 Hz, *J* = 16.2 Hz, 2H), 5.04 (dt, *J* = 13.5 Hz, *J* = 6.3 Hz, 1H), 4.65 (s, 2H), 4.24-4.17 (m, 2H), 3.60-3.40 (m, 4H), 3.30-3.04 (m, 4H), 2.81 (s, 3H), 2.16-1.99 (m, 1H), 1.97-1.87 (m, 1H).

6.12. *Tert-butyl 4-((1-(2-((6-bromochroman-4-yl)amino)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl)piperazine-1-carboxylate (WR12)*

Compound WR12 was synthesized following the procedure described in 6.1. White solid, yield 42%, m.p: 206-207°C. ESI-HRMS calcd for C<sub>23</sub>H<sub>31</sub>BrN<sub>6</sub>O<sub>4</sub> (M + H<sup>+</sup>): 535.1668, 537.1648; found, 535.1654, 537.1635. <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ: 9.58 (s, 1H), 9.03 (d, *J* = 8.4 Hz, 1H), 7.36-7.32 (m, 2H), 6.81-6.77 (m, 1H), 5.34 (dt, *J* = 31.2 Hz, *J* = 16.2 Hz, 2H), 5.04 (dt, *J* = 13.2 Hz, *J* = 6.0 Hz, 1H), 4.49 (s, 2H), 4.30-4.13 (m, 2H), 4.06-4.02 (m, 2H), 3.47-3.00 (m, 6H), 2.13-2.00 (m, 1H), 1.97-1.87 (m, 1H), 1.41 (s, 9H).