

Drug Discoveries & Therapeutics

Volume 6, Number 4 August, 2012



www.ddtjournal.com



ISSN: 1881-7831 Online ISSN: 1881-784X CODEN: DDTRBX Issues/Vear: 6 Language: English Publisher: IACMHR Co., Ltd.

Drug Discoveries & Therapeutics is one of a series of peer-reviewed journals of the International Research and Cooperation Association for Bio & Socio-Sciences Advancement (IRCA-BSSA) Group and is published bimonthly by the International Advancement Center for Medicine & Health Research Co., Ltd. (IACMHR Co., Ltd.) and supported by the IRCA-BSSA and Shandong University China-Japan Cooperation Center for Drug Discovery & Screening (SDU-DDSC).

Drug Discoveries & Therapeutics publishes contributions in all fields of pharmaceutical and therapeutic research such as medicinal chemistry, pharmacology, pharmaceutical analysis, pharmaceutics, pharmaceutical administration, and experimental and clinical studies of effects, mechanisms, or uses of various treatments. Studies in drug-related fields such as biology, biochemistry, physiology, microbiology, and immunology are also within the scope of this journal.

Drug Discoveries & Therapeutics publishes Original Articles, Brief Reports, Reviews, Policy Forum articles, Case Reports, News, and Letters on all aspects of the field of pharmaceutical research. All contributions should seek to promote international collaboration in pharmaceutical science.

Editorial Board

Editor-in-Chief:

Kazuhisa SEKIMIZU The University of Tokyo, Tokyo, Japan

Co-Editors-in-Chief:

Xishan HAO Tianjin Medical University, Tianjin, China Norihiro KOKUDO The University of Tokyo, Tokyo, Japan Hongxiang LOU Shandong University, Ji'nan, China Yun YEN City of Hope National Medical Center, Duarte, CA, USA

Chief Director & Executive Editor:

Wei TANG The University of Tokyo, Tokyo, Japan

Managing Editor:

Hiroshi HAMAMOTO The University of Tokyo, Tokyo, Japan Munehiro NAKATA Tokai University, Hiratsuka, Japan

Senior Editors:

Guanhua DU Chinese Academy of Medical Science and Peking Union Medical College, Beijing, China Xiao-Kang LI National Research Institute for Child Health and Development, Tokyo, Japan Masahiro MURAKAMI Osaka Ohtani University, Osaka, Japan Yutaka ORIHARA The University of Tokyo, Tokyo, Japan Tomofumi SANTA The University of Tokyo, Tokyo, Japan Wenfang XU Shandong University, Ji'nan, China

Web Editor:

Yu CHEN The University of Tokyo, Tokyo, Japan

Proofreaders:

Curtis BENTLEY Roswell, GA, USA Thomas R. LEBON Los Angeles, CA, USA

Editorial and Head Office:

Pearl City Koishikawa 603, 2-4-5 Kasuga, Bunkyo-ku, Tokyo 112-0003, Japan Tel.: +81-3-5840-9697 Fax: +81-3-5840-9698 E-mail: office@ddtjournal.com

Drug Discoveries & Therapeutics

Editorial and Head Office

Pearl City Koishikawa 603, 2-4-5 Kasuga, Bunkyo-ku, Tokyo 112-0003, Japan

Tel: +81-3-5840-9697, Fax: +81-3-5840-9698 E-mail: office@ddtjournal.com URL: www.ddtjournal.com

Editorial Board Members

Alex ALMASAN (Cleveland, OH) John K. BUOLAMWINI (Memphis, TN) Shousong CAO (Buffalo, NY) Jang-Yang CHANG (Tainan) Fen-Er CHEN (Shanghai) Zhe-Sheng CHEN (Queens, NY) Zilin CHEN (Wuhan, Hubei) Shaofeng DUAN (Lawrence, KS) Chandradhar DWIVEDI (Brookings, SD) Mohamed F. EL-MILIGI (6th of October City) Hao FANG (Ji'nan, Shandong) Marcus L. FORREST (Lawrence, KS) Takeshi FUKUSHIMA (Funabashi, Chiba) Harald HAMACHER (Tübingen, Baden-Württemberg) Kenji HAMASE (Fukuoka, Fukuoka) Xiaojiang HAO (Kunming, Yunnan) Kiyoshi HASEGAWA (Tokyo) Waseem HASSAN (Rio de Janeiro) Langchong HE (Xi'an, Shaanxi) Rodney J. Y. HO (Seattle, WA) Hsing-Pang HSIEH (Zhunan, Miaoli)

Yongzhou HU (Hangzhou, Zhejiang) Yu HUANG (Hong Kong) Hans E. JUNGINGER (Marburg, Hesse) Amrit B. KARMARKAR (Karad, Maharashra) Toshiaki KATADA (Tokyo) Gagan KAUSHAL (Charleston, WV) Ibrahim S. KHATTAB (Kuwait) Shiroh KISHIOKA (Wakayama, Wakayama) Robert Kam-Ming KO (Hong Kong) Nobuyuki KOBAYASHI (Nagasaki, Nagasaki) Toshiro KONISHI (Tokyo) Chun-Guang LI (Melbourne) Minyong LI (Ji'nan, Shandong) Jikai LIU (Kunming, Yunnan) Xinyong LIU (Ji'nan, Shandong) Yuxiu LIU (Nanjing, Jiangsu) Xingyuan MA (Shanghai) Ken-ichi MAFUNE (Tokyo) Sridhar MANI (Bronx, NY) Tohru MIZUSHIMA (Tokyo) Abdulla M. MOLOKHIA (Alexandria)

Yoshinobu NAKANISHI (Kanazawa, Ishikawa) Xiao-Ming OU (Jackson, MS) Weisan PAN (Shenyang, Liaoning) Rakesh P. PATEL (Mehsana, Gujarat) Shivanand P. PUTHLI (Mumbai, Maharashtra) Shafiqur RAHMAN (Brookings, SD) Adel SAKR (Cairo) Gary K. SCHWARTZ (New York, NY) Yuemao SHEN (Ji'nan, Shandong) Brahma N. SINGH (New York, NY) Tianqiang SONG (Tianjin) Sanjay K. SRIVASTAVA (Amarillo, TX) Hongbin SUN (Nanjing, Jiangsu) Chandan M. THOMAS (Bradenton, FL) Murat TURKOGLU (Istanbul) Fengshan WANG (Ji'nan, Shandong) Hui WANG (Shanghai) Quanxing WANG (Shanghai) Stephen G. WARD (Bath) Yuhong XU (Shanghai) Bing YAN (Ji'nan, Shandong)

Yasuko YOKOTA (Tokyo) Takako YOKOZAWA (Toyama, Toyama) Rongmin YU (Guangzhou, Guangdong) Guangxi ZHAI (Ji'nan, Shandong) Liangren ZHANG (Beijing) Lining ZHANG (Ji'nan, Shandong) Na ZHANG (Ji'nan, Shandong) Ruiwen ZHANG (Amarillo, TX) Xiu-Mei ZHANG (Ji'nan, Shandong) Yongxiang ZHANG (Beijing)

(As of August 2012)

Reviews

| 169 - 177 | Research progress in the radioprotective effect of superoxide dismutase. <i>Xiaojing Huang, Chunxia Song, Chuanqing Zhong, Fengshan Wang</i> |
|-----------|--|
| 178 - 193 | An overview on antiepileptic drugs. Nirupam Das, Meenakshi Dhanawat, Sushant K. Shrivastava |

Brief Report

| 194 - 197 | Anti-inflammatory activities of fractions from Geranium nepalense and | | |
|-----------|---|--|--|
| | related polyphenols. | | |
| | Chunhua Lu, Yaoyao Li, Liji Li, Lanying Liang, Yuemao Shen | | |

Original Articles

| 198 - 204 | Synthesis and structure-activity relationship study of 2-(substituted benzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2 <i>H</i> -thiazolo[3,2- <i>a</i>]pyrimidin-3(7 <i>H</i>)-one derivatives as anticancer agents. | | |
|-----------|---|--|--|
| | Theivendran Panneer Selvam, Viswanathen Karthick, Palanirajan Vijayaraj Kumar, Mohamed Ashraf Ali | | |
| 205 - 211 | Antioxidant and hepatoprotective activity of an ethanol extract of <i>Syzygium jambos</i> (L.) leaves. | | |
| | Md. Rafikul Islam, Mst. Shahnaj Parvin, Md. Ekramul Islam | | |
| 212 - 217 | Antimicrobial screening of some Egyptian plants and active flavones from <i>Lagerstroemia indica</i> leaves. | | |
| | Yasser Diab, Khaled Atalla, Khaled Elbanna | | |
| 218 - 225 | Optimization of cell wall skeleton derived from Mycobacterium bovis | | |
| | BCG Tokyo 172 (SMP-105) emulsion in delayed-type hypersensitivity and antitumor models. | | |
| | Masanori Miyauchi, Masashi Murata, Akihisa Fukushima, Toshiyuki Sato, | | |
| | Masae Nakagawa, Tetsuya Fujii, Norimasa Koseki, Nobuyuki Chiba, Yasuo Kashiwazaki | | |

Commentary

| 226 - 229 | Animal welfare and use of silkworm as a model animal. | | |
|-----------|---|--|--|
| | Nobukazu Sekimizu, Atmika Paudel, Hiroshi Hamamoto | | |

Guide for Authors

Copyright

Review

Research progress in the radioprotective effect of superoxide dismutase

Xiaojing Huang^{1,2}, Chunxia Song^{1,2,3}, Chuanqing Zhong^{1,2,4}, Fengshan Wang^{1,2,*}

¹ Key Laboratory of Chemical Biology of Natural Products (Ministry of Education), Ji'nan, Shandong, China;

² Institute of Biochemical and Biotechnological Drugs, School of Pharmaceutical Sciences, Shandong University, Ji'nan, Shandong China;

³ Marine College, Shandong University at Weihai, Weihai, Shandong, China;

⁴ Municipal and Environmental Engineering College, Shandong Jianzhu University, Ji'nan, Shandong, China.

ABSTRACT: Irradiation from diverse sources is ubiquitous and closely associated with human activity. Radiation therapy (RT), an important component of the multiple radiation origins, contributes significantly to oncotherapy by killing tumor cells. On the other hand, RT can also cause some undesired normal tissue injuries that afflict numerous cancer patients. Although many promising radioprotective agents are emerging, few of them have entered the market successfully due to various limitations. At present, the most accepted hypothesis for the radiation-caused injury involves reactive oxygen species (ROS) generation. Superoxide dismutase (SOD), the unique enzyme responsible for the dismutation of superoxide radicals, is expected to occupy an indispensable position in the treatment of ROS-mediated tissue injuries originating from exposure to radiation. This review focuses on the mechanism of radioprotection by SOD at the tissue or organ level, cellular level, and molecular level, respectively, in order to provide references for further investigation of radiation injury and development of new radioprotectors.

Keywords: Superoxide dismutase, radioprotection, radiation, reactive oxygen species

1. Introduction

Irradiation from diverse sources is ubiquitous and closely associated with human activity. As shown in Figure 1 (1), among all sources of radiation, natural radiation, including radon, thoron, cosmic radiation, and natural

*Address correspondence to:

radioactivity in soils and food, dominates in the average doses of individual radiation adsorption. However, the dose from natural radiation is not under human control. By contrast, artificial radiation, which consists primarily of medical exposure of patients, accounts for approximately 14 percent of the individual radiation absorption and has been attracting more and more attention over recent years. Moderate application of man-made radiation is extremely beneficial as shown by the excellent performance of radiation therapy in oncology (2). However, some adverse effects accompany the therapeutic benefit as a consequence of the unavoidable exposure of the surrounding normal tissues to radiation. In this sense, besides the skin being irradiated directly, other internal radiosensitive organs also cannot escape from being injured to different degrees (3), which become an impediment, counteracting the efficacy of radiation therapy (RT). In order to minimize these undesired side effects, many efforts have been made to improve the RT technology, such as image-guided radiotherapy, proton radiotherapy, and intensity-modulated radiotherapy. Even though these advanced techniques have the advantage of improved accuracy and control of irradiation, the patients still confront the potential risk of normal tissue injuries (2). Thus, to seek the radiation modifiers with selective protection for normal tissues has been a realm of intense investigation.

Despite the fact that many promising radioprotective candidates are emerging, amifostine (WR2721) is the only one approved for clinical use to date. In addition to its high efficacy in ameliorating xerostomia resulting from irradiation (4,5), high frequencies of deleterious

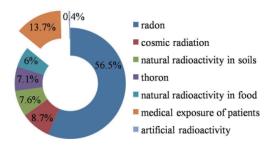


Figure 1. Dose contribution to the individual radiation absorption from all sources of radiation.

Dr. Fengshan Wang, Key Laboratory of Chemical Biology of Natural Products (Ministry of Education), Institute of Biochemical and Biotechnological Drugs, School of Pharmaceutical Sciences, Shandong University, 44 Wenhua Road West, Ji'nan 250012, Shandong, China. E-mail: fswang@sdu.edu.cn

side effects (nausea, cutaneous reactions, hypotension, *etc.*) and even tumor radioprotection have been reported, which limit its use (*6-9*). Therefore, the search for other radioprotectors with high potency and low toxicity should be the primary subject of further research.

At present, various compounds are being evaluated and examined for their radioprotective properties. It has been reported that ReIB, one of the five NF-kB family members existing in mammals, can improve the radioresistance of prostate cancer cells through up-regulation of the mitochondria-localized manganese-superoxide dismutase (Mn-SOD) expression (10). Meanwhile, Murley et al. have carried out a series of research on the delayed radioprotection for RKO36 cells (a strain of human colon carcinoma cells). They found that preincubation of RKO36 cells with WR1065 (the free thiol form of amifostine) or tumor necrosis factor alpha (TNF- α) could effectively stimulate the expression of Mn-SOD, thereby enhancing the adaptive response of the cells to the subsequent radiation challenge (11, 12). Additionally, in the study by Zhang et al., CpGoligodeoxynucleotide (CpG-ODN) was shown to effectively relieve bone marrow hemopoiesis radiation injury. Interestingly, the mechanism by which CpG-ODN acted was also through activating the NF-kB pathway and elevating Mn-SOD content (13).

These several series of evidence indicate that SOD plays a significant role in radioprotection, and it is of great importance to investigate further details of its mechanism of action so as to develop it as a radioprotector. This review describes the radioprotective studies of SOD based on the hypothesis of reactive oxygen species (ROS) generation associated with radiation injury.

2. Reactive oxygen species and radiation injury

Reactive oxygen species (ROS), *in vivo* byproducts of oxygen metabolism, comprise a multitude of family members such as superoxide radical (O_2^{-}), hydrogen peroxide (H₂O₂), singlet oxygen (¹O₂), hydroxyl radical (OH) and so on (14). Though the physiological levels of ROS are critical for a variety of cellular functions, such as cell growth, stress adaption, injury response, and cellular phenotype development (15), inevitable toxicities can be induced by overdoses of ROS under some pathological conditions (16). Homeostasis of ROS is not only affected by endogenous factors but also by exogenous ones (Table 1).

| Table 1. Summary of diverse sources of ROS | Table 1. | Summary | of diverse | sources | of ROS |
|--|----------|---------|------------|---------|--------|
|--|----------|---------|------------|---------|--------|

| Endogenous sources | Exogenous sources |
|--|---|
| NAD(P)H oxidase Xanthine oxidase P-450 monooxygenases Lipoxygenases Cyclooxygenases, <i>etc.</i> | Radiations Pathogens Metals: Fe, Cu, Zn Xenobiotics, <i>etc.</i> |

Note: This table summarizes the diverse sources of ROS based on the authors' interpretation of the references (14) and (17). Readers can refer to the web version of these articles.

Ionizing radiation (IR), possessing great strength of penetration, usually exerts its harmful effects on organisms and biomolecules through both direct and indirect effects. The former is referred to the irreversible injuries caused by radiation selectively impacting certain biomolecules, of which DNA damage is the most notable one. The latter is associated with the condition, in which radiation interacts with non-targeted molecules, induces abnormal levels of ROS exceeding the capacity of the organism to clear them, and, consequently, leads to oxidative stress-mediated damaging effects (2). Owing to the highly oxidative activity, •OH, the radiolysis product of water, contributes a lot to the adverse reactions immediately after radiation by breaking chemical bonds and promoting lipid peroxidation. This explains the significant role of water in the indirect effects of IR (18). Moreover, it has been demonstrated that the radiationexposed organisms perpetuate elevated levels of ROS (19-21), caused to a large extent by the mitochondrial dysfunction. Under such condition, molecular oxygen (O_2) is partly reduced to generate considerable amounts of $O_2^{\overline{2}}$ and H₂O₂ (22). Then, by the way of Fenton and Haber-Weiss reactions, respectively, both $O_2^{\overline{2}}$ and H_2O_2 can be further converted to \cdot OH (23), which is the most toxic of all ROS responsible for the majority of IR-mediated adverse reactions. Depending on the photochemical reaction between radiation and some endogenous photosensitizers localized in the cellular or mitochondrial membrane, such as nicotinamide adenine dinucleotide phosphate (NADPH) oxidase and xanthine oxidase (24), huge doses of ROS are generated, overwhelming the antioxidant defense system in organisms, which result in a series of serious adverse effects.

Depending upon the level of injury, the IR-induced damages are usually categorized as follows: tissue or organ level effects, cellular level effects and molecular level effects (see Figure 2).

3. Superoxide dismutase and its radioprotective effects

Approximately eighty years ago, Keilin *et al.* isolated a blue protein containing copper from bovine erythrocytes for the first time and named it ergthrocuprein. However, they had no idea about its bioactivity. Not until 1969 did

| Tissue or organic level | Cellular level | Molecular level |
|-------------------------|-------------------|--------------------|
| Skin injury | G1 arrest | Lipid peroxidation |
| Myelotoxicity | S delay | Mitochondrial |
| Pneumonitis | G2 accumulation | membrane |
| Xerostomia | Cell death | depolarization |
| Esophagitis | etc | DNA damages |
| etc | | etc |

Figure 2. IR-induced damaging effects at different levels.

McCord and Fridorich discover that this protein has the enzymatic activity of catalyzing $O_2^{\frac{1}{2}}$ dismutation and formally denominate it superoxide dismutase (SOD) (25), which is regarded as a thumping breakthrough that triggered extensive research on SOD from then on.

As naturally present antioxidant enzymes, SODs exist in mammals in diverse forms. There are Cu, Zn-SOD in cytoplasm and nucleus, Mn-SOD oriented in mitochondria, and EC-SOD present mainly in extracellular spaces (25-27). Their three-dimensional structures are depicted in Figure 3.

All of the SODs can accelerate the dismutation of O_2^{-1} with a powerful potency to yield O_2 and H_2O_2 ; the latter is further decomposed into nontoxic products by

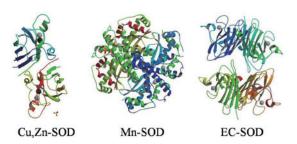


Figure 3. Structures of different SODs in mammals from the Protein Data Bank (PDB). A, Human Cu, Zn-SOD (PDB ID 1PU0); B, Human Mn-SOD (PDB ID 1EM1); C, Human EC-SOD (PDB ID 2JLP). Human Cu, Zn-SOD is a homodimer with a molecular weight of 32 kDa. Each monomer has one Cu and Zn acting as their active center. Containing the same metal ions as Cu, Zn-SOD, human EC-SOD is a homotetramer with a molecular weight of approximately 135 kDa. Human Mn-SOD is also a homotetramer with molecular weights ranging from 86 to 88 kDa. Distinctively, Mn is essential for its enzymatic activity.

catalase (CAT) or glutathione peroxidase (GPX). Besides the conventional enzymes mentioned above, some other catalysts such as thioredoxins (TRXs) and glutaredoxins (GRXs) can also facilitate the degradation of H_2O_2 via modulating the redox balance of disulfides (see Figure 4) (28). Hence, a naturally occurring antioxidant enzymatic defense system is established in the organism involving SOD, CAT, GPX, TRXs, GRXs, and so on.

With the growing interest in SOD, it has become clear that SOD acts fundamentally for defeating ROSmediated diseases such as carcinoma, inflammation and aging (29,30). However, these diseases are beyond the scope of this review. Herein, we focus on the protective effects of SOD against IR-induced normal tissue injury at the tissue or organ level, cellular level, and molecular level, respectively.

3.1. Radioprotection at the tissue or organ level

Generally, large doses of radiation can lead to a remarkable reduction of parenchymal cells and development of tissue fibrosis, while the vessel wall of mesenchyma becomes thickened and microcirculation gets blocked. All of these together may provoke the burst of organ dysfunction eventually, if not repaired promptly (31). However, due to the dissimilar radiosensitivity, different tissues or organs show marked diversity in their responses to the radiation.

Skin, the firstly affected target of radiation, is one of the most acknowledged tissues with respect to the IRinduced injury. During the early period following the

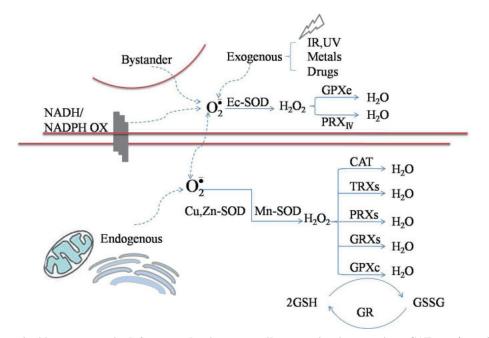


Figure 4. The antioxidant enzymatic defense mechanism naturally occurring in organism. CAT: catalase; GPXc: classic (intracellular) glutathione peroxidase; GPXe: extracellular glutathione peroxidase; GSH: reduced glutathione; GSSG: oxidized glutathione; GR: glutathione reductase; GRXs: glutaredoxins; PRXs: peroxiredoxins (thioredoxin peroxidase); PRX_{IV}: peroxiredoxin IV; TRXs: thioredoxins; NADH/NADPH OX: nicotinamide adenine dinucleotide reduced/nicotinamide adenine dinucleotide phosphate reduced oxidases; IR: ionizing radiation; UV: ultraviolet.

radiation, it is common to observe the appearance of local hyperemia and erythema on skin surface. Thereafter, owing to the long-term, persistent and epigenetic effects of radiation, the lesion may evolve into a chronic skin injury which has no efficient treatment (31). In the study by Yan et al., after a single dose of 37 Gy was given to mice on their right hind legs, AAV2-Mn-SOD-hrGFP, a recombinant adeno-associated virus vector expressing Mn-SOD tagged with humanized recombinant green fluorescent protein, was injected subcutaneously in the experimental group and an equal volume of AAV2-IRES (internal ribosome entry site)-hrGFP in the control group. Although all the irradiated mice demonstrated severe skin injury initially, the mice in the experimental group displayed pronounced mitigation and accelerated healing process (p < 0.05) two weeks after radiation compared with the control group. These results indicate the marked relief of IR-induced skin injury provided by Mn-SOD expression (32).

Because of the high radiosensitivity, it is obvious that hematopoietic cell proliferation can be suppressed upon the exposure to radiation (33). Numerous studies have been performed to explore the role of SOD in the adaptive response of the hematopoietic organ to the radiation. It was observed that intravenous injection of bovine SOD to mice could significantly promote the recovery of erythrocytes, reticulocytes and white blood cells from X-irradiation-induced loss (34,35). Moreover, Eastgate *et al.* verified that interleukin-1 (IL-1) administration was able to provide radioprotection of the irradiated mice from myelotoxicity as well as the explanted murine bone marrow cells from the damaging effects of IR mainly due to the effect of increased Mn-SOD expression triggered by IL-1 treatment (36).

Lung, another organ vulnerable to radiation damage, is subjected to injury regularly during the radiotherapy of patients with lung carcinoma or other thoracic cancers. Depending on the time of onset of lung injury as well as the pathological features exhibited, there are two kinds of IR-induced lung injury: acute radiation pneumonitis manifesting exudative inflammation and interstitial edema, and lung fibrosis characterized by the thickening of alveolar wall due to long-time exposure to radiation (37). However, these two types of lung injury are thought to be independent (38). Provided that lung fibrosis arising from radiation was a result of a cascade of cytokines among which TGF-B1 was a critical one (39), Machtay et al. used RT-PCR to detect the level of TGF-B1 expression in mouse lung homogenate and demonstrated that the group of mice treated with PEG-AOE (PEG-antioxidant enzyme composed of PEG-SOD and PEG-CAT at the ratio of 1:1) showed a notably diminished level of TGF-B1 compared with the group that received irradiation alone. In addition, further analysis indicated a remarkable reduction in the lung hydroxyproline content by PEG-AOE administration (40). The data above supports the concept that PEG-AOE has a great potential to reverse radiation-induced lung fibrosis.

For the head-and-neck cancer patients, local radiotherapy always results in xerostomia and oral mucositis (41), which bring them serious discomfort. By means of flow cytometry, it was found that the sharp reduction of salivary secretion in the mice irradiated at head-and-neck region was caused by the overproduction of ROS resulting from radiation. However, this phenomenon was not observed in the group of mice with PC-SOD (lecithinized SOD) treatment due to the capability of PC-SOD to scavenge $O_2^{\frac{1}{2}}$ during the whole experimental process (42). Subsequently, another study by Nagler et al. showed a similar trend in the Wistar rats administered with Mn-SOD, which showed a dramatic resistance against hyposalivation induced by local headand-neck radiation (43). Collectively, these findings suggest that SOD can effectively protect the saliva gland from radiation injury and neutralize IR-mediated hyposalivation.

Esophagitis is a major complication developed in the non-small cell lung cancer patients receiving RT. Stickle *et al.* found that intraesophageal injection of Mn-SOD-PL (plasmid liposome) prior to radiation could significantly prevent the development of vacuole in the esophageal squamous lining cells (p < 0.001) and elevate the mouse survival (p = 0.0009) suggesting the protective effect provided by Mn-SOD-PL-mediated SOD expression against esophagitis (44).

3.2. Radioprotection at the cellular level

It is well accepted that cell cycle is susceptible to radiation which can induce G1 arrest, S delay, and G2 accumulation (45). Nonetheless, attributable to the long-time evolution, the cell has developed a series of cell cycle checkpoints including G1/S checkpoint, S checkpoint, and G2/M checkpoint. All the checkpoints above collaborate to initiate related repair mechanisms and to guarantee the normal transition from one phase to another depending upon the activity of cell phase-specific cyclins and cyclin-dependent protein kinases (CDKs). A representative profile for the regulation of mammalian cell cycle is described in Figure 5 (46).

During S phase, the cell needs to absorb appropriate amounts of ribonucleosides to maintain nucleic acid synthesis. In the study by Epperly *et al.*, 5-bromo-2deoxyuridine (BuDR) was given to the pre-irradiated mice by intraperitoneal injection and the intake of BuDR by oral cavity mucosal cells was measured one hour later to estimate the state of S phase. It was found that the groups of mice irradiated or irradiated with WR2721 administration alone showed BuDR intake multiple times higher than the normal control. However, this phenomenon was suppressed significantly in the mice treated with Mn-SOD-PL alone or Mn-SOD-PL combined with WR2721, which verified the hypothesis that SOD could suppress the radiation-induced G1 arrest to exert its protective effects (*47*).

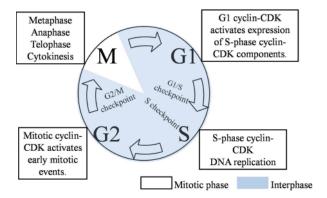


Figure 5. Regulation of mammalian cell cycle. G1: 1st gap; S: DNA synthesis phase; G2: 2nd gap; M: mitosis and cytokinesis. As shown in this figure, cell cycle consists of mitotic phases and interphases which are further classified into G1, S, and G2. G1 is also called presynthetic phase during which DNA pre-replication complexes accumulate and prepare for DNA synthesis. S is regarded as the core of interphase. In this phase, DNA is replicated adequately to enable the final cell division. G2 is referred to post-synthetic phase, during which DNA replication is completed and some early mitotic events are activated to be ready for the entry to M phase. M comprises metaphase, anaphase, telophase and cytokinesis. After separation of both chromosomes and cytoplasm, two new daughter cells carrying the same genetic material as that of the parent cell are obtained. Obviously, the ordered transition among different phases cannot do without the assistance of corresponding cyclins specific for every phase. It is these cyclins that act as the key regulatory factor in respective checkpoint.

G2/M checkpoint is devoted to prohibit the cell with damaged DNA from entering M phase directly. In particular, compound Cdc2(Chk1)-CyclinB involved in the transition from G2 to M is inactivated either by ATM-Chk2-Cdc25 or ATR-Chk1-Cdc25 pathway (48). Gao et al. showed that radiation induced considerable DNA damage and apoptosis as measured by the large increase of the percentage of cells with sub-G1 content. Yet, this outcome was not observed in the irradiated cells overexpressing Cu, Zn-SOD, where the protein CyclinB1 content decreased by 60-70% compared with the control and the percentage of cells accumulated in G2 phase increased significantly. The results demonstrate that Cu, Zn-SOD is able to provide radioprotection by the way of down-regulating CyclinB1 activity, retarding the G2/M transition and promoting DNA repair (46). Additionally, in accordance with the finding above, Kalen et al. obtained a similar conclusion that Mn-SOD could efficiently initiate the G2/M checkpoint to produce cytoprotection (49).

On the other hand, SOD is envisioned to perform its cellular level radioprotection through suppressing the abnormal proliferation. It is well accepted that stem cells usually multiply slowly to make the exact response to the signal from the external environment and determine whether to proliferate directly or differentiate for the sake of reducing DNA mutation and preventing tumorigenesis (*50*). Take the mouse esophageal side population (SP) stem cells as an example: samples from the mice irradiated alone showed a significant increase in the content of PCNA (proliferating cell nuclear antigen) compared to the

background (p < 0.0001), while in contrast, the esophageal SP stem cells from mice given Mn-SOD-PL kept the PCNA level commensurate to the normal control without any perturbation of multi-direction differentiation capacity, which confirms the theory described above regarding the action of SOD (51).

Clearly, the cellular level radioprotection provided by SOD can be ascribed to its capacity of either regulating the cell cycle checkpoint or inhibiting the abnormal proliferation of the cell. Therefore, it is concluded that SOD is able to act as a promising radioprotector to maintain the steady-state of the cell proliferation and restrict the inclination to carcinogenesis.

3.3. Radioprotection at the molecular level

As described previously, based on the interaction between radiation and *in vivo* biomolecules, excessive ROS are generated, which lead to the various pathological symptoms, implicating the significance of the molecular level radioprotection conducted by SOD.

Through direct interaction with radiation, lipids, the major constituents included in the construction of biomembrane, can be oxidized to produce considerable peroxidized lipids and malonaldehyde (MDA), which pose a threat to the integrity of the membrane structure (52). Early in 1976, a related study was conducted to determine the role of SOD involved in preserving the phospholipid biomembrane in vitro when it was exposed to radiation. The data revealed a notable increase in the amount of peroxidized lipids in the irradiated biomembrane as evidenced by the increased absorbance at 232 nm. Adversely, the same phenomenon was not observed in the biomembrane pre-incubated with bovine SOD at an extremely low concentration of 1 ng/mL, demonstrating the striking protection effect of SOD on biomembrane in vitro against lipid peroxidization caused by radiation (53). Recently, Epperly et al. used the irradiated mouse model transfected with Mn-SOD-PL to verify the hypothesis that the in vivo lipid peroxidization was partly regulated by the cytokines involved in the cell division to substitute the injured cells. By means of RNase protection assay, a detectable up-regulation of cytokines such as IFNy and TNFy was observed in the irradiated control mice but not in the mice with Mn-SOD-PL administration. Consistent with that, the latter mice also showed relatively lower level of peroxidized lipids after receiving radiation as compared with the irradiated control value, which confirmed their hypothesis successfully (54).

In addition, it is also worthwhile to mention the SODinitiated radioprotective effect on mitochondria, the essential organelle in which membrane depolarization is responsible for numerous detrimental reactions such as the release of cytochrome c, the activation of caspase, the uncoupling of oxidative phosphorylation (55). Over the past years, it has been proved that overexpression of Mn-SOD in irradiated cells significantly decreased the occurrence of mitochondrial membrane depolarization which was commonly seen in the control group (56). Besides, when Gorman *et al.* investigated the bystander effect of radiation, a remarkable genomic instability coupled with mitochondrial membrane depolarization was observed in the bystander cells. However, after transfection with Mn-SOD-PL, the biological reactions above were significantly inhibited. These results demonstrate the efficient radioprotection by SOD on mitochondria (57).

Similarly, DNA also seems to be the target biomolecule of radiation. It is well established that after radiation, a broad range of DNA damages are induced such as base damage, single strand breaks (SSBs), and double strand breaks (DSBs) if initial damage on DNA is not repaired properly. Among them, DSBs is regarded as the most deleterious one for its ability to arouse homologous recombination (HR) (58-61). Then, HR can further generate base insertion, depletion, translocation along with high carcinogenicity (59-61). Peroxynitrite, the product of the reaction between nitrogen monoxidum and superoxide, is able to trigger the formation of genomic rearrangement directly, indicating the key role of ROS scavenging in blocking cancer generation (see Figure 6). In the study by Niu *et al.*, fluorescent yellow direct repeat (FYDR) mice were employed to estimate the incidence of HR in vivo measured by the number of fluorescent recombinant cells using flow cytometry. The data showed a significant increase in fluorescent recombinant esophageal cells in the irradiated FYDR mice compared to the control, which indicated that a large degree of HR was stimulated by radiation. Whereas, in the irradiated FYDR mice with Mn-SOD-PL administration, the level of fluorescent recombinant cell counts was almost near the normal value (62). This study provides a powerful evidence for the potential of SOD to prevent HR induced by radiation. Furthermore, using agarose gel electrophoresis, Liu et al. found that the radiation-induced plasmid DNA damages, such as the increased amount of open circular, could be effectively suppressed by the pre-incubation of Hep-SOD (heparin-SOD conjugate) in vitro (63).

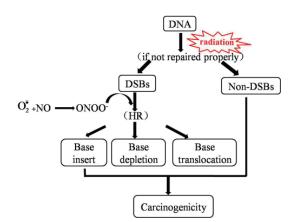


Figure 6. DNA damages triggered by radiation. HR: homologous recombination; DSBs: double strand breaks; Non-DSBs: single strand breaks, translocation, base damage, *etc*.

4. Conclusion

As the unique enzyme capable of dismutating $O_2^{\frac{1}{2}}$, SOD is expected to occupy an indispensable position in the treatment of ROS-mediated normal tissue injuries originating from exposure to radiation. Although SODrelated antiradiation research has been continued for nearly forty years and many positive outcomes have been obtained, hardly any drug based on SOD has been approved for radioprotective use in the clinic. Up to now, Orgotein is the only SOD product used as a radioprotector mainly in animals because of its inclination to induce allergic reaction in human (64). Other factors restricting its entry into clinical treatment include its large molecular weight, inability to pass the cell membrane freely, short half-life (65), rapid metabolic rate, narrow time-window of action (66) and so on. To solve these problems, scientific community has resorted to the investigation of SOD-based gene therapy, SOD conjugates, and non-enzymatic SOD mimics.

SOD-based gene therapy mediated by the plasmid liposome or recombinant virus vector showed positive outcomes in numerous research studies as described in this paper. This strategy addresses the poor membrane permeability and low expression of native SOD found in organisms. Additional work needs to be done to explore the availability of SOD-based gene therapy in human subjects. SOD conjugates obtained through the way of chemical modification have advantages of prolonged halflife, improved cell membrane permeability, augmented bioactivities and efficient targeting compared with the native SOD. An outstanding example of these compounds is Hep-SOD which has been verified for its superior radioprotection owing to its long half-life, and enhanced tolerance to high temperature, strong acid/base, and enzymolysis of trypsin (67,68). Besides these advanced forms of SOD, nonenzymatic SOD mimics have also become a favorite form of radioprotective agent to some researchers in recent years. This class of synthetic low molecular-weight compounds containing a metal ion as the active center also shows prolonged half-lives and widened time-windows compared to native SOD. Among them, M40403 (a manganese (II) complex with a bis (cyclo-hexylpyridine) substituted macrocyclic ligand) has been approved by FDA as a radioprotector for cancer patients (69).

From the discussion above, we firmly believe the great potential of SOD-based compounds to be developed as novel radioprotectors in the future. At present, the most important task is to continue studying further their pharmacokinetics, toxicity, optimal route of administration and to strive for their radioprotective application in the clinic as soon as possible.

Acknowledgements

This project was supported by the Important National Science & Technology Specific Projects of China (Grant No. 2010ZX09401-302-2-30).

References

- Radiological Protection Institute of Ireland. Your radiation exposure: Sources of radiation. *http://www.rpii. ie/Your-Health/ Your-radiation-exposure.aspx* (accessed October 16, 2011).
- Citrin D, Cotrim AP, Hyodo F, Baum BJ, Krishna MC, Mitchell JB. Radioprotectors and mitigators of radiationinduced normal tissue injury. Oncologist. 2010; 15:360-371.
- Christodouleas JP, Forrest RD, Ainsley CG, Tochner Z, Hahn SM, Glatstein E. Short-term and long-term health risk of nuclear-power-plant accidents. N Engl J Med. 2011; 364:2334-2341.
- Brown DQ, Pittock JW 3rd, Rubinstein JS. Early results of the screening program for radioprotectors. Int J Radiat Oncol Biol Phys. 1982; 8:565-570.
- Cassatt DR, Fazenbaker CA, Bachy CM, Hanson MS. Preclinical modeling of improved amifostine (Ethyol) use in radiation therapy. Semin Radiat Oncol. 2002; 12:97-102.
- Koukourakis MI, Kyrias G, Kakolyris S, Kouroussis C, Frangiadaki C, Giatromanolaki A, Retalis G, Georgoulias V. Subcutaneous administration of amifostine during fractionated radiotherapy: A randomized phase II study. J Clin Oncol. 2000; 18:2226-2233.
- Andreassen CN, Grau C, Lindegaard JC. Chemical radioprotection: A critical review of amifostine as a cytoprotector in radiotherapy. Semin Radiat Oncol. 2003; 13:62-72.
- Wasserman TH, Brizel DM. The role of amifostine as a radioprotector. Oncology (Williston Park). 2001; 15: 1349-1354; discussion 1357-1360.
- Antonadou D, Throuvalas N, Petridis A, Bolanos N, Sagriotis A, Synodinou M. Effect of amifostine on toxicities associated with radiochemotherapy in patients with locally advanced non-small-cell lung cancer. Int J Radiat Oncol Biol Phys. 2003; 57:402-408.
- Holley AK, Xu Y, St Clair DK, St Clair WH. ReIB regulates manganese superoxide dismutase gene and resistance to ionizing radiation of prostate cancer cells. Ann N Y Acad Sci. 2010; 1201:129-136.
- Murley JS, Kataoka Y, Miller RC, Li JJ, Woloschak G, Grdina DJ. SOD2-mediated effects induced by WR1065 and low-dose ionizing radiation on micronucleus formation in RKO human colon carcinoma cells. Radiat Res. 2011; 175:57-65.
- Murley JS, Kataoka Y, Baker KL, Diamond AM, Morgan WF, Grdina DJ. Manganese superoxide dismutase (SOD2)-mediated delayed radioprotection induced by the free thiol form of amifostine and tumor necrosis factor alpha. Radiat Res. 2007; 167:465-474.
- Zhang C, Ni J, Gao F, Sun D, Zhou C, Cheng Y, Cai J, Li B. The mechanism for the ameliorative effect of CpGoligodeoxynucleotides on bone marrow hemopoiesis radiation injury. Basic Clin Pharmacol Toxicol. 2011; 109:11-16.
- Afonso V, Champy R, Mitrovic D, Collin P, Lomri A. Reactive oxygen species and superoxide dismutases: Role in joint diseases. Joint Bone Spine. 2007; 74:324-329.
- Sugamura K, Keaney JF Jr. Reactive oxygen species in cardiovascular disease. Free Radic Biol Med. 2011; 51:978-992.
- 16. Silva JP, Coutinho OP. Free radicals in the regulation of

damage and cell death-basic mechanisms and prevention. Drug Discov Ther. 2010; 4:144-167.

- Schäfer M, Werner S. Oxidative stress in normal and impaired wound repair. Pharmacol Res. 2008; 58:165-171.
- Sonntag CV. The Chemical Basis of Radiation Biology. Taylor & Francis, London, UK, 1987; pp. 31-56.
- Greenberger JS, Epperly MW, Gretton J, Jefferson M, Nie S, Bernarding M, Kagan V, Guo HL. Radioprotective gene therapy. Curr Gene Ther. 2003; 3:183-195.
- Greenberger JS, Epperly MW. Radioprotective antioxidant gene therapy: Potential mechanisms of action. Gene Ther Mol Biol. 2004; 8:31-44.
- Greenberger JS, Epperly MW. Antioxidant therapeutic approaches toward amelioration of the pulmonary pathophysiological damaging effects of ionizing irradiatioin. Curr Respir Med Rev. 2007; 3:29-39.
- Riley PA. Free radicals in biology: Oxidative stress and the effects of ionizing radiation. Int J Radiat Biol. 1994; 65:27-33.
- Hall EJ, Giaccia AJ. Radiobiology for the radiologist. Lippi cott Williams & Wilkins, Philadelphia, USA, 2000.
- Lavi R, Shainberg A, Shneyvays V, Hochauser E, Isaac A, Zinman T, Friedmann H, Lubart R. Detailed analysis of reactive oxygen species induced by visible light in various cell types. Lasers Surg Med. 2010; 42:473-480.
- McCord JM, Fridovich I. Superoxide dismutase. An enzymatic function for erythrocuprein (hemocuprein). J Biol Chem. 1969; 244:6049-6055.
- Marklund SL. Human copper-containing superoxide dismutase of high molecular weight. Proc Nati Acad Sci U S A. 1982; 79:7634-7638.
- Weisiger RA, Fridovich I. Mitochrondrial superoxide dismutase. Site of synthesis and intramitochondrial localization. J Biol Chem. 1973; 248:4793-4796.
- Kinnula VL, Crapo JD. Superoxide dismutases in the lung and human lung diseases. Am J Respir Crit Care Med. 2003; 167:1600-1619.
- Dalle-Donne I, Rossi R, Colombo R, Giustarini D, Milzani A. Biomarkers of oxidative damage in human disease. Clin Chem. 2006; 52:601-623.
- Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol. 2007; 39:44-84.
- Bentzen SM. Preventing or reducing late side effects of radiation therapy: Radiobiology meets molecular pathology. Nat Rev Cancer. 2006; 6:702-713.
- Yan S, Brown SL, Kolozsvary A, Freytag SO, Lu M, Kim JH. Mitigation of radiation-induced skin injury by AAV2-mediated MnSOD gene therapy. J Gene Med. 2008; 10:1012-1018.
- 33. Williams JP, Brown SL, Georges G E, *et al.* Animal models for medical countermeasures to radiation exposure. Radiat Res. 2010; 173:557-578.
- Petkau A, Kelly K, Chelack WS, Barefoot C. Protective effect of superoxide dismutase on erythrocytes of X-irradiated mice. Biochem Biophys Res Commun. 1976; 70:452-458.
- Petkau A, Chelack WS, Pleskach SD. Protection of superoxide dismutase of white blood cells in X-irradiated mice. Life Sci. 1978; 22:867-882.
- Eastgate J, Moreb J, Nick HS, Suzuki K, Taniguchi N, Zucali JR. A role for manganese superoxide dismutase in radioprotection of hematopoietic stem cells by

interleukin-1. Blood. 1993; 81:639-646.

- Liu J, Wang X, Wang F, Teng L, Cao J. Attenuation effects of heparin-superoxide dismutase conjugate on bleomycin-induced lung fibrosis *in vivo* and radiationinduced inflammatory cytokine expression *in vitro*. Biomed Pharmacother. 2009; 63:484-491.
- Graves PR, Siddiqui F, Anscher MS, Movsas B. Radiation pulmonary toxicity: from mechanisms to management. Semin Radiat Oncol. 2010; 20:201-207.
- Rubin P, Johnston CJ, Williams JP, McDonald S, Finkelstein JN. A perpetual cascade of cytokines postirradiation leads to pulmonary fibrosis. Int J Radiat Oncol Biol Phys. 1995; 33:99-109.
- Machtay M, Scherpereel A, Santiago J, Lee J, McDonough J, Kinniry P, Arguiri E, Shuvaev VV, Sun J, Cengel K, Solomides CC, Christofidou-Solomidou M. Systemic polyethylene glycol-modified (PEGylated) superoxide dismutase and catalase mixture attenuates radiation pulmonary fibrosis in the C57/bl6 mouse. Radiother Oncol. 2006; 81:196-205.
- Barasch A, Epstein JB. Management of cancer therapyinduced oral mucositis. Dermatol Ther. 2011; 24:424-431.
- Tai Y, Inoue H, Sakurai T, Yamada H, Morito M, Ide F, Mishima K, Saito I. Protective effect of lecithinized SOD on reactive oxygen species-induced xerostomia. Radiat Res. 2009; 172:331-338.
- Nagler RM, Reznick AZ, Slavin S, Nagler A. Partial protection of rat parotid glands from irradiation-induced hyposalivation by manganese superoxide dismutase. Arch Oral Biol. 2000; 45:741-747.
- Stickle RL, Epperly MW, Klein E, Bray JA, Greenberger JS. Prevention of irradiation-induced esophagitis by plasmid/liposome delivery of the human manganese superoxide dismutase transgene. Radiat Oncol Investig. 1999; 7:204-217.
- Bernhard EJ, Maity A, Muschel RJ, McKenna WG. Effects of ionizing radiation on cell cycle progression. A review. Radiat Environ Biophys. 1995; 34:79-83.
- Gao Z, Sarsour EH, Kalen AL, Li L, Kumar MG, Goswami PC. Late ROS accumulation and radiosensitivity in SOD1-overexpressing human glioma cells. Free Radic Biol Med. 2008; 45:1501-1509.
- Epperly MW, Carpenter M, Agarwal A, Mitra P, Nie S, Greenberger JS. Intraoral manganese superoxide dismutase-plasmid/liposome (MnSOD-PL) radioprotective gene therapy decreases ionizing irradiation-induced murine mucosal cell cycling and apoptosis. In Vivo. 2004; 18:401-410.
- Fernet M, Mégnin-Chanet F, Hall J, Favaudon V. Control of the G2/M checkpoints after exposure to low doses of ionizing radiation: implications for hyperradiosensitivity. DNA Repair (Amst). 2010; 9:48-57.
- Kalen AL, Sarsour EH, Venkataraman S, Goswami PC. Mn-superoxide dismutase overexpression enhances G2 accumulation and radioresistance in human oral squamous carcinoma cells. Antioxid Redox Signal. 2006; 8:1273-1281.
- Giebel B, Bruns I. Self-renewal versus differentiation in hematopoietic stem and progenitor cells: A focus on asymmetric cell divisions. Curr Stem Cell Res Ther. 2008; 3:9-16.
- 51. Niu Y, Shen H, Epperly M, Zhang X, Nie S, Cao S, Greenberger JS. Protection of esophageal multi-lineage progenitors of squamous epithelium (stem cells) from ionizing irradiation by manganese superoxide dismutase-

plasmid/liposome (MnSOD-PL) gene therapy. In Vivo. 2005; 19:965-974.

- Tsimikas S, Brilakis ES, Miller ER, McConnell JP, Lennon RJ, Kornman KS, Witztum JL, Berger PB. Oxidized phospholipids, Lp(a) lipoprotein, and coronary artery disease. N Engl J Med. 2005; 353:46-57.
- Petkau A, Chelack WS. Radioprotective effect of superoxide dismutase on model phospholipid membranes. Biochim Biophys Acta. 1976; 433:445-456.
- Epperly MW, Tyurina YY, Nie S, Niu YY, Zhang X, Kagan V, Greenberger JS. MnSOD-plasmid liposome gene therapy decreases ionizing irradiation-induced lipid peroxidation of the esophagus. In Vivo. 2005; 19:997-1004.
- Bernardi P, Scorrano L, Colonna R, Petronilli V, Di Lisa F. Mitochondria and cell death. Mechanistic aspects and methodological issues. Eur J Biochem. 1999; 264:687-701.
- 56. Epperly MW, Melendez JA, Zhang X, Nie S, Pearce L, Peterson J, Franicola D, Dixon T, Greenberger BA, Komanduri P, Wang H, Greenberger JS. Mitochondrial targeting of a catalase transgene product by plasmid liposomes increases radioresistance *in vitro* and *in vivo*. Radiat Res. 2009; 171:588-595.
- 57. Gorman S, Tosetto M, Lyng F, Howe O, Sheahan K, O'Donoghue D, Hyland J, Mulcahy H, O'Sullivan J. Radiation and chemotherapy bystander effects induce early genomic instability events: Telomere shortening and bridge formation coupled with mitochondrial dysfunction. Mutat Res. 2009; 669:131-138.
- Helleday T, Lo J, van Gent DC, Engelward BP. DNA double-strand break repair: From mechanistic understanding to cancer treatment. DNA Repair (Amst). 2007; 6:923-935.
- Wiktor-Brown DM, Kwon HS, Nam YS, So PT, Engelward BP. Integrated one- and two-photon imaging platform reveals clonal expansion as a major driver of mutation load. Proc Nati Acad Sci U S A. 2008; 105:10314-10319.
- Harper JV, Reynolds P, Leatherbarrow EL, Botchway SW, Parker AW, O'Neill P. Induction of persistent double strand breaks following multiphoton irradiation of cycling and G1-arrested mammalian cells-replicationinduced double strand breaks. Photochem Photobiol. 2008; 84:1506-1514.
- 61. Natarajan AT, Palitti F. DNA repair and chromosomal alterations. Mutat Res. 2008; 657:3-7.
- Niu Y, Wang H, Wiktor-Brown D, Rugo R, Shen H, Huq MS, Engelward B, Epperly M, Greenberger JS. Irradiated esophageal cells are protected from radiationinduced recombination by MnSOD gene therapy. Radiat Res. 2010; 173:453-461.
- 63. Liu JF, Wang X, Tan HN, Liu H, Wang YG, Chen RQ, Cao JC, Wang FS. Effect of heparin-superoxide dismutase on γ-radiation induced DNA damage *in vitro* and *in vivo*. Drug Discov Ther. 2010; 4:355-361.
- Nielsen OS, Overgaard J, Overgaard M, Steenholdt S, Jakobsen A, Sell A. Orgotein in radiation treatment of bladder cancer. A report on allergic reactions and lack of radioprotective effect. Acta Oncol. 1987; 26:101-104.
- Hosseinimehr SJ. Trends in the development of raidioprotective agents. Drug Discov Today. 2007; 12:794-805.
- 66. Bafana A, Dutt S, Kumar A, Kumar S, Ahuja PS. The basic and applied aspects of superoxide dismutase. J Mol

Catal B Enzym. 2011; 68:129-138.

- Liu J, Zhao T, Tan H, Cheng Y, Cao J, Wang F. Pharmacokinetic analysis of *in vivo* disposition of heparinsuperoxide dismutase. Biomed Pharmacother. 2010; 64:686-691.
- 68. Zhang HW, Wang FS, Shao W, Zheng XL, Qi JZ, Cao JC, Zhang TM. Characterization and stability investigation of Cu, Zn-superoxide dismutase covalently

modified by low molecular weight heparin. Biochemistry (Mosc). 2006; 71 (Suppl. 1):S96-S100, 5.

69. Thompson JS, Chu Y, Glass J, Tapp AA, Brown SA. The manganese superoxide dismutase mimetic, M40403, protects adult mice from lethal total body irradiation. Free Radic Res. 2010; 44:529-540.

(Received July 28, 2012; Accepted August 6, 2012)

Review

An overview on antiepileptic drugs

Nirupam Das, Meenakshi Dhanawat, Sushant K. Shrivastava*

Department of Pharmaceutics, Indian Institute of Technology, Banaras Hindu University, Varanasi, India.

ABSTRACT: Epilepsy is the most common chronic neurological disorder of the brain. For several decades different kinds of medications have been used to treat epilepsy. Even though many surgical advances has been made and implemented, medications remain the basis of treatment. The search for noble antiepileptic drugs (AEDs) with more selective activity and lower toxicity continues to be an area of intensive investigation in medicinal chemistry. Additionally, drug resistance is an important clinical problem in epilepsy and is associated with an increased risk of morbidity and mortality. This review intends to present a comprehensive overview on AED in particular along with discussion on some aspects of associated drug resistance and combination therapy.

Keywords: Epilepsy, anticonvulsants, resistance, combination therapy

1. Introduction

Epilepsy is a chronic neurological disorder in which clusters of nerve cells, or neurons, in the brain sometimes signal abnormally that may remain localized (focal epilepsy) or become widespread (generalized epilepsy). The term epilepsy is derived from the Greek word epilepsia, which means "falling sickness" and can be called "seizure", "ictus", or "convulsion" (1). Both the electrical and the behavioral aspect of seizures can be quite variable and complex, even in a single patient. Seizures can be caused by a variety of pathologic conditions, including acquired injuries and genetic abnormalities. In addition, many physiologic disturbances of brain function can produce seizures and the prevalence of epilepsy varies from adults to children (2). Three to five percent of the population has a seizure sometime in their life and half to one per cent of the population have

'active epilepsy'. The heterogenicity of epilepsy makes it difficult to establish precise epidemiological statistics (3).

Etiology of epilepsy. Approximately 60% of all epilepsies are idiopathic. Almost any type of brain pathology can cause seizures/epilepsy. The underlying etiology is numerous and the abnormalities may range from symptomatic seizures due to tumor, infection, and trauma to cryptogenic forms. Cerebrovascular disease is the most commonly identified cause among adults, while prenatal insults seem to be most common among children (4). An imbalance between glutamate and γ -aminobutyric acid (GABA) neurotransmitter systems can lead to hyperexcitability. Catecholaminergic neurotransmitter systems and opioid peptides are also shown to play a role in epileptogenesis (5). Enhancement of excitatory transmission and simultaneous failure of inhibitory mechanisms together with changes in intrinsic neuronal properties results in repetitive neuronal discharges (6). Additionally, innate immunity/inflammation, adaptive immune responses, and inflammation markers including autoantibodies also play a role in the pathophysiology of several types of epilepsy (7).

Classification. Classification of epilepsy is the foundation for treatment. Several neuropsychiatrists with a special interest in epilepsy put forward the idea that the existing systems of classification of psychiatric disorders and personality disorders are inadequate as far as epilepsy is concerned. Classification of seizure type is dependent on the accuracy of history, availability and sophistication of diagnostic tests used, and age at which the patient's seizure type was classified. International Classification of Epileptic Seizures System (ICES) was introduced in 1970 and revised in 1981. Generally seizures can be classified as partial, generalized, and unclassified ones. The Commission on classification and terminology of the International League Against Epilepsy (ILAE) recently recommended new terminology and concepts on focal and generalized epilepsy. According to etiologic classification, viz., the idiopathic, symptomatic, and cryptogenic forms of epilepsy have been conceptualized as genetic, structural/metabolic, and unknown forms of epilepsy (8). A general classification of epilepsy is shown in Figure 1.

Global burden. The WHO global burden of disease (GBD) 2004 update estimates that about 40 million individuals globally have epilepsy. Inclusion of epilepsy

^{*}Address correspondence to:

Dr. Sushant Kumar Shrivastava, Department of Pharmaceutics, Indian Institute of Technology, Banaras Hindu University, Varanasi-221 005, India.

E-mail: skshrivastava.phe@itbhu.ac.in; mdanawat. rs.phe@itbhu.ac.in

Figure 1. The classification of epilepsy.

caused by other disease or injury may increase the total number of persons affected in the world to about 50 million (9). The incidence of epilepsy in developed countries is between 40-70/100,000/year and the ratio is much higher (120/100,000/year) in resource poor countries (10).

Medication. An appropriate diagnosis together with proper selection and utilization of currently available antiepileptic drugs (AEDs) is necessary for therapeutic success in the management of epilepsy. With the range of drugs currently available, there are immense opportunities for patient-tailored drug therapy. However the management of epilepsy is primarily based on optimum use of AEDs with the choice of drugs varying considerably among physicians and across countries. The choice is primarily based on evidence of efficacy and effectiveness for the individual's seizure type, but other patient-specific factors, including age, sex, childbearing potential, adverse-effect profile, comorbidities, and concomitant medications are also needed to be considered (11). Further, better understanding of pharmacoresistance would help to replace the current empiricism with a more patient-centric approach towards the management of epilepsy.

2. Antiepileptic drugs (structures see Figure 2)

Bromides were the first medication introduced by Sir Charles Locock in 1857 to provide control for seizures. Bromides were rendered obsolete due to their side effects and are being replaced by newer therapeutics. However, it has now been re-established as an addon therapy in some selected cases of intractable generalized tonic clonic seizure (12). Phenobarbital (PB) (1912) (1), a member of the barbiturate class serendipitously discovered by Alfred Hauptmann was as effective as bromides with a less toxic profile, is easier to administer and subsequently replaced the bromides. Primidone (2) is another member of the same family whose mechanism of antiepileptic action is not known. Primidone per se has anticonvulsant activity as do its two metabolites viz., PB and phenylethylmalonamide (PEMA) (3). In the 1930s, the introduction of sulfanilamide was a major medical advance, but in 1937 a sulfanilamide preparation containing diethylene glycol was one of the most consequential mass poisonings of the 20th century known as the elixir sulfanilamide disaster. This led to new regulations for

the preparation, safety, testing, labeling, distribution, and marketing of drugs. Hydantoin (glycolylurea) was first isolated in 1861. The precise mechanisms by which hydantoins work are unknown, but they are thought to exert their therapeutic effect by depressing abnormal neuronal discharges in the central nervous system (CNS). The hydantoins include phenytoin (Dilantin[®]) (4) and mephenytoin (Mesantoin[®]). Phenytoin (5,5-diphenylhydantoin, dilantin; 1938) known as the 'miracle' drug of its day was discovered by H. Houston Merritt and Tracy Putnam. This drug was used as a first choice, or when phenobarbital failed. It was one of the most widely used drugs, effective in tonic-clonic and partial seizures. An unknown substance positive to ninhydrin was found in 1949 by Roberts and Frankel in chromatographed fresh human brain tissue, which was later identified as GABA, the inhibitory neurotransmitter (13). The discovery of valproate's effectiveness as an AED created a new therapeutic paradigm. This drug was thought to be effective in enhancing GABA in the nervous system, and became one of the first drugs in which a mode of action was proposed. This drug has been licensed in the UK for clinical use since 1973 and in the USA since 1978. Its own metabolism may be enhanced by other anti-epileptic agents. In the 1970s and 1980s, the AEDs most frequently used to prevent seizures were phenobarbital, phenytoin, and carbamazepine. Later on they were found to cause major malformations, microcephaly, growth retardation, and distinctive minor abnormalities of the face and fingers in infants exposed to them during pregnancy. Gabapentin (5) (Neurontin[®]), marketed in the U.S. in 1993, was the first approved AED not metabolized in the liver, making drug interactions a lesser problem. The mode of action of gabapentin is largely unknown (14). Parker et al. (2004) (15) demonstrated that gabapentin selectively activates presynaptic GABA_B heteroreceptors. Recent studies suggest that it interacts with an auxiliary $\alpha 2\beta$ subunit of voltage-sensitive calcium channels and inhibits the calcium currents leading in turn to reduced neurotransmitter release and attenuation of postsynaptic excitability (16). Carbamazepine (Tegretol[®]) (6) is considered as a drug of choice for tonic clonic seizures, partial seizures, and trigeminal neuralgia. It works by decreasing nerve impulses that cause seizures and pain. Succinimides such as ethosuximide (Zarontin[®]) (7) and methsuccimide (Celontin[®]) are also widely used for absence (petitmal epilepsy) seizures. Another class of compounds that are widely used in the management of epilepsy is benzodiazepines (BDZs). Clinical advantages of these drugs include rapid onset of action, high efficacy rates and minimal toxicity. Among the approximately 35 BZDs available, clorazepate $(\text{Tranxene}^{\mathbb{R}})$ (8) has a distinctive and favorable profile that includes a long half-life of its active metabolite and slow onset of tolerance (17). Standard AEDs usually

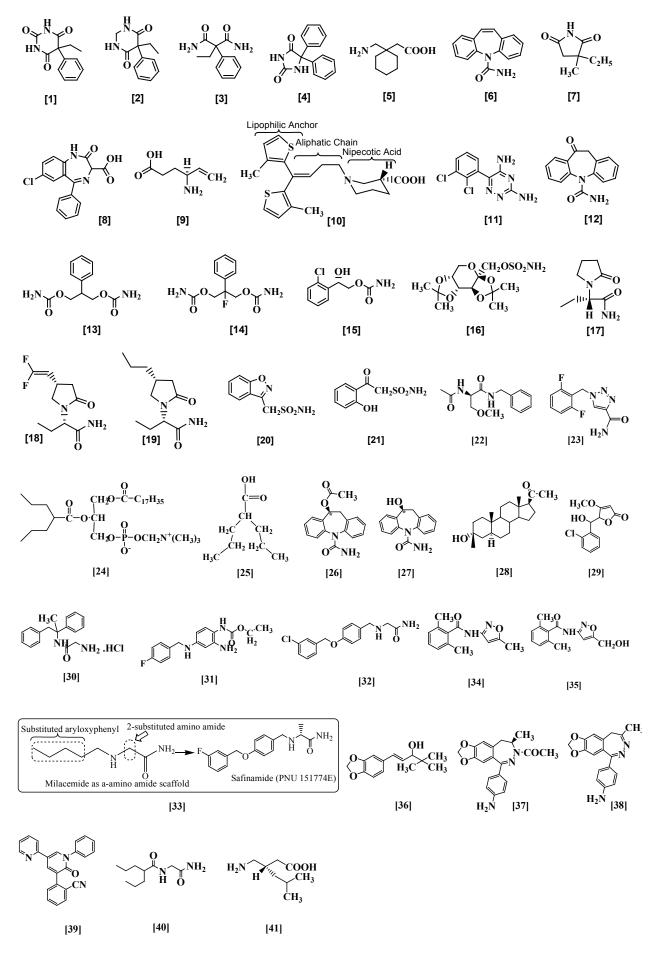


Figure 2. Chemical structures of antiepileptic drugs.

180

www.ddtjournal.com

produce side effects in 50% or more of patients treated. Rare but serious idiosyncratic reactions were reported which include agranulocytosis, Stevens-Johnson syndrome, aplastic anaemia, hepatic failure, allergic dermatitis, serum sickness, and pancreatitis (18). Once the seizure type and epilepsy syndrome have been determined, an AED can be appropriately selected. During the past decade, a number of new AEDs have been developed with diverse mechanism of action (Table 1). Most of the AEDs are efficacious for partial-onset seizures and were originally approved based on their efficacy as add-on therapy in patients with refractory partial-onset seizures. For patients with generalizedonset seizures the choice of therapy is narrower and includes valproate as well as the newer agents such as lamotrigine and topiramate. From the last 30 years many newer drugs were introduced with better safety profiles. The aim of epilepsy therapy is to keep the patient free of seizures without interfering with normal brain function. The currently available armamentarium of AEDs is discussed below.

Vigabatrin (γ -vinyl GABA, Sabril[®]) (9). Vigabatrin (VGB), a structural analogue of GABA possesses a vinyl appendage. It irreversibly inhibits GABA-transaminase (GABA-T), the enzyme responsible for the catabolism of GABA, thereby increasing the whole-brain levels of GABA making it more available to its receptor site (19). Thus VGB acts as an indirect GABA agonist. It has emerged as a first choice AED in the treatment of refractory epilepsies such as infantile spasms (also called WEST syndrome), particularly those accompanied by tuberous sclerosis (20). Long term vigabatrin treatment is associated with persistent

| visual field problems that have led to a reduction in the | | |
|---|--|--|
| use of the drug (21). In order to minimize the chance | | |
| of visual field defects, the treatment of infantile spasm | | |
| may be started for brief periods at very early onset with | | |
| low doses of VGB (22). | | |

Tiagabine (10). Tiagabine (TGB) [(R)-(-)-1-[4,4-bis(3-methyl-2-thienyl)-3-butenyl]3-piperidine carboxylic acid hydrochloride] is a nipecotic acid analogue, into which a lipophilic anchor has been incorporated to facilitate crossing of the blood-brain barrier after oral administration. It is a highly selective and potent inhibitor of GABA transporter 1 (GAT-1) in presynaptic neurons and glial cells. TGB selectively increases the amount of the inhibitory neurotransmitter GABA at the GABAergic synapse and is the first of its kind to be introduced into clinical practice (23). Isobolographic analysis of the interactions of TGB with three conventional AEDs: valproate (VPA), carbamazepine (CBZ), and phenobarbital (PB) in amygdala-kindled rats showed synergistic interaction (fixed-ratio combinations of 1:3, 1:1, and 3:1, respectively). The study suggested that TGB appears to be a valuable drug for an add-on therapy of refractory complex partial seizures in humans (24). However, a recent study revealed that the efficacy of tiagabine in newly diagnosed epilepsy is relatively low when prescribed along with other AEDs. A critical side effect such as induction of non-convulsive status epilepticus limits its use (25).

Lamotrigine (Lamictal[®]) (11). Lamotrigine (LTG), a triazine derivative that inhibits the release of glutamate (an excitatory amino acid) has been approved for use as an adjunct drug in treatment of refractory partial seizure

| Drugs | Mechanism of action |
|--|---|
| Vigabatrin | Irreversibly inhibits GABA-T |
| Tiagabine | Inhibitor of GAT-1 |
| ^Lamotrigine | Inhibits the release of excitatory neurotransmitter glutamate. It also inhibits the voltage-sensitive Na ⁺ channels (VDSC); Blockade of α 4 β 2-nAChR |
| Carbamazepine, oxcarbazepine, eslicarbazepine | Stabilize the inactivated state of VDSC |
| ^Felbamate, fluorofelbamate | Inhibits NMDA receptor. Also potentiates GABA-mediated inhibition and blocks VDSC |
| Carisbamate, rufinamide, losigamone, soretolide, valrocemide | Yet to be ascertained |
| ^Topiramate | Selectively blocks excitatory synaptic transmission mediated by GluR5 kainate receptors; also acts at VDSC |
| Levetiracetam, seletracetam, brivaracetam | Interacts with the synaptic vesicle protein 2A |
| ^Zonisamide | Block sodium channels and reduce voltage dependent T-type Ca ²⁺ currents; also modulates dopaminergic, GABA ergic, and serotonergic systems |
| ^Lacosamide | Enhances slow inactivation of VDSC and modulates CRMP-2 |
| Ganaxolone | Positive allosteric modulation of the GABA _A receptor |
| Remacemide | Potent Na ⁺ channel blocker and non-competitive NMDA channel antagonist. |
| Retigabine | It is a KCNQ K ⁺ channel opener that involves opening of neuronal Kv7.2 (KCNQ2) voltage activated K ⁺ channels |
| ^Safinamide | Antagonize the Ca ²⁺ and Na ⁺ channels; also reversibly inhibit MAO-B |
| Stiripentol | It is positive allosteric modulator of GABA _A receptor |
| Talampanel, perampanel | Non-competitively blocks AMPA receptor |
| Pregabalin | It binds potently to the α 2- γ subunit, an auxiliary protein associated with voltage-gated Ca ²⁺ channels |

Table 1. Antiepileptic drugs and their mechanism of action

^ Indicates multiple mechanisms.

with or without generalized tonic/clonic seizures. It also inhibits the voltage-sensitive sodium channels thereby stabilizing the neuronal membrane (26). Recent studies further suggested that the neuronal a4β2-nAChR (neuronal nicotinic acetylcholine receptor) is likely an important target. The blockade of $\alpha 4\beta 2$ -nAChR might represent the mechanism through which LTG effectively controls some types of epilepsy such as autosomal dominant nocturnal frontal lobe epilepsy or juvenile myoclonic epilepsy (27). Among the adverse effects, idiosyncratic drug reactions, especially skin rashes are considered fatal and may require discontinuation of the drug. The parent drug rather than a reactive metabolite causes LTG-induced skin rashes. Rash is relatively more common in children than in adults and it is safe when used in general practice to treat epilepsy inadequately controlled by other medications. Serious adverse events were rarely reported and included Stevens-Johnson syndrome (28). Long term treatment with LTG may cause hepatic inflammation and it has been found that dextran conjugate prodrug has the potential to reduce the hepatotoxicity (29). LTG monotherapy was found to be an effective treatment for children with newly diagnosed childhood absence seizures and an extendedrelease formulation (LTG-XR) may be given once daily for increasing compliance (30). Further a randomized, double-blind, placebo-controlled study reported that adjunctive therapy with LTG-XR administered once daily to a target of 200 to 500 mg/day significantly reduced weekly frequency of primary generalized tonicclonic (PGTC) seizures and increased the percentage of patients with $a \ge 50\%$ reduction in PGTC seizure frequency (31). A retrospective population-based study by Knoester et al. (2005) suggests that LTG was effective in 40% of the patients with refractory epilepsy measured by reduction in seizure frequency and retention time as observed in 165 patients. The drug is known to have a more favorable side-effect profile than conventional AEDs (32). An unblinded randomized controlled trial carried out by the Standard and New Antiepileptic Drugs (SANAD) study group found lamotrigine to be clinically better than CBZ, the standard drug treatment, for time to treatment failure outcomes and is considered as a cost-effective alternative for patients diagnosed with partial onset seizures (33).

Oxcarbazepine (TRILEPTAL[®]) (12). Oxcarbazepine (OBZ), 10,11-dihydro-10-oxo-5H-dibenz[b,f]azepine-5-carboxamide is a prodrug which is activated to eslicarbazepine in the liver. OBZ is primarily used in the treatment of epilepsy. It is also used to treat anxiety and mood disorders and benign motor ticks. OBZ is a structural derivative of CBZ, with a ketone in place of the carbon-carbon double bond on the dibenzazepine ring. This difference helps reduce the impact on the liver of metabolizing the drug, and also prevents the serious forms of anemia or agranulocytosis occasionally associated with CBZ (34). *Felbamate (Felbatol*[®]) (13). Felbamate (FBM) is structurally related to meprobamate. Its activity in epilepsy probably involves effects on the NMDA receptor. It also potentiates GABA-mediated inhibition and blocks voltage-dependent sodium channels. FBM is an effective and safe AED for either monotherapy or add-on treatment in adults with refractory partial seizures. It is also effective and safe for the treatment of refractory Lennox-Gastaut syndrome in both children and adults (35).

Fluorofelbamate (14). Fluorofelbamate (FFBM, 2-phenyl-2-fluoro-1,3-propanediol dicarbamate) is new chemical entity different from FBM in that fluorine is substituted for hydrogen in the two position of the propane. Mazarati et al. (2002) (36) studied the effectiveness of FFBM using a rat model of selfsustaining status epilepticus (SSSE). They found that FFBM exhibited a much better activity profile including no recurrent seizure activity in aborting SSSE when injected at both its early and advanced stages where diazepam and phenytoin failed to abort SSSE when administered after 40 or 70 min after the onset of stimulation. The drug candidate is designed to retain the activity of felbamate but with a different metabolic pathway that restricts the formation atropaldehyde/ acid-glutathione adduct (ATPAL-GSH and ATPA-GSH) the reactive aldehyde and acid metabolite of FBM. Thus fluorofelbamate is devoid of serious idiosyncratic toxicity associated with FBM (37). The presence of the fluoro atom protects the amide groups by its inductive effect and does not undergo the formation of ATPAL-GSH and ATPA-GSH (38).

Carisbamate (15). Carisbamate (CBM), or RWJ-333369 ((S)-2-O-carbamoyl-1-o-chlorophenylethanol), is a novel neuromodulator under investigation for the adjunctive treatment of epilepsy. This AED is structurally similar to felbamate. CBM was found to possess a broad spectrum of activity in rodent seizure and epilepsy models. Molecular action that contributes to its broad-spectrum antiepileptic activity is yet to be ascertained (39). A study was initiated to investigate mechanisms underlying the antiepileptic effects of carisbamate using the hippocampal neuronal culture models of status epilepticus and spontaneous epileptiform discharges (40). CBM has demonstrated antiepileptic activity in a variety of in vivo seizure models including hippocampal, corneal kindling, and the Genetic Absence Epilepsy Rats of Strasbourg (GAERS) model of absence epilepsy (41). It was also found to be effective in protecting against spontaneous recurrent seizures in kainate-treated animals (42) and in genetic models of epilepsy. It delays or prevents the Li-pilocarpine model of status epilepticus (43). It is rapidly and almost completely absorbed from the gut with a bioavailability of approximately 95% and with a peak plasma concentration achieved within 1-3 h (44).

Topiramate (Topamax[®]) (16). Topiramate (TPM) $(2,3:4,5-bis-O-(1-methylidene)-\beta-D-fructopyranose$ sulphamate) is a sulphamate substituted monosaccharide. The specific mechanism of action of TPM is not well understood. Preliminary reports suggested that it has a multiple mode of action. It acts at voltage-dependent sodium channels blocking the spread of seizures, it enhances GABA_A evoked chloride currents at a nonbenzodiazepine receptor site and it also antagonizes the 2-amino-3-(3-hydroxy-5-methyl-4-isoxazolyl) propionic acid (AMPA) subtype of glutamate receptors (45). Gryder and Rogawski (2003) (46) established that it selectively blocks excitatory synaptic transmission mediated by GluR5 kainate receptors. It is rapidly absorbed with linear pharmacokinetics and in the absence of enzyme inducers viz., phenytoin or carbamazepine has a plasma elimination half-life of 20-30 h. It has relatively low potential for interaction with other AEDs. However during TPM adjunctive therapy with phenytoin, CBZ, and VPA, the minimum plasma concentration level of TPM was reduced to 50%, 40%, and 14%, respectively. In addition, administration of TPM in patients already taking phenytoin twice a day, a 25% rise in the concentrations of phenytoin were found and may require a downward adjustment of phenytoin dosage (47). Besides in patients with pharmacoresistant epilepsy, Luna-Tortos et al. (2009) (48) provided evidence that brain levels of topiramate may be affected by overexpression of P-glycoprotein. Enhanced elimination of TPM was also observed during pregnancy. In particular the plasma concentration of TPM was found to decline approximately by 40% in the 2nd and 3rd trimester (49). To evaluate the efficacy of TPM in infants, Grosso et al. (2005) (50) found that TPM is effective across a broad range of seizure types in infants aged less than 2 years. The study also provides evidence regarding the usefulness of TPM in cryptogenic infantile spasms but it showed poor efficacy in symptomatic infantile spasms and in Dravet's syndrome. A randomized double-blind clinical trial demonstrated that TPM may be an alternative for phenytoin in patients for whom urgent treatment is required. TPM was found to be well tolerated as observed from the lower rate of incidence of adverse effects in the TPM treatment group (51).

Levetiracetam (Keppra[®]) (17). Levetiracetam (LEV), a water soluble pyrrolidone derivative, is the S-enantiomer of α -ethyl-2-oxo-pyrrolidine acetamide. Although LEV shares some targets (such as delayed rectifier channels and N- and P/Q-type calcium channels) with other AEDs, it is a novel AED with a unique mechanism of action related to an interaction with synaptic vesicle protein 2A (SV2A) (52). This anticonvulsant drug is structurally related to the nootropic drug piracetam. In contrast to the activity of the (S)-isomer, the (R)-form of LEV was at least 150-fold less potent in the audiogenic seizure susceptible

mouse and largely inactive in other models (53). It has been approved by the USFDA for adjunctive therapy in the treatment of partial-onset seizures in patients 16 years of age and older with epilepsy (54).

LEV analog. Seleteracetam and brivaracetam exemplifiethe analog of prototype anticonvulsant LEV that exerts site selectivity and illustrates the possibility of widening the target specificity respectively.

Seletracetam (18). Seletracetam (STM) (UCB 44212; (2*S*)-2-[(4*S*)-4-(2,2-difluorovinyl)-2oxopyrrolidin-1-yl] butanamide, is a structural analog of the AED levetiracetam which binds selectively and stereospecifically to SV2A (a novel binding site, synaptic vesicle protein 2A) with a 10-fold greater affinity than LEV. SV2A is thought to be involved with synaptic vesicle exocytosis and neurotransmitter release (55). SV2A represents a novel molecular target that seems to have an important role in the pharmacological activity of STM. The SV2A protein is thought to assist with the coordination of synaptic vesicle exocytosis and neurotransmitter release (56). Proteins involved in exocytosis, and SV2 in particular, could be considered as promising novel targets for the development of new CNS drug therapies. Discovering the mechanism of drug action through this receptor triggered a drug discovery program which led to the identification of brivaracetam (currently in phase III clinical trials for epilepsy), and seletracetam. Studies show that STM binds selectively to SV2A, without direct modulation of Na⁺ channels (57,58). A study was performed by Klitguard and coworkers on an *in vitro* high K⁺ low Ca²⁺ concentration fluid (HKLCF) model of epilepsy (mice and rat) and they have concluded that STM induces a more potent and complete suppression of neuronal synchronization than LEV. Furthermore, STM showed no psychomimetic effects and a very high tolerability index in both kindled and GAERS rats, which is markedly superior to that of LEV and other AEDs (59). Apart from various similarities seletracetam differs from levetiracetam by a very potent and selective effect against Zn^{2+} inhibition of glycine-gated currents as well as a more potent inhibition of high-voltage-operated Ca²⁺ currents and epileptiform elevation of intracellular Ca²⁺ concentrations involving multiple high-voltage-operated Ca²⁺ channels. In pharmacokinetic studies seletracetam was found to reach C_{max} within 1 h. The linear, timeindependent pharmacokinetics of the drug combined with a rapid and almost complete absorption indicates that STM has a major uncomplicated pharmacokinetic profile (60).

Brivaracetam (UCB 34714) (19). has had a drug discovery program carried out by Kenda *et al.* (2004) (61) for ligands with significant affinity to LBS (levetiracetam binding site) as a novel molecular target. Brivaracetam (BVT) ((2S)-2-[(4R)-2-oxo-4-propylpyrrolidinyl] butanamide), the 4-*n*-propyl structural analog of LEV, emerged as the single most

potent drug candidate among a series of compounds having a 4-substituted lactam ring by small-sized hydrophobic groups. It is approximately 10 times more potent than LEV as an antiseizure agent in audiogenic seizure-prone mice. Further Tai and Truong (2007) (62) demonstrated that BVT possesses more potent antiseizure and anti-myoclonic activity than LEV in an established rat model of cardiac arrest induced posthypoxic myoclonus. It possesses a binding affinity for the synaptic vesicle protein 2A (SV2A) and also shows an ability to inhibit Na⁺ channels. BVT has a half-life of 8 h and its metabolites are not pharmacologically active. In various experimental models of epilepsy, brivaracetam exhibited properties superior to LEV as an AED and has an excellent tolerability profile in humans (63). A phase II clinical trial established that it produced a dose-dependent reduction in the frequency of seizures in adults with refractory partial seizures. The drug is currently under phase III clinical trials (64).

Zonisamide (Zonegran[®]) (20). Zonisamide (ZNS) is a benzisoxazole with a sulfonamide side chain (1, 2-benzisoxazole-3-methanesulfonamide). The main actions of ZNS are blockade of sodium channels and reduction of voltage dependent T-type Ca²⁺ currents (65). It also enhances neuronal inhibition via modulation of neurotransmitter systems, including dopaminergic, GABAergic and serotonergic systems. It may enhance GABA function through interaction at allosteric or other binding sites and/or by influencing GABA transport. ZNS is also reported to be a weak inhibitor of carbonic anhydrase (66). It has a favorable pharmacokinetic profile as it is rapidly and completely absorbed and has a long half-life (63-69 h in healthy volunteers) which allows twice-daily, or even once-daily dosing. ZNS undergoes acetylation to form N-acetyl ZNS, and reduction to form the open ring metabolite, 2-sulfamoylacetyl phenol (21) that undergoes urinary excretion via glucuronide conjugation (67). It has been shown to be effective in patients whose seizures are resistant to other AEDs. Patients most often received ZNS as monotherapy. When ZNS was added to therapy with other AEDs, the dosage of the other AEDs was reduced. ZNZ or CBZ are favored over phenytoin, clobazam, valproate, or phenobarbital for simple partial and complex partial seizures (68). The drug has been approved in the US and Europe as an adjunctive therapy for refractory partial seizures in adults. It has several CNS dose-dependent side effects and by slow titration of dose the incidence could be minimized (69). A recent observational study was carried out by Kelemen et al. for accessing the efficacy and tolerability of ZNS in different resistant generalized epileptic syndromes. At a mean dose of 367 mg/day (100-600 mg/day), it was observed that ZNS showed excellent efficacy against progressive myoclonic epilepsy type 1. They suggested that the free radical scavenging and possible neuroprotective effects of zonisamide may be beneficial in acquired symptomatic epilepsies (70).

Lacosamide (Vimpat[®]) (22). Lacosamide (LCM) (formally known as harkoseride) is a new AED discovered by high throughput animal screening. Systematic evaluation of more than 100 N-benzyl-2acetamidopropionamide derivatives of this compound in animal models led to the identification of LCM (71). It is the first drug to come from a class of compounds known as functionalized amino acids and it is an optical antipode of the naturally occurring amino acid L-serine (72). It has dual mode of action as LCM enhances slow inactivation of voltage-gated sodium channels and modulates the collapsin response mediator protein-2 (CRMP-2), a protein, which is part of neuronal signal transduction pathways and which is attributed to neuroprotection (73). In contrast to AEDs such as phenytoin, CBZ, and LTG that block sodium channels when activated, LCM facilitates slow inactivation of sodium channels both in terms of kinetics and voltage dependency (74). Clinically, LCM is at present in a late stage of development as an adjunctive treatment for patients with uncontrolled partial-onset seizures. It provides high oral bioavailability unaffected by food, good tolerability with twice daily dosing, and minimal drug-drug interactions (75).

Rufinamide (Banzel[®]; Inovelon[®]) (23). Rufinamide (RFM) (1-[(2,6-difluorophenyl) methyl]-1H-1,2, 3-triazole-4-carboxamide) is a triazole derivative structurally unrelated to any currently marketed AEDs. Comparative studies of rufinamide with established AEDs (phenytoin, phenobarbital, ethosuximide, valproate) in several rodent seizure models showed the superiority of rufinamide to other AEDs tested in terms of protective indices in the electrically and chemically induced seizure tests, and the MES safety ratio (76). The drug is effective orally and is relatively well absorbed in the lower dose range. The main route of metabolism involves hydrolysis of the carboxamide group by carboxylesterases to an inactive derivative that is eliminated mainly by renal excretion via glucuronide conjugation (77). The precise mechanisms of action of RFM are unknown, however in vitro studies suggest that modulation of sodium channels activity, particularly prolongation of the inactive state may be the main mechanism of its antiepileptic activity. It possesses several favourable properties which might pave its way as the orphan drug for the treatment of partial seizures and drop attacks associated with Lennox-Gastaut syndrome (78). Coppola et al. (2011) (79) showed that it is also effective and well tolerated as an adjunctive drug for the treatment of refractory childhood-onset epileptic encephalopathies. Vendrame et al. (2010) (80) in single-centric studies observed that it is also has potential for treatment of a wide range of other seizure types including both partial and generalized epilepsy syndromes in the pediatric population. Further it showed no effect on cognitive function in patients with refractory partial seizures (81).

DP-valproic acid (DP-VPA) (24). DP-VPA (SPD 421, DP 16, TVA, RAP-valproate), a novel prodrug of VPA (25) in which the VPA moiety is covalently bound to the phospholipid lecithin based on a new drug delivery technology known as Regulated Activation of Prodrug (D-RAPTM). The ED_{50} value is 50-fold lower than VPA with a longer half-life. It showed a high absorption rate and bioavailability with negligible hepatic metabolism. Side effects are restricted to dosedependent gastrointestinal problems (82). It is currently under development for the treatment of partial and generalized seizures (83). The absorption pattern of DP-VPA follows a unique pattern whereby the complex permeates through the gut wall and enters intact to the enterocyte. Then it associates itself with chylomicrons and reaches the systemic blood circulation via the lymphatic route (84).

Eslicarbazepine acetate (BIA 2-093) (26). Eslicarbazepine acetate (ESL) [(S)-(-)-10-acetoxy-10,11dihydro-5*H*-dibenz[b,f]azepine-5-carboxamide], formerly known as BIA 2-093 is a novel central nervous system (CNS)-active agent. It belongs to the members of first-line AEDs represented by carbamazepine (firstgeneration) and oxcarbazepine (second-generation) having the dibenz/b,f/azepine nucleus bearing the 5-carboxamide substitute but is structurally different at the 10,11-position (85). This molecular variation results in differences in metabolism, preventing the formation of toxic epoxide metabolites such as carbamazepine-10,11 epoxide. It is the prodrug of ESL (S-licarbazepine (27)), the entity responsible for pharmacological activity. It is currently under clinical development for the treatment of epilepsy and bipolar disorder and acts by inhibiting voltagegated sodium channels. Among the other dibenz[b,f] azepine-5-carboxamide derivatives ESL has the highest protective index (86). A recent phase III study of ESL demonstrated that ESL in a once-daily dosage of 800 and 1,200 mg was effective in reducing standardized seizure frequency. It was well tolerated as adjunctive therapy for partial-onset seizures in patients who were refractory to treatment with standard AED therapy. Mild to moderate dizziness, headache, diplopia, somnolence, and vertigo were the most commonly reported dose related adverse effects (87).

Ganaxolone (28). Ganaxolone (GNX) (3α -hydroxy-3 β -methyl- 5α -pregnan-20-one), a neuroactive steroid currently in clinical trials represents a potential AED. It is a beta methylated synthetic analogue of allopregnanolone (3α -hydroxy-21xi, 22-oxido-21homo- 5α -pregnan-20-one) and thought to act through positive allosteric modulation of the GABA_A receptor (88). In healthy human volunteers, GNX, administered in doses ranging from 50 to 1,500 mg, either as drug alone or formulated with pharmaceutical grade excipients, is rapidly absorbed from the gastrointestinal tract after oral administration. The 3β -methyl substituent minimizes metabolism at the 3β -hydroxyl group so GNX is orally active, is not converted to the hormonally active 3-keto form, and hence lacks hormonal side effects (89). GNX has been shown to be well tolerated in adults and children and the commonly observed adverse events in children were agitation and somnolence. It is currently undergoing further development against newly diagnosed infantile spasms (90), in adults with refractory partial-onset seizures (91) and in women with catamenial epilepsy (92).

Losigamone (29). Losigamone (LSG), a racemic mixture of 5-a-5 (2-chlorphenylhydroxmethyl)-4methoxy-2(5H)-furanone is related to β -methoxybutenolides and is very similar to fadyenolides and piperolides isolated from *Piper fadyenii* and *Piper* sanctum, respectively. It is the first drug that has been identified using medicinal plant-based drug discovery (93). The mechanism of action for LSG is not clearly known, although several have been proposed (94). Initially it was thought that the anticonvulsant effect of losigamone may be due to NMDA (N-methyl-Daspartic acid) antagonism and inhibition of excitatory amino acid release (95). In vitro and in vivo experiments carried out on genetically epilepsy prone DBA/2 mice by Jones and Davies suggested that the clinically effective anticonvulsant activity of LSG is attributed to its S (+)-enantiomer rather than R (-)-enantiomer or its racemic mixture (96). Present data suggests that the drug decreases neuronal excitability via a decrease in the persistent Na⁺ current in rat hippocampal neurons (97). In a multi-center, double-blind, randomized clinical trial LSG was found to be an effective and safe add-on drug for refractory partial epilepsy in adults. The median reduction in seizure frequency as well as responder rate was significantly greater for a dose of 1,500 mg/day than for 1,200 mg/day, indicating a dose-response relationship (98).

Remacemide hydrochloride (30). Remacemide hydrochloride (RMD) [(±)-2-amino-N-(1-methyl-1,2diphenylethyl)-acetamide monohydrate] is a racemic mixture and the (-) isomer is more potent than the (+)isomer in a maximal electroshock seizure (MES) test in mice. The major route of metabolism of remacemide involves desglycination and the principal metabolite is desglycinyl-remacemide (DGR). The drug may be considered as a prodrug because the anticonvulsant effects of the drug may be primarily mediated by DGR (99). It is a two-fold more potent Na^+ channel blocker and a 100-fold more potent non-competitive NMDA channel antagonist. Further, it also exhibits a greater efficacy than RMD itself in a variety of animal seizure models (100). Regardless of the activity of DGR, RMD exhibits inconsistent clinical efficacy as add-on therapy because DGR appears to be more susceptible to hepatic enzyme induction than the parent compound (101). Studies also showed that remacemide was significantly less effective than carbamazepine in preventing seizure

recurrence. Although significant pharmacodynamic interactions were observed between remacemide and other AEDs (valproate, CBZ, phenytoin, and phenobarbital) (102), unfavorable pharmacokinetic interactions make RMD an unsuitable candidate for adjunctive treatment of epilepsy (103).

Retigabine (31). Retigabine (RTG), N-(2-amino-4-(4-fluorobenzylamino) phenyl) carbamic acid ethyl ester effective against partial-onset seizures is the first novel KCNQ opener in the late stages of clinical development with an excellent safety profile (104). Rundfeldt (1997) (105) demonstrated that RTG initiates a membrane conductance which is selective for K⁺ ions and it contributes to the anticonvulsant activity. Further studies established that it acts as a KCNQ potassium channel opener that involves opening of neuronal Kv7.2 (KCNQ2) voltage activated K^+ channels (106). Besides opening of peripheral KCNQ channels it hyperpolarizes the axotomized terminals that may constitute a novel and selective mechanism for attenuation of neuropathic pain symptoms (107). RTG as an adjunctive drug displayed promising improvement in patients with partial drug-resistant epilepsy. The most prominent adverse effects due to retigabine add-on therapy were dizziness, somnolence, and fatigue. It is metabolized primarily by glucuronidation to N-glucuronide metabolites and by acetylation (108).

Safinamide (32). By retaining the acetamide portion and replacement of the pentylamino moiety of milacemide (with residues present in the structures of substrates and inhibitors of the MAO (Mono Amine Oxidase), Pevarello et al. (1998) (109) derived the lead 2-[[4-(3-chlorobenzoxy)benzyl]amino]acetamide. As an outcome of this study, safinamide (SAF), ((S)-2-[[4-(3-fluorobenzoxy)benzyl]amino]propanamide methanesulfonate) (33), a 2-substituted amino amide emerged as a potent, orally active AED with a good safety margin. It has been shown to antagonize the calcium and sodium channels; as well as inhibit monoamine oxidase type-B (MAO-B) and the inhibition is reversible. Selectivity of SAF for the B isoform of the enzyme versus A is 5,000 and 1,000 times higher in rat and human brains, respectively (110).

Soretolide (D-2916) (34). Soretolide (SRT), (2,6dimethylbenzamide N-(5-methyl-3-isoxazolyl)) is a new potent anticonvulsant exhibiting similar pharmacological properties to those of carbamazepine. Maurizis *et al.* hypothesized that SRT follows two metabolic degradation pathways. The active metabolite D3187 (35) has a better ability to cross the blood-brain barrier than the unchanged drug in female rats which may be attributed to the longer anticonvulsant activity of SRT (111). It is effective in the MES test in rodents but STR and its active metabolite are ineffective in protecting against PTZ-induced clonic seizures and in blocking generalized seizures in the hippocampal kindling rat model (112).

Stiripentol (36). Stiripentol (STP) is an efficient drug for add-on therapy in severe myoclonic epilepsy in infancy. When combined with CBZ and clobazam, it prevents the formation of the inactive metabolite of CBZ, epoxy-carbamazepine, and hydroxylation of the active metabolite of clobazam into hydroxynorclobazam respectively (113). In vitro and in vivo studies suggest that STP can be considered as a "booster" of clobazam as the inhibitory effect of STP on CYP2C19 ($Ki = 0.14 \mu M$) was found to potentiate the antiepileptic effect of clobazam (114). It is positive allosteric modulator acting directly upon the GABA_A receptor. Although it does not solely depend on the subunit composition of the receptor, STP elicits higher activity at the α 3- or δ -subunit containing receptors. This drug with target selectivity is of particular importance if the said receptor subunits are responsible for any kind of neuronal dysfunction associated with neuronal hyperexcitability (115).

AMPA receptor antagonist talampanel (37). 2-Amino -3-(3-hydroxy-5-methyl-4-isoxazolyl) propionic acid receptors (AMPARs) play important roles in neurotransmission in the CNS and in the synaptic plasticity that underlies learning processes and memory (116). However, under certain pathological conditions the AMPARs over-activation determines neuronal cell death related to various neurological diseases such as stroke, Huntington's chorea, epilepsy, etc. Therefore, AMPAR antagonists have been considered useful as therapeutic agents for these disorders, particularly in epileptic seizures and are emerging as a promising new target for epilepsy therapy (117). The majority of the researches on AMPAR receptor antagonists are on the non-competitive (allosteric) AMPAR antagonists interacting with an allosteric AMPA binding site (Figure 3). The noncompetitive antagonists have the advantage of remaining effective independently of the level of glutamate or the polarization state of the synaptic membrane during a neurological disease (118).

Moreover, they do not influence the normal glutamatergic activity after prolonged use. Thus, in recent years some important classes of these ligands have been developed. The first lead to be identified as a selective, non-competitive AMPA receptor antagonist is 1-(4-aminophenyl)-4-methyl-7,8-methylenedioxy-5H-2,3-benzodiazepine (GYKI 52466) (38) (119). Based on this template, various 2,3-benzodiazepine compounds were synthesized and evaluated. Among all the compounds, the dioxolo-benzodiazepine talampanel also named LY300164 emerged as a highly active molecule. Its phase II clinical trials in the U.S. in patients with severe epilepsy not responsive to other drugs have yielded positive results. Phase III trials in epilepsy are underway to confirm and expand these results (120). Talampanel (TLP) (GYKI-53773, LY300164), a non-competitive AMPA receptor blocker has undergone initial assessment in patients with

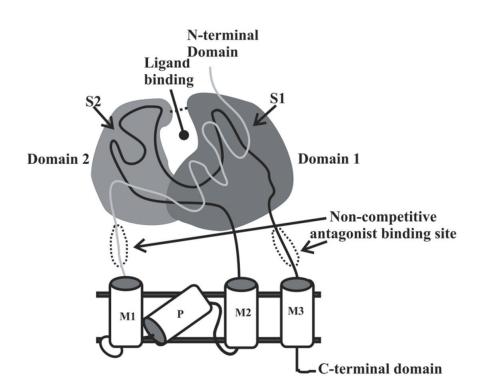


Figure 3. Schematic model of an AMPA receptor subunit. The model illustrates the agonist binding core (such as glutamate and AMPA) and competitive antagonist binding site. The putative sites of action of allosteric non-competitive antagonist talampanel are also shown. The agonist binding core consists of two domains (S1 and S2) attached by linkers to the cation channel domain consisting of three membrane-spanning segments (M1-M3) and a pore loop (P). The C-terminal domain is intracellular and the *N*-terminal domain is extracellular. (Figure reproduced with permission, Bialer *et al.*, 2007)

epilepsy. In an early adjunctive-therapy crossover trial in 49 patients with refractory partial seizures, median seizure frequency was 21% lower on talampanel than on the placebo, a statistically significant effect (121). A recent review that deals with talampanel extensively summarizes that the antiepileptic is generally well tolerated in adults with refractory complex partial seizures. The most commonly reported adverse event is dizziness and at higher doses sedation and ataxia may occur (122). A new potent noncompetitive AMPA receptor antagonist perampanel (39) that has demonstrated efficacy and good tolerability in the treatment of refractory partial onset seizures is in late stage clinical development (123). Whether the drug will prove adequately effective to reduce the morbidity and mortality of epilepsy is yet to be ensured.

Valrocemide/ TV1901 (40). Valrocemide (VGD) is a combination of VPA and glycinamide (N-valproyl glycinamide), a chemical derived from glycine, an amino acid that can have an antiepileptic effect if its concentration in the brain is increased (124). Isoherranen et al. (2001) (125) investigated the anticonvulsant activity of VGD in various animal (rodent) models of human epilepsy to determine its anticonvulsant profile and safety margin. The results obtained in this study suggest that VGD has a broad spectrum of anticonvulsant activity and promising potential as a new AED. VGD is currently under development by Teva and Acorda therapeutics as a potential therapeutic for the treatment of epilepsy. In the year 2003, a phase II trial using valrocemide as an adjunct therapy in refractory epilepsy patients had been completed and phase III trials were being planned. VGD was also being investigated for potential utility in the treatment of bipolar disorder and neuropathic pain (*126*).

Pregabalin (Lyrica[®]) (41). Pregabalin (PGB) is the alkylated analogue of the neurotransmitter GABA. Its binds potently to the $\alpha 2\gamma$ subunit, an auxiliary protein associated with voltage-gated calcium channels (VGCC) in the CNS and reduces calcium influx at nerve terminals thereby (127) modulating the release of excitatory neurotransmitters in "hyper-excited" neurons, restoring them to normal physiological state (128). It is indicated as an adjunctive therapy in adults with partial seizures with or without secondary generalization, peripheral neuropathic pain and in patients with generalized anxiety disorder or social anxiety disorder (129). A recent study by Briggs et al. suggested that PGB at a higher dose is effective in reducing the absolute frequency of secondarily generalized tonicclonic seizures in patients with clinically refractory partial epilepsy, but not secondary generalization (130).

The majority of the newer AEDs used clinically are derived from structural modification of the existing drugs. These include vigabatrin, oxcarbazepine, fluorofelbamate, brivaracetam, DP-valproic acid, eslicarbazepine, valrocemide, and pregabalin. The drugs are developed with an objective to augment the efficacy and safety margin and few of them are effective in combination. Although it may provide a solution to contained epilepsy an absolute seizure-free state is still not attainable and often "evergreening" may be one of the criteria for such structural modifications. This phenomenon of existing drug modification also does not encourage sustained research for exploring new targets. Nevertheless, few drugs acting on newer molecular targets viz., SV2A protein, AMPA receptor etc. with promising clinical trial results might shape a better therapeutic outcome. Few currently available investigational agents act at diverse targets that are involved in the pathogenesis of this complex neurological disorder and therefore, it is impossible to anticipate all the agents that provide an equivalent level of potency and efficacy. Additionally, the complexity increases when taking into consideration the nature of epileptogenesis in an individual patient. Application of plant-based drug discovery is still in its infancy as far as epilepsy is concerned and demands more screening of this novel source for generation of leads or prototype drugs as exemplified by losigamone.

3. Pharmacoresistance

Pharmacoresistance to medication is an important clinical problem in epilepsy. The phenomenon is observed in approximately one third of patients, and is associated with an increased risk of death and other ill consequences. Pharmacoresistance in epilepsy may be defined as seizures that continue to occur despite treatment trials with at least three appropriate AEDs at maximum tolerated doses (131). When a patient fails to respond to two or three appropriate AEDs then the chance of significant benefit from other drugs is 10% or less (132).

There are a number of factors associated with drug resistance and varies from one patient to another including early age of seizure onset combined with high seizure frequency, type of epileptic syndrome and seizure, structural brain lesions (e.g. cortical dysplasia) or electroencephalographic abnormalities and history of status epilepticus (133). At present two hypotheses have been asserted to explain the development of pharmacoresistance to AEDs viz., the target hypothesis and the transporter hypothesis. The target hypothesis denotes that epilepsy related changes in the molecular properties of the drug targets contribute to pharmacoresistance. For example, Remy et al. (2003) (134) studied cellular mechanisms underlying drug resistance in resected hippocampal tissues from patients. They suggested that a loss of Na⁺ channel drug sensitivity may constitute the development of resistance. The transporter hypothesis accentuates that resistance develops due to overexpression or increase in functioning of multidrug transporters in the brain, leading to poor penetration of AEDs into brain targets and thereby contributing to multidrug resistance (MDR)

in epilepsy. To substantiate the hypothesis, Volk and Loscher using a rat model of temporal lobe epilepsy demonstrated that there is an increase in expression of multidrug transporter proteins such as the ATP-binding cassette sub-family B member 1 (ABCB1, also known as MDR1 and P-glycoprotein 170) in the brain of the rat with drug-resistant spontaneous seizures (135). Siddiqui et al. (2003) (136) recognized a genetic factor associated with resistance to AEDs. They hypothesized that polymorphisms in the drug transporter gene (CC genotype at the ABCB1 C3435T) is associated with increased expression of the ABCB1 protein which inturn influences the response to AED treatment. Although no genetic stratification is underpinned, a recent study found a significant association between ABCB1 polymorphisms and drug resistance when patients were stratified by the same type of epilepsy and/or in those treated with the same AEDs (137). Researchers' efforts might target development of AEDs that are not recognized by MDR proteins or that can evade ABCB1. Alternatively, agents that inhibit these proteins (138) can be concomitantly administered with the currently available AEDs thereby decreasing the incidence of pharmacoresistance.

4. Combination therapy

Monotherapy is generally recommended for patients with newly diagnosed epilepsy. Combination therapy (CT) should only be initiated upon unresponsiveness to monotherapy. As the mechanism of action does not generally provide much guidance while combining AEDs, it is important to know the efficacy of each drug in different seizure types. CT has been found to be successful in about 30% of patients. A non-randomized trial suggested the efficacy of combination therapy in achieving seizure-freedom in epilepsies refractory to single drug treatment (139). Drug interaction is a common phenomenon observed during CT. This may be avoided by choosing non-interacting drugs. If such alternatives are not available, interacting drugs may be administered together by monitoring the plasma drug concentration followed by adjustment of dosage (140). For attaining a viable therapeutic outcome, CT requires rational combination to be tailored on an individual basis. In general, if the efficacy of two AEDs combined is shown to be additive or supra-additive and the burden of side effects is less than additive, the combination is considered to be advantageous. On the contrary if there is no observed efficacy or it is less than additive while the side effect burden is equal to additive, the combination is regarded as unfavorable (141). With reservation to evidence-based findings, Karceski et al. (2009) (142) summarized the choice of specific medications as possible add-on agents that were identified as showing efficacy based on Class I and Class II evidence in the 2004 American Academy

| Existing agent | Appropriate add-ons | | |
|-------------------------|---|--|--|
| Existing agent | Symptomatic localization related epilepsy | Idiopathic generalized epilepsy | |
| Felbamate | - | Valproate, lamotrigine | |
| Lamotrigine | Levetiracetam, topiramate, oxcarbazepine | 1 , 2 | |
| Levetiracetam | Lamotrigine, carbamazepine, oxcarbazepine | Valproate, lamotrigine, topiramate | |
| Topiramate | Lamotrigine, levetiracetam, carbamazepine | Lamotrigine, valproate, levetiracetam | |
| Valproate | Levetiracetam, oxcarbazepine, lamotrigine | Lamotrigine, topiramate, levetiracetam, zonisamide | |
| Vagus nerve stimulation | _ | Valproate, lamotrigine, topiramate | |
| Zonisamide | Levetiracetam, lamotrigine | Lamotrigine, valproate, levetiracetam | |
| Tiagabine | Lamotrigine, levetiracetam, topiramate | _ | |
| Phenytoin | Levetiracetam, lamotrigine | - | |
| Phenobarbital | Lamotrigine, levetiracetam, oxcarbazepine | - | |
| Oxcarbazepine | Levetiracetam, lamotrigine, topiramate | _ | |
| Gabapentin | Lamotrigine, levetiracetam, oxcarbazepine | _ | |
| Carbamazepine | Levetiracetam, lamotrigine | _ | |

Table 2. Combination/add-on therapy

Bold italic indicates treatment of choice.

of Neurology (AAN)/American Epilepsy Society (AES) guidelines (Table 2). Among the various possible combinations, the information from the table implied that LEV may be considered as the universal addon drug. Furthermore, recent evidence revealed that a combination of LEV with other AEDs, particularly those enhancing GABAergic inhibition, lead to additive/ synergistic effects on seizure protection with minimal side effects and pharmacokinetic interactions (*143*).

5. Conclusion

Despite the discovery of a number of AEDs, the management of epilepsy still remains an intricate task. Due to the prevalence of resistance to monotherapy, combination therapy proves workable. The utilization of available drugs to combat resistance requires rational adaptation of data arising out of clinical trials. Most of the currently available AEDs possess multiple mechanisms of action. With a few exceptions, the precise primary mode of action of some newer AEDs remains to be discovered. A multidiciplinary approach to identify potential receptor site, mechanism of action, and reason for resistance would pave the way for better therapeutic interventions towards the management of epilepsy.

References

- Riviello JJ. Classification of seizures and epilepsy. Curr Neurol Neurosci Rep. 2003; 3:325-331.
- 2. Kwan P, Brodie MJ. Neuropsychological effects of epilepsy and antiepileptic drugs. Lancet. 2001; 357:216-222.
- Eriksson KJ, Koivikko MJ. Prevalence, classification, and severity of epilepsy and epileptic syndromes in children. Epilepsia. 1997; 38:1275-1282.
- Robinson S. Systemic prenatal insults disrupt telencephalon development: Implications for potential interventions. Epilepsy Behav. 2005; 7:345-363.
- Engelborghs S, D'Hooge R, De Deyn PP. Pathophysiology of epilepsy. Acta Neurol Belg. 2000; 100:201-213.

- Duncan JS, Sander JW, Sisodiya SM, Walker MC. Adult epilepsy. Lancet. 2006; 367:1087-1100.
- Granata T, Cross H, Theodore W, Avanzini G. Immunemediated epilepsies. Epilepsia. 2011; 52:5-11.
- Berg AT, Scheffer IE. New concepts in classification of the epilepsies: entering the 21st century. Epilepsia. 2011; 52:1058-1062.
- World Health Organization. Epilepsy. http://www.who.int/ mediacentre/factsheets/fs999/en/ (accessed July 29, 2012).
- de Boer HM, Mula M, Sander JW. The global burden and stigma of epilepsy. Epilepsy Behav. 2008; 12:540-546.
- 11. Perucca E, Tomson T. The pharmacological treatment of epilepsy in adults. Lancet Neurol. 2011; 10:446-456.
- 12. Pearce JMS. Bromide. The first effective antiepileptic agent. J Neurol Neurosurg Psychiatry. 2002; 72:412.
- Cereghino JJ. The major advances in epilepsy in the 20th century and what we can expect (hope for) in the future. Epilepsia. 2009; 50:351-357.
- Jensen AA, Mosbacher J, Elg S, Lingenhoehl K, Lohmann T, Johansen TN, Abrahamsen B, Mattsson JP, Lehmann A, Bettler B, Bräuner-Osborne H. The anticonvulsant gabapentin (neurontin) does not act through gammaaminobutyric acid-B receptors. Mol Pharmacol. 2002; 61:1377-1384.
- Parker DA, Ong J, Marino V, Kerr DI. Gabapentin activates presynaptic GABA B heteroreceptors in rat cortical slices. Eur J Pharmacol. 2004; 495:137-143.
- 16. Sills GJ. The mechanisms of action of gabapentin and pregabalin. Curr Opin Pharmacol. 2006; 6:108-113.
- Riss J, Cloyd J, Gates J, Collins S. Benzodiazepines in epilepsy: Pharmacology and pharmacokinetics. Acta Neurol Scand. 2008; 118:69-86.
- Shorvon S, Stefan H. Overview of the safety of newer antiepileptic drugs. Epilepsia. 1997; 38:S45-S51.
- Willmore LJ, Abelson MB, Ben-Menachem E, Pellock JM, Shields WD. Vigabatrin: 2008 update. Epilepsia. 2009; 50:163-173.
- Hwang H, Kim KJ. New antiepileptic drugs in pediatric epilepsy. Brain Dev. 2008; 30:549-555.
- McDonagh J, Stephen LJ, Dolan FM *et al*. Peripheral retinal dysfunction in patients taking vigabatrin. Neurology. 2003; 61:1690-1694.
- 22. Parisi P, Bombardieri R, Curatolo P. Current role of vigabatrin in infantile spasms. Eur J Paediatr Neurol. 2007;

11:331-336.

- Landmark CJ. Targets for antiepileptic drugs in the synapse. Med Sci Monit. 2007; 13:RA1-RA7.
- Borowicz KK, Zadrozniak M, Luszczki JJ, Czuczwar SJ. Interactions between tiagabine and conventional antiepileptic drugs in the rat model of complex partial seizures. J Neural Transm. 2008; 115:661-667.
- Bauer J, Cooper-Mahkorn D. Tiagabine: Efficacy and safety in partial seizures-current status. Neuropsychiatr Dis Treat. 2008; 4:731-736.
- Messenheimer JA. Lamotrigine. Epilepsia. 1995; 36: S87-S95.
- Zheng C, Yang K, Liu Q, Wang MY, Shen J, Valles AS, Lukas RJ, Barrantes FJ, Wu J. The anticonvulsive drug lamotrigine blocks neuronal α4β2 nicotinic acetylcholine receptors. Pharmacol Exp Ther. 2010; 335:401-408.
- Mackay FJ, Wilton LV, Peace GL, Freemantle SN, Mann RD. Safety of long-term lamotrigine in epilepsy. Epilepsia. 1997; 38:881-886.
- 29. Pugazhendhy S, Shrivastava PK, Sinha SK, Shrivastava SK. Lamotrigine-dextran conjugates-synthesis, characterization, and biological evaluation. Med Chem Res. 2001; 20:595-600.
- Blaszczyk B, Czuczwar SJ. Efficacy, safety, and potential of extended-release lamotrigine in the treatment of epileptic patients. Neuropsychiatr Dis Treat. 2010; 6:145-150.
- Biton V, Di Memmo J, Shukla R, Lee YY, Poverennova I, Demchenko V, Saiers J, Adams B, Hammer A, Vuong A, Messenheimer J. Adjunctive lamotrigine XR for primary generalized tonic-clonic seizures in a randomized, placebo-controlled study. Epilepsy Behav. 2010; 19:352-358.
- Knoester PD, Keyser A, Renier WO, Egberts ACG, Hekster YA, Deckers CLP. Effectiveness of lamotrigine in clinical practice: Results of a retrospective populationbased study. Epilepsy Res. 2005; 65:93-100.
- 33. Marson AG, Al-Kharusi AM, Alwaidh M, et al. The SANAD study of effectiveness of carbamazepine, gabapentin, lamotrigine, oxcarbazepine, or topiramate for treatment of partial epilepsy: An unblinded randomised controlled trial. Lancet. 2007; 369:1000-1015.
- Shorvon S. Oxcarbazepine: A review. Seizure. 2000; 9:75-79.
- Schmidt D. Felbamate: Successful development of a new compound for the treatment of epilepsy. Epilepsia. 1993; 34:S30-S33.
- Mazarati AM, Sofia RD, Wasterlain CG. Anticonvulsant and antiepileptogenic effects of fluorofelbamate in experimental status epilepticus. Seizure. 2002; 11:423-430.
- Roecklein BA, Sacks HJ, Mortko H, Stables J. Fluorofelbamate. Neurotherapeutics. 2007; 4:97-101.
- Landmark CJ, Johannessen SI. Modifications of antiepileptic drugs for improved tolerability and efficacy. Perspect Medicin Chem. 2008; 2:21-39.
- Novak GP, Kelley M, Zannikos P, Klein B. Carisbamate (RWJ-333369). Neurotherapeutics. 2007; 4:106-109.
- 40. Deshpande LS, Nagarkatti N, Sombati S, DeLorenzo RJ. The novel antiepileptic drug carisbamate (RWJ 333369) is effective in inhibiting spontaneous recurrent seizure discharges and blocking sustained repetitive firing in cultured hippocampal neurons. Epilepsy Res. 2008; 79:158-165.
- Nehlig A, Rigoulot MA, Boehrer A. A new drug, RWJ-333369, displays potent antiepileptic properties in genetic models of absence and audiogenic epilepsy.

Epilepsia. 2005; 46:215.

- 42. Grabenstatter HL, Dudek FE. The use of chronic models in antiepileptic drug discovery: The effect of RWJ-333369 on spontaneous motor seizures in rats with kainate-induced epilepsy. Epilepsia. 2004; 45:197.
- Francois J, Ferrandon A, Koning E, Nehlig A. A new drug RWJ 333369 protects limbic areas in the lithiumpilocarpine model (lipilo) of epilepsy and delays or prevents the occurrence of spontaneous seizures (abstract). Epilepsia. 2005; 46:269.
- 44. Yao C, Doose DR, Novak G, Bialer M. Pharmacokinetics of the new antiepileptic and CNS drug RWJ-333369 following single and multiple dosing to humans. Epilepsia. 2006; 47:1822-1829.
- Walker MC, Sander JW. Topiramate: A new antiepileptic drug for refractory epilepsy. Seizure. 1996; 5:199-203.
- Gryder DS, Rogawski MA. Selective antagonism of GluR5 kainate-receptor-mediated synaptic currents by topiramate in rat basolateral amygdala neurons. J Neurosci. 2003; 23:7069-7074.
- Rosenfeld WE. Topiramate: A review of preclinical, pharmacokinetic, and clinical data. Clin Ther. 1997; 19:1294-1308.
- Luna-Tortos C, Rambeck B, Jurgens UH, Loscher W. The antiepileptic drug topiramate is a substrate for human P-glycoprotein but not multidrug resistance proteins. Pharm Res. 2009; 26:2464-2470.
- Ohman I, Sabers A, de Flon P, Luef G, Tomson T. Pharmacokinetics of topiramate during pregnancy. Epilepsy Res. 2009; 87:124-129.
- Grosso S, Galimberti D, Farnetani MA, Cioni M, Mostardini R, Vivarelli R, Di Bartolo RM, Bernardoni E, Berardi R, Morgese G, Balestri P. Efficacy and safety of topiramate in infants according to epilepsy syndromes. Seizure. 2005; 14:183-189.
- 51. Ramsay E, Faught E, Krumholz A, Naritoku D, Privitera M, Schwarzman L, Mao L, Wiegand F, Hulihan J. Efficacy, tolerability, and safety of rapid initiation of topiramate versus phenytoin in patients with new-onset epilepsy: A randomized double-blind clinical trial. Epilepsia. 2010; 51:1970-1977.
- Hamann M, Sander SE, Richter A. Brivaracetam and seletracetam, two new SV2A ligands, improve paroxysmal dystonia in the dt sz mutant hamster. Eur J Pharmacol. 2008; 601:99-102.
- Noyer M, Gillard M, Matagne A, Henichart JP, Wulfert E. The novel antiepileptic drug levetiracetam (ucb L059) appears to act *via* a specific binding site in CNS membranes. Eur J Pharmacol. 1995; 286:137-146.
- 54. Ulloa CM, Towfigh A, Safdieh J. Review of levetiracetam, with a focus on the extended release formulation, as adjuvant therapy in controlling partial-onset seizures. Neuropsychiatr Dis Treat. 2009 5:467-476.
- Lynch BA, Lambeng N, Nocka K, Kensel-Hammes P, Bajjalieh SM, Matagne A, Fuks B. The synaptic vesicle protein SV2A is the binding site for the antiepileptic drug levetiracetam. Proc Natl Acad Sci U S A. 2004; 101:9861-9866.
- Crowder KM, Gunther JM, Jones TA, Hale BD, Zhang HZ, Peterson MR, Scheller RH, Chavkin C, Bajjalieh SM. Abnormal neurotransmission in mice lacking synaptic vesicle protein 2A (SV2A). Proc Nat Acad Sci U S A. 1999; 96:15268-15273.
- 57. Zona C, Niespodziany I, Pieri M, Klitgaard H, Margineanu DG. Seletracetam (ucb 44212), a new pyrrolidone

derivative, lacks effect on Na⁺ currents in rat brain neurons *in vitro*. Epilepsia. 2005; 46:116.

- Rigo JM, Nguyen L, Hans G, Belachew S, Moonen G, Matagne A, Klitgaard H. Seletracetam (ucb 44212): Effect on inhibitory and excitatory neurotransmission. Epilepsia. 2005; 46:110-111.
- Matagne A, Margineanu DG, Potschka H, Loscher W, Michel P, Kenda B, Klitgaard H. Profile of the new pyrrolidone derivative seletracetam (ucb 44212) in animal models of epilepsy. Eur J Pharmacol. 2009; 614:30-37.
- Bennett B, Matagne A, Michel P, Leonard M, Cornet M, Meeus MA, Toublanc N. Seletracetam (UCB 44212). Neurotherapeutics. 2007; 4:117-122.
- Kenda BM, Matagne AC, Talaga PE, Pasau PM, Differding E, Lallemand BI, Frycia AM, Moureau FG, Klitgaard HV, Gillard MR, Fuks B, Michel P. Discovery of 4-substituted pyrrolidone butanamides as new agents with significant antiepileptic activity. J Med Chem. 2004; 47:530-549.
- Tai KK, Truong DD. Brivaracetam is superior to levetiracetam in a rat model of post-hypoxic myoclonus. J Neural Transm. 2007; 114:1547-1551.
- Matagne A, Margineanu DG, Kenda B, Michel P, Klitgaard H. Anti-convulsive and anti-epileptic properties of brivaracetam (ucb 34714), a high-affinity ligand for the synaptic vesicle protein, SV2A. Br J Pharmacol. 2008; 154:1662-1671.
- Rogawski M. Brivaracetam: A rational drug discovery success story. Br J Pharmacol. 2008; 154:1555-1557.
- Stefan H, Feuerstein TJ. Novel anticonvulsant drugs. Pharmacol Ther. 2007; 113:165-183.
- Baulac M. Introduction to zonisamide. Epilepsy Res. 2006; 68:S3-S9.
- 67. Leppik IE. Zonisamide: Chemistry, mechanism of action, and pharmacokinetics. Seizure. 2004; 13:S5-S9.
- Ohtahara S. Zonisamide in the management of epilepsy-Japanese experience. Epilepsy Res. 2006; 68:S25-S33.
- Zaccara G, Tramacere L, Cincotta M. Drug safety evaluation of zonisamide for the treatment of epilepsy. Expert Opin Drug Saf. 2011; 10:623-631.
- Kelemen A, Rasonyl G, Neuwirth M, Barcs G, Szucs A, Jakus R, Fabo D, Juhos V, Palfy B, Halasz P. Our clinical experience with zonisamide in resistant generalized epilepsy syndromes. Ideggyogy Sz. 2011; 64:187-192.
- 71. Perucca E, Yasothan U, Clincke G, Kirkpatrick P. Lacosamide. Nat Rev Drug Discov. 2008; 7:973-974.
- Luszczki JJ. Third-generation antiepileptic drugs: Mechanisms of action, pharmacokinetics and interactions. Pharmacol Rep. 2009; 61:197-216.
- Saussele T. Lacosamide. A new antiepileptic drug as adjunctive therapy in patients with partial-onset seizures. Med Monatsschr Pharm. 2008; 31:374-377.
- Curia G, Biagini G, Perucca E, Avoli M. Lacosamide: A new approach to target voltage-gated sodium currents in epileptic disorders. CNS Drugs. 2009; 23:555-568.
- 75. Doty P, Rudd GD, Stoehr T, Thomas D. Lacosamide. Neurotherapeutics. 2007; 4:145-148.
- White HS, Franklin MR, Kupferberg HJ, Schmutz M, Stables JP, Wolf HH. The anticonvulsant profile of rufinamide (CGP 33101) in rodent seizure models. Epilepsia. 2008; 49:1213-1220.
- Perucca E, Cloyd J, Critchley D, Fuseau E. Rufinamide: Clinical pharmacokinetics and concentration-response relationships in patients with epilepsy. Epilepsia. 2008; 49:1123-1141.
- 78. Herman ST. Adopting an Orphan Drug: Rufinamide for

Lennox-Gastaut Syndrome. Epilepsy Curr. 2009; 9:72-74.

- 79. Coppola G, Grosso S, Franzoni E, Veggiotti P, Zamponi N, Parisi P, Spalice A, Habetswallner F, Fels A, Verrotti A, D'Aniello A, Mangano S, Balestri A, Curatolo P, Pascotto A. Rufinamide in refractory childhood epileptic encephalopathies other than Lennox-Gastaut syndrome. Eur J Neurol. 2011; 18:246-251.
- Vendrame M, Loddenkemper T, Gooty VD, Takeoka M, Rotenberg A, Bergin AM, Eksioglu YZ, Poduri A, Duffy FH, Libenson M, Bourgeois BF, Kothare SV. Experience with rufinamide in a pediatric population: a single center's experience. Pediatr Neuro. 2010; 43:155-158.
- Wisniewski CS. Rufinamide: A new antiepileptic medication for the treatment of seizures associated with lennox-gastaut syndrome. Ann Pharmacother. 2010; 44:658-667.
- Labiner DM. DP-VPA D-Pharm. Curr Opin Investig Drugs. 2002; 3:921-923.
- Trojnar MK, Wierzchowska-Cioch E, Krzyzanowski M, Jargiello M, Czuczwar SJ. New generation of valproic acid. Pol J Pharmacol. 2004; 56:283-288.
- Dahan A, Duvdevani R, Shapiro I, Elmann A, Finkelstein E, Hoffman A. The oral absorption of phospholipid prodrugs: *In vivo* and *in vitro* mechanistic investigation of trafficking of a lecithin-valproic acid conjugate following oral administration. J Control Release. 2008; 126:1-9.
- Bialer M, Johannessen SI, Levy RH, Perucca E, Tomson T, White HS. Progress report on new antiepileptic drugs: A summary of the Ninth Eilat Conference (EILAT IX). Epilepsy Res. 2009; 83:1-43.
- Ambrosio AF, Soares-Da-Silva P, Carvalho CM, Carvalho AP. Mechanisms of action of carbamazepine and its derivatives, oxcarbazepine, BIA 2-093, and BIA 2-024. Neurochem Res. 2002; 27:121-130.
- Elger C, Halasz P, Maia J, Almeida L, Soares-da-Silva P. Efficacy and safety of eslicarbazepine acetate as adjunctive treatment in adults with refractory partial-onset seizures: A randomized, double-blind, placebo-controlled, parallelgroup phase III study. Epilepsia. 2009; 50:454-463.
- Ungard JT, Beekman M, Gasior M, Carter RB, Dijkstra D, Witkin JM. Modification of behavioral effects of drugs in mice by neuroactive steroids. Psychopharmacology. 2000; 148:336-343.
- Monaghan EP, McAculey JW, Data JL. Ganaxolone: A novel positive allosteric modulator of the GABA_A receptor complex for the treatment of epilepsy. Exp Opin Invest Drugs. 1999; 8:1663-1671.
- Malphrus AD, Wilfong AA. Use of the newer antiepileptic drugs in pediatric epilepsies. Curr Treat Options Neurol. 2007; 9:256-267.
- Pieribone VA, Tsai J, Soufflet C, Rey E, Shaw K, Giller E, Dulac O. Clinical evaluation of ganaxolone in pediatric and adolescent patients with refractory epilepsy. Epilepsia. 2007; 48:1870-1874.
- Reddy DS, Rogawski MA. Neurosteroid replacement therapy for catamenial epilepsy. Neurotherapeutics. 2009; 6:392-401.
- Chatterjee SS, Noldner M. Losigamone: From plant extract to antiepileptic drug. CNS Drug Rev. 1997; 3:225-244.
- Dimpfel W, Chatterjee SS, Noldner M, Ticku MK. Effects of the anticonvulsant losigamone and its isomers on the GABA_A receptor system. Epilepsia. 1995; 36:983-989.
- Srinivasan J, Richens A, Davies JA. The effect of losigamone (AO-33) on electrical activity and excitatory amino acid release in mouse cortical slices. Br J Pharmacol.

1997; 122:1490-1494.

- Jones FA, Davies JA. The anticonvulsant effects of the enantiomers of losigamone. Br J Pharmacol. 1999; 128:1223-1228.
- Gebhardt C, Breustedt JM, Noldner M, Chatterjee SS, Heinemann U. The antiepileptic drug losigamone decreases the persistent Na⁺ current in rat hippocampal neurons. Brain Res. 2001; 920:27-31.
- Baulac M, Klement S. Efficacy and safety of losigamone in partial seizures: A randomized double-blind study. Epilepsy Res. 2003; 55:177-189.
- Santangeli S, Sills GJ, Stone TW, Brodie MJ. Differential effects of remacemide and desglycinyl-remacemide on epileptiform burst firing in the rat hippocampal slice. Neurosci Lett. 2002; 321:33-36.
- Małek R, Borowicz KK, Kimber-Trojnar Z, Sobieszek G, Piskorska B, Czuczwar SJ. Remacemide-a novel potential antiepileptic drug. Pol J Pharmacol. 2003; 55:691-698.
- 101. Sills GJ, Santangeli S, Forrest G, Brodie MJ. Influence of cytochrome P450 induction on the pharmacokinetics and pharmacodynamics of remacemide hydrochloride. Epilepsy Res. 2002; 49:247-254.
- 102. Wesnes KA, Edgar C, Dean AD, Wroe SJ. The cognitive and psychomotor effects of remacemide and carbamazepine in newly diagnosed epilepsy. Epilepsy Behav. 2009; 14:522-528.
- 103. Borowicz KK, Malek R, Luszczki JJ, Ratnaraj N, Patsalos PN, Czuczwar SJ. Isobolographic analysis of interactions between remacemide and conventional antiepileptic drugs in the mouse model of maximal electroshock. Epilepsy Behav. 2007; 11:6-12.
- 104. Porter RJ, Nohria V, Rundfeldt C. Retigabine. Neurotherapeutics. 2007; 4:149-154.
- 105. Rundfeldt C. The new anticonvulsant retigabine (D-23129) acts as an opener of K⁺ channels in neuronal cells. Eur J Pharmacol. 1997; 336:243-249.
- 106. Wuttke TV, Seebohm G, Bail S, Maljevic S, Lerche H. The new anticonvulsant retigabine favors voltage-dependent opening of the Kv7.2 (KCNQ2) channel by binding to its activation gate. Mol Pharmacol. 2005; 67:1009-1017.
- 107. Roza C, Lopez-Garcia JA. Retigabine, the specific KCNQ channel opener, blocks ectopic discharges in axotomized sensory fibres. Pain. 2008; 138:537-545.
- Czuczwar P, Wojtak A, Cioczek-Czuczwar A, Parada-Turska J, Maciejewski R, Czuczwar SJ. Retigabine: The newer potential antiepileptic drug. Pharmacol Rep. 2010; 62:211-219.
- 109. Pevarello P, Bonsignori A, Dostert P, Heidempergher F, Pinciroli V, Colombo M, McArthur RA, Salvati P, Post C, Fariello RG, Varasi M. Synthesis and anticonvulsant activity of a new class of 2-[(arylalkyl)amino]alkanamide derivatives. J Med Chem. 1998; 41:579-590.
- Fariello RG. Safinamide. Neurotherapeutics. 2007; 4:110-116.
- 111. Maurizis JC, Madelmont JC, Rapp M, Marijnen C, Cerf MC, Gillardin JM, Lepage F, Veyre A. Disposition and metabolism of 2,6-dimethylbenzamide *N*-(5-methyl-3isoxazolyl) (D2916) in male and female rats. Drug Metab Dispos. 1997; 25:33-39.
- 112. Perucca E, Kupferberg HJ. Drugs in early clinical development. In: Antiepileptic drugs (Levy RH, Mattson RH, Meldrum BS, Perucca E, eds.). Lippincott Williams & Wilkins, Philadelphia, USA, 2002; pp. 913-992.
- 113. Chiron C, Marchand MC, Tran A, Rey E, d'Athis P, Vincent J, Dulac O, Pons G. Stiripentol in severe myoclonic

epilepsy in infancy: A randomised placebo-controlled syndrome-dedicated trial. Lancet. 2000; 356:1638-1642.

- 114. Giraud C, Treluyer JM, Rey E, Chiron C, Vincent J, Pons G, Tran A. *In vitro* and *in vivo* inhibitory effect of stiripentol on clobazam metabolism. Drug Metab Dispos. 2006; 34:608-611.
- 115. Fisher JL. The anti-convulsant stiripentol acts directly on the GABA (A) receptor as a positive allosteric modulator. Neuropharmacology. 2009; 56:190-197.
- Lees GJ. Pharmacology of AMPA/Kainate receptor ligands and their therapeutic potential in neurological and psychiatric disorders. Drugs. 2000; 59:33-78.
- 117. De Sarro G, Gitto R, Russo E, Ibbadu GF, Barreca ML, De Luca L, Chimirri A. AMPA receptor antagonists as potential anticonvulsant drugs. Curr Top Med Chem. 2005; 5:31-42.
- 118. Chimirri A, Gitto R, Zappala M. AMPA receptor antagonists. Exp Opin Ther Patents. 1999; 9:557-570.
- 119. Donevan SD, Rogawski MA. GYKI 52466, a 2,3-benzodiazepine, is a highly selective, noncompetitive antagonist of AMPA/kainate receptor responses. Neuron. 1993; 10:51-59.
- 120. Gitto R, Barreca ML, De Luca L, De Sarro G, Ferreri G, Quartarone S, Russo E, Constanti A, Chimirri A. Discovery of a novel and highly potent noncompetitive AMPA receptor antagonist. J Med Chem. 2003; 46:197-200.
- 121. Perucca E, French J, Bialer M. Development of new antiepileptic drugs: Challenges, incentives, and recent advances. Lancet Neurol. 2007; 6:793-804.
- 122. Howes JF, Bell C. Talampanel. Neurotherapeutics. 2007; 4:126-129.
- 123. Rogawski MA. Revisiting AMPA receptors as an antiepileptic drug target. Epilepsy Curr. 2011; 11:56-63.
- 124. Valrocemide licensed for worldwide development, production and marketing. http://www.epilepsy.org.uk/ node/1133 (accessed July 29, 2012).
- 125. Isoherranen N, Woodhead JH, White HS, Bialer M. Anticonvulsant profile of valrocemide (TV1901): A new antiepileptic drug. Epilepsia. 2001; 42:831-836.
- Hovinga CA. Valrocemide (Teva/Acorda). Curr Opin Investig Drugs. 2004; 5:101-106.
- 127. Hamandi K, Sander JW. Pregabalin: A new antiepileptic drug for refractory epilepsy. Seizure. 2006; 15:73-78.
- Kavoussi R. Pregabalin: From molecule to medicine. Eur Neuropsychopharmacol. 2006; 2:S128-S133.
- Dworkin RH, Kirkpatrick P. Fresh from the pipeline: Pregabalin. Nat Rev Drug Discov. 2005; 4:455-456.
- Briggs DE, Lee CM, Spiegel K, French JA. Reduction of secondarily generalized tonic-clonic (SGTC) seizures with pregabalin. Epilepsy Res. 2008; 82:86-92.
- Sisodiya SM. Genetics of drug resistance in epilepsy. Curr Neurol Neurosci Rep. 2005; 5:307-311.
- Pati S, Alexopoulos AV. Pharmacoresistant epilepsy: From pathogenesis to current and emerging therapies. Cleve Clin J Med. 2010; 77:457-467.
- Regesta G, Tanganelli P. Clinical aspects and biological bases of drug-resistant epilepsies. Epilepsy Res. 1999; 34:109-122.
- 134. Remy S, Gabriel S, Urban BW, Dietrich D, Lehmann TN, Elger CE, Heinemann U, Beck H. A novel mechanism underlying drug resistance in chronic epilepsy. Ann Neurol. 2003; 53:469-479.
- 135. Volk HA, Loscher W. Multidrug resistance in epilepsy: Rats with drug-resistant seizures exhibit enhanced brain expression of P-glycoprotein compared with rats with

- 136. Siddiqui A, Kerb R, Weale ME, Brinkmann U, Smith A, Goldstein DB, Wood NW, Sisodiya SM. Association of multidrug resistance in epilepsy with a polymorphism in the drug-transporter gene *ABCB1*. N Engl J Med. 2003; 348:1442-1448.
- 137. Sanchez MB, Herranz JL, Leno C, Arteaga R, Oterino A, Valdizan EM, Nicolas JM, Adin J, Armijo JA. Genetic factors associated with drug-resistance of epilepsy: Relevance of stratification by patient age and aetiology of epilepsy. Seizure. 2010; 19:93-101.
- 138. Friedenberg WR, Rue M, Blood EA, Dalton WS, Shustik C, Larson RA, Sonneveld P, Greipp PR. Phase III study of PSC-833 (valspodar) in combination with vincristine, doxorubicin, and dexamethasone (valspodar/VAD) versus VAD alone in patients with recurring or refractory multiple myeloma (E1A95): A trial of the Eastern Cooperative Oncology Group. Cancer. 2006; 106:830-838.

- Peltola J, Peltola M, Raitanen J, Keranen T, Kharazmi E, Auvinen A. Seizure-freedom with combination therapy in localization-related epilepsy. Seizure. 2008; 17:276-280.
- 140. Patsalos PN, Froscher W, Pisani F, van Rijn CM. The importance of drug interactions in epilepsy therapy. Epilepsia. 2002; 43:365-385.
- 141. French JA, Faught E. Rational polytherapy. Epilepsia. 2009; 50:63-68.
- 142. Karceski S, Morrell MJ, Carpenter D. Treatment of epilepsy in adults: Expert opinion. Epilep Behav. 2005; 7: S1-S64.
- 143. Kaminski RM, Matagne A, Patsalos PN, Klitgaard H. Benefit of combination therapy in epilepsy: A review of the preclinical evidence with levetiracetam. Epilepsia. 2009; 50:387-397.

(Received January 20, 2012; Revised August 7, 2012; Accepted August 11, 2012)

Brief Report

DOI: 10.5582/ddt.2012.v6.4.194

Anti-inflammatory activities of fractions from *Geranium nepalense* and related polyphenols

Chunhua Lu¹, Yaoyao Li¹, Liji Li², Lanying Liang³, Yuemao Shen^{1,*}

¹ Key Laboratory of Chemical Biology (Ministry of Education), School of Pharmaceutical Sciences, Shandong University, Ji'nan, Shandong, China;

² Kunming Institute of Medical Sciences, Kunming, Yunnan, China;

³ Chinese Medicine Hospital of Yuncheng County, Yuncheng, Shandong, China.

ABSTRACT: Geranium nepalense Sweet is a common Chinese herbal medicine and has been used as influenza, dysentery, antiphlogistic and analgesic tonic, hemostatic, stomachic, and antidiabetic drugs. The anti-inflammatory effects of G. nepalense on tetradecanoyl phorbol acetate (TPA)-induced mouse ear edema were studied in this work. The results showed that ethyl acetate fraction of the water extract of G. nepalense possessed significant activity at 2.5 g/kg (p < 0.01) with aspirin as a positive control (0.6 g/kg). Six polyphenolic compounds, including three flavonoids, *i.e.* kaempferol, kaempferol-7-O-β-D-glucopyranoside, and quercetin-7-O-αrhamnopyranoside, and two tannins, *i.e.* pyrogallol and gallic acid, and one lignin, i.e. epipinoresinol, were isolated and characterized from ethyl acetate fraction. The isolation of polyphenols provides a clue for beneficial effects of G. nepalense in the demonstrated anti-inflammatory activity.

Keywords: Geranium nepalense, anti-inflammatory activity, TPA-induced mouse ear edema, polyphenolic, flavonoids

1. Introduction

Geranium nepalense Sweet (Geraniaceae) is widely distributed in China (1). It has been used to treat various inflammatory conditions, including influenze, dysentery, antiphlogistic and analgesic, and used as a Chinese herbal medicine (2,3). The extract of G. nepalense inhibited tetradecanoyl phorbol acetate (TPA)-induced edema in mouse ears in our screening for anti-inflammatory components. Here, we report the anti-inflammatory activities of extract fractions and the compounds isolated from the active extract of *G. nepalense.*

2. Materials and Methods

2.1. General experimental procedures

The NMR spectra were obtained on a Bruker AM-400 spectrometer operating at 400 MHz for ¹H-NMR and 100 MHz for ¹³C-NMR, respectively. The spectra of electro spray ionization-mass spectrometry (ESI-MS) were recorded on a Finnigan LCQ Advantage Max ion trap mass spectrometer (Thermo Finnigan, USA). The isolation process was conducted on silica gel (200-300 meshes, Qingdao Marine Chemical, China), Sephadex LH-20 (25-100 μ m, Fluka, Switzerland). Thin layer chromatography (TLC) was carried out on silica gel GF254 plates (0.2 mm thickness, 5 × 10 cm, Qingdao Marine Chemical, China).

2.2. Plant material

G. nepalense Sweet was collected in Songhuaba, Kunming, Yunnan, China. The authentication process was carried out by Dr. Jianying Xiang (Kunming Institute of Botany, Chinese Academic of Sciences). A voucher specimen was deposited in the Kunming Institute of Botany, Chinese Academic of Sciences (Kunming, Yunnan, China).

2.3. Animals

The Kunming mice were purchased from Yunnan Baiyao Group Company Limited (Kunming, Yunnan, China). Animal Ethics Committee (AEC) approvals were obtained for the experimental protocols. The AEC oversees animal programs, facilities and procedures. Mice were housed in a climate-controlled environment with a 12 h light/dark cycle and were provided with free access to food and water during the experiment.

^{*}Address correspondence to:

Dr. Yuemao Shen, School of Pharmaceutical Sciences, Shandong University, No. 44 West Wenhua Road, Ji'nan, Shandong 250012, China.

E-mail: yshen@sdu.edu.cn; yshen1965@gmail.com

2.4. Extraction and fractionation

The dried and cut material (2.0 kg) was soaked in distilled water and boiled for three times. The water solution was combined and concentrated *in vacuo* to about 500 mL. The concentrated water solution was then partitioned with ethyl acetate and *n*-butanol successively. After revaporation under reduced pressure, ethyl acetate fraction (37 g) (marked as GN-EA) and *n*-butanol fraction (33 g) (marked as GN-BU) were obtained respectively. Finally, the left water fraction was concentrated to dry and marked as GN-W.

2.5. Anti-inflammatory activities of extract fractions of G. nepalense

The anti-inflammatory activities of GN-EA, GN-Bu, and GN-W were evaluated in a TPA-induced mouse ear edema model. The inflammation model was established according to Hu's and Agut's method (4,5).

A total of 75 Kunming mice were allotted to five groups of 15 each in a completely randomized design. The mice of each group were treated by gastric perfusion of none, 0.6 g/kg aspirin, 2.5 g/kg GN-EA, 2.5 g/kg GN-Bu, and 7.5 g/kg GN-W 30 min prior to each TPA (0.05 mL) treatment once a day for 3 days.

The mice were sacrificed 1 h after the last TPA treatment. Ear punches (7 mm diameter) were taken from each group and weighted. The t test with different samples was adopted for comparison between groups.

2.6. Isolation of GN-EA fraction

The ethyl acetate fraction (32 g) was subjected to column chromatography (CC) over silica gel eluted with a solvent system of CHCl₃/MeOH in gradient (100:1; 100:5; 100:10, and MeOH) to obtain 9 subfractions (Fr. 1-9) based on the TLC analysis. Fr. 4 (606 mg) was subjected to CC over silica gel (15 g) and eluted with petroleum ether/acetone (10:1; 5:1; 3:1) to obtain Fr. 4a (25 mg) and further purified by CC over Sephadex LH-20 eluted with MeOH to yield 1 (8 mg). Fr. 8 was purified by repeated CC over Sephadex LH-20 eluted with MeOH to yield 2 (90.5 mg) and 3 (24.0 mg). Fr. 5 (900 mg) was subjected to CC over Sephadex LH-20 eluted with MeOH to obtained 4 (670 mg). Fr. 6 was purified by CC over Sephadex LH-20 eluted with MeOH to yield 5 (700 mg). Fr. 1 (400 mg) was subjected to CC over silica gel (15 g) and eluted with petroleum ether/acetone (10:1; 5:1) to obtain Fr. 1a (25 mg). Fr. 1a was further purified by CC over Sephadex LH-20 eluted with MeOH to yield 6 (6.5 mg).

3. Results and Discussion

3.1. Anti-inflammatory activities of G. nepalense fractions

Preliminary phytochemical screening and evaluation of

anti-inflammatory components indicated that organic acids, flavonoids, polyphenolic, tannin, and essential oil may be responsible, at least in part, for the antiinflammatory effects of the total extract of *Geranium* (*3*,*6*). Therefore, the TPA-induced ear edema model was employed with the objective of seeking the major bioactive fraction from *G. nepalense*. Both GN-EA and GN-Bu fractions exhibited significant (p < 0.01) antiinflammatory activities on TPA-induced ear edema model at 2.5 g/kg in our study (Table 1).

3.2. Chemical structures of the isolated compounds

The ethyl acetate extract of *G. nepalense* was isolated by repeated column chromatography (Sephadex LH-20 and silica gel) to afford six pure compounds. These compounds were subjected to ¹H-NMR, ¹³C-NMR, and ESI-MS analyses for structure identification (Data are shown in the Appendix). They are elucidated to be kaempferol (1) (7-9), kaempferol-7-O- β -Dglucopyranoside (2) (10-12), quercetin-7-O- α rhamnopyranoside (3) (13), pyrogallol (4) (14), gallic acid (5) (15), and epipinoresinol (6) (16) according to their ¹H- and ¹³C-NMR spectral data and compared with spectral values in literatures. This is the first report for the isolation of compounds **1-6** (Figure 1) from this plant.

 Table 1. Anti-inflammatory activity of GN fractions of G.

 nepalense

| Treatment | Rats (individuals) | Dose (g/kg) | Weight of ear edema (mg) |
|-----------|--------------------|-------------|--------------------------|
| Model | 15 | 0.0 | 7.33 ± 3.04 |
| Aspirin | 15 | 0.6 | $2.84 \pm 1.94^{**}$ |
| GN-EA | 15 | 2.5 | $2.48 \pm 2.02^{**}$ |
| GN-Bu | 15 | 2.5 | $3.36 \pm 1.76^{**}$ |
| GN-W | 15 | 7.5 | $5.62 \pm 2.86^{\#}$ |

All values are expressed as mean of 15 mice in each group. Statistically significant: ** p < 0.01 compared to control, # compared to GN-EA.

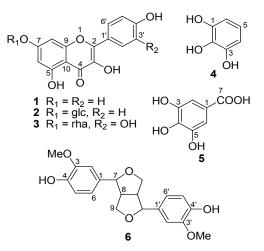


Figure 1. The chemical structures of compounds 1-6.

Previously studies have suggested that the ethyl acetate extract of G. carolinianum was the antiinflammatory active fraction (17). The polyphenolic compounds of Geranium spp. such as flavonoids and tannins have been shown to possess free radical scavenging/antioxidant anti-inflammatory activity both in vivo and in vitro (12,18,19,20). The anti-inflammatory effect of G. nepalense was evaluated by TPA-induced ear edema model in vivo in this study. The results showed that both GN-EA and GN-Bu fractions inhibited TPAinduced inflammation. Six compounds were isolated from the GN-EA fraction. The chemical structures of them were elucidated to be flavonoids (1-3), pyrogallol (4), gallic acid (5), and epipinoresinol (6), which all belong to polyphenolic. 1 isolated from Hibiscus cannabinus L. showed significant anti-inflammatory (21). **3** and **6** were also isolated from the leaves of Brasenia schreberi, and both compounds exhibited anti-inflammatory activities (22). Our results suggested that highly-enriched 4 and 5 may be the best active constituents related to the traditional utilization of this herb.

Acknowledgements

This work was partially supported by the National Science Fund for Distinguished Young Scholars to Y.-M. Shen (30325044) and the Key Project of Chinese Ministry of Education (306010).

References

- http://www.efloras.org/florataxon.aspx?flora_ id=2&taxon_id=200012393 (accessed February 24, 2012).
- The flora of China || GERANIACEAE || Sect. Sibirica R. Knuth in Engl. Pflanzenr. Heft || Geranium nepalense Sweet 43(1). http://www.plants.csdb.cn/eflora/View/ Search/Chs_contents.aspx?L_name=Geranium%20 nepalense%20Sweet (accessed February 24, 2012).
- 3. Zhou HY. The basic study of wilford granesbill herb. Guowai Yiyao: Zhiwuyao Fen'ce. 1996; 11:164-166.
- Hu XQ. Studies on analgesia liniment of muskiness. Zhong Cheng Yao. 1990; 12:26-27.
- Agut J, Tarrida N, Sacristan A. Antiinflammatory activity of topically applied sertaconazole nitrate. Methods Find Exp Clin Pharmacol. 1996; 18:233-234.
- Du SS, Zhang WS, Wu C, Xu YC, Wei LX. Chemical Constituents of *Geraniun eristemon*. Zhongguo Zhong Yao Za Zhi. 2003; 28:625-626.
- Adebayo AH, Tan NH, Akindahunsi AA, Zeng GZ, Zhang YM. Anticancer and antiradical scavenging activity of *Ageratum conyzoides* L. (Asteraceae). Pharmacogn Mag. 2010; 6:62-66.
- Thakur A, Jain V, Hingorani L, Laddha KS. Phytochemical studies on *Cissus quadrangularis* Linn. Pharmacogn Res. 2009; 1:213-215.
- Wan C, Yu Y, Zhou S, Tian S, Cao S. Isolation and identification of phenolic compounds from *Gynura divaricata* leaves. Pharmacogn Mag. 2011; 7:101-108.
- 10. Ebada S, Ayoub N, Singab A, AI-Azizi M. Phytophenolics from *Peltophorum africanum* Sond. (Fabaceae) with

promising hepatoprotective activity. Pharmacogn Mag. 2008; 4:287-293.

- Fiorentino A, D'Abrosca B, Pacifico S, Golino A, Mastellone C, Oriano P, Monaco P. Reactive oxygen species scavenging activity of flavone glycosides from *Melilotus neapolitana*. Molecules. 2007; 12:263-270.
- Sang S, Cheng X, Zhu N, Stark RE, Badmaev V, Ghai G, Rosen RT, Ho CT. Flavonol glycosides and novel iridoid glycoside from the leaves of *Morinda citrifolia*. J Agric Food Chem. 2001; 49:4478-4481.
- Wei Y, Xie Q, Dong W, Ito Y. Separation of epigallocatechin and flavonoids from *Hypericum perforatum* L. by high-speed counter-current chromatography and preparative high-performance liquid chromatography. J Chromatogr A. 2009; 1216:4313-4318.
- Meselhy MR, Nakamura N, Hattori M. Biotransformation of (-)-epicatechin 3-O-gallate by human intestinal bacteria. Chem Pharm Bull (Tokyo). 1997; 45:888-893.
- 15. Zhang WJ, Liu YQ, Li XC, Yang CR. Chemical constituents of ecological tea from Yunnan. Acta Bot Yunnanica. 1995; 17:204-208.
- Phenylpropanoids (Zhan YM, ed.). Chemical Industry Press, Beijing, China, 2005.
- 17. Hu YQ, Liu DL, Zhou YC, Lei ZY. Studies on the anti-inflammatory and analgesic activity of *Geranium carolinianum*. Xibei Yaoxue Zazhi. 2003; 18:113-115.
- Kuo YJ, Hwang SY, Wu MD, Liao CC, Liang YH, Kuo YH, Ho HO. Cytotoxic constituents from *Podocarpus fasciculus*. Chem Pharm Bull (Tokyo). 2008; 56:585-588.
- Saija A, Scalese M, Lanza M, Marzullo D, Bonina F, Castelli F. Flavonoids as antioxidant agents: Importance of their interaction with biomembranes. Free Radic Biol Med. 1995; 19:481-486.
- Latte KP, Kolodziej H. Antioxidant properties of phenolic compounds from *Pelargonium reniforme*. J Agric Food Chem. 2004; 52:4899-4902.
- Rho HS, Ghimeray AK, Yoo DS, Ahn SM, Kwon SS, Lee KH, Cho DH, Cho JY. Kaempferol and kaempferol rhamnosides with depigmenting and anti-inflammatory properties. Molecules. 2011; 18:3338-3344.
- Legault J, Perron T, Mshvildadze V, Girard-Lalancette K, Perron S, Laprise C, Sirois P, Pichette A. Antioxidant and anti-inflammatory activities of quercetin 7-*O*-β-Dglucopyranoside from the leaves of *Brasenia schreberi*. J Med Food. 2011; 14:1127-1134.

(Received February 24, 2012; Revised August 8, 2012; Accepted August 9, 2012)

Appendix

¹H-NMR, ¹³C-NMR, and MS analyses of the isolated compounds 1-6.

Kaempferol (1), yellow powder, $C_{15}H_{10}O_6$, ESI-MS: *m/z* 285 [M – H]⁻. ¹H-NMR (400 Hz, DMSO-*d*₆) δ : 7.98 (d, *J* = 8.8, H-2', 6', 2H), 6.96 (d, *J* = 8.8, H-3', 5', 2H), 6.74 (d, *J* = 2.0, H-8), 6.36 (d, *J* = 2.0, H-6). ¹³C-NMR (100 Hz, DMSO-*d*₆) δ : 150.3 (C-2), 138.7 (C-3), 178.4 (C-4), 161.4 (C-5), 92.3 (C-6), 160.2 (C-7), 97.7 (C-8), 160.9

(C-9), 105.2 (C-10), 130.5 (C-1'), 130.2 (C-2', 6'), 115.6 (C-3', 5'), 155.3 (C-4').

Kaempferol-7-*O*-β-*D*-glucopyranoside (2), $C_{21}H_{20}O_{11}$, ESI-MS: *m/z* 447 [M – H]⁻. ¹H-NMR (400 Hz, CD₃OD) δ: 8.25 (d, *J* = 8.8, H-2', 6', 2H), 7.73 (d, *J* = 8.8, H-3', 5', 2H), 6.25 (d, *J* = 2.1, H-8), 6.19 (d, *J* = 2.1, H-6), 5.82 (d, *J* = 9.7, glc-H-1). ¹³C-NMR (100 Hz, CD₃OD) δ: 146.8 (C-2), 138.7 (C-3), 176.4 (C-4), 161.4 (C-5), 98.7 (C-6), 164.2 (C-7), 93.7 (C-8), 157.1 (C-9), 104.6 (C-10), 132.0 (C-1'), 128.6 (C-2', 6'), 115.4 (C-3', 5'), 155.6 (C-4'), 103.8 (glc-C-1), 75.0 (glc-C-2), 78.6 (glc-C-3), 71.3 (glc-C-4), 79.0 (glc-C-5), 62.4 (glc-C-6).

Quercetin-7-*O*-α*-rhamnopyranoside* (3), C₂₁H₂₀O₁₁, ESI-MS: *m/z* 447 [M – H]⁻; ¹H-NMR (400 MHz, CD₃OD) δ: 7.33 (d, *J* = 1.5 Hz, H-2'), 6.90 (d, *J* = 8.3 Hz, H-5'), 7.29 (dd, *J* = 1.5, 8.3 Hz, H-6'), 6.19 (br. s, H-6), 6.35 (br. s, H-8), 5.34 (br. s, rha-H-1), 0.93 (d, *J* = 6.1 Hz, rha-H-6); ¹³C-NMR (100 MHz, CD₃OD) δ: 148.9 (C-2), 136.2 (C-3), 179.6 (C-4), 158.5 (C-5), 99.8 (C-6), 165.8 (C-7), 94.7 (C-8), 158.2 (C-9), 105.9 (C-10), 133.0 (C-1'), 116.4 (C-2'), 146.4 (C-3',), 147.8 (C-4'), 117.0 (C-5'), 123.9 (C- 6'), 103.5 (rha-C-1), 71.9 (rha-C-2), 72.0 (rha-C-3), 72.1 (rha-C-4), 73.3 (rha-C-5), 17.6 (rha-C-6).

Pyrogallol (4), brown needle, C₆H₆O₃, EI-MS *m/z* (%): 126 (100, M⁺); ¹H-NMR (400 MHz, DMSO-*d*₆) δ: 6.79 (t, J = 8.0, H-5), 6.63 (d, J = 8.0, H-4, 6, 2H); ¹³C-NMR (100 MHz, DMSO-*d*₆) δ: 110.7 (C-4, C-6), 120.7 (C-5), 140.0 (C-2), 147.8 (C-1, 3).

Gallic acid (5), colorless needles, $C_7H_6O_5$, EI-MS *m/z* (%): 170 (100, M⁺), 153 (80), 126 (92); ¹H-NMR (400 MHz, C_5D_5N) δ : 8.08 (s, H-2, H-6); ¹³C-NMR (100 MHz, C_5D_5N) δ : 123.0 (C-1), 110.7 (C-2, C-6), 147.8 (C-3, C-5), 140.7 (C-4), 169.9 (C-7).

Epipinoresinol (6), white powder, $C_{20}H_{22}O_6$, ESI-MS: *m/z* 357 [M – H]⁻; ¹H-NMR (400 MHz, C_5D_5N) δ : 4.94 (d, *J* = 3.3, H-7, 7', 2H), 3.23 (m, H-8, 8', 2H), 4.31 (m, H-9a, 9'a, 2H), 4.00 (m, H-9b, 9'b, 2H), 3.76 (s, C-3, 3'-OMe, 6H); ¹³C-NMR (100 MHz, C_5D_5N) δ : 133.2 (C-1, 1'), 111.0 (C-2, 2'), 148.9 (C-3, 3'), 147.9 (C-4, 4'), 116.5 (C-5, 5'), 119.8 (C-6, 6'), 86.5 (C-7, 7'), 54.8 (C-8, 8'), 72.0 (C-9, 9'), 58.2 (C-3, 3'-OMe).

Original Article

Synthesis and structure-activity relationship study of 2-(substituted benzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2*H*-thiazolo[3,2-*a*]pyrimidin-3(7*H*)-one derivatives as anticancer agents

Theivendran Panneer Selvam^{1,*}, Viswanathen Karthick², Palanirajan Vijayaraj Kumar³, Mohamed Ashraf Ali⁴

¹ Department of Pharmaceutical Chemistry, PES's Rajaram and TarabaiBandekar College of Pharmacy, Ponda, Goa, India;

² Department of Biotechnology, Acharya Nagarjuna University, Guntur, Andrapradesh, India;

³ School of Pharmacy, UCSI (University College Sadaya International) University, Kuala Lumpur, Selangor, Malaysia;

⁴ Institute for Research in Molecular Medicine (INFORM), University of Sains Malaysia, Penang, Malaysia.

ABSTRACT: The synthesis and structure-activity relationship (SAR) study of a series of 2-(substituted benzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2Hthiazolo[3,2-a]pyrimidin-3(7H)-one (4a-4j) derivatives as anticancer agents are described. This series of thiazolopyrimidines were synthesized by the reaction of 7-(4-fluoro phenyl)-5-(furan-2-yl)-2H-thiazolo[3,2-a] pyrimidin-3(7H)-one (3) with appropriate substituted aldehydes in the presence of anhydrous sodium acetate and glacial acetic acid. Their structures were confirmed by IR, ¹H-NMR, mass, and elemental analyses. These novel thiazolopyrimidine derivatives were screened for their anticancer activity on the U937 human histocytic lymphoma cell line by 3-(4,5-dimethyl thiazole-2-yl)-2,5-diphenyltetrazoliumbromide (MTT) assay. The comparison of anticancer activity of thiazolopyrimidine was performed considering their structures. This study was done using 2-(substituted benzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2Hthiazolo[3,2-a]pyrimidin-3(7H)-one (4a-4j) as a basic model, showing that i) presence of a hydrogen donor/ acceptor domain [thiazolo[3,2-a]pyrimidin-3(7H)one] on the thiazolopyrimidine ring; ii) presence of a hydrophobic [(4-fluorophenyl)] aryl ring system on the thiazolopyrimidine ring; iii) presence of an electron donor moiety [5-(furan-2-yl)] on the thiazolopyrimidine ring; iv) ortho and para substitution of the distal aryl ring [2-(substituted benzylidene)] function strongly influenced anticancer activity. Among these compounds (4a-4j) para substituted derivatives 4c, 4e, 4f, 4g, 4h, and 4j showed significant anticancer activity.

Keywords: Thiazolopyrimidine, benzylidene aryl ring, anticancer activity

1. Introduction

Thiazole, pyrimidine and related pyrimidines are classes of fused heterocycles that are of considerable interest because of the diverse range of their biological properties. These are among a wide variety of nitrogen heterocycles that have been explored for developing pharmaceutically important molecules. Thiazolopyrimidine and related fused heterocycles are of interest as potential bioactive molecules, which can be considered as thia-analogues of the natural purine bases such as adenine and guanine, and have acquired a growing importance in the field of medicinal chemistry because of their biological potential. They are known to exhibit pharmacological activities such as analgesic, antiinflammatory, antiarrhythmic, antiparkinsonian, and anticancer activities (*1-8*).

Cancer is a collection of different life threatening diseases characterized by uncontrolled growth of cells leading to invasion of surrounding tissue and often spreading to other parts of the body. When it comes to understanding and controlling cancer scientists are now working from a position of strength because a foundation of knowledge about cancer has been built over the past 50 years. There is an urgent need for novel effective drug regimens for the treatment of cancer because the current chemotherapy suffers from a slim therapeutic index, with significant toxicity from effective drug doses or tumor recurrence at low drug doses. The new anticancer chemotherapeutic agents search continues to be an active area of research at many companies and research centers (9,10). Searching for new anticancer agents having heterocyclic nucleus continues worldwide at various laboratories (11-13).

In the last several decades, fused pyrimidine

^{*}*Address correspondence to:*

Dr. T. Panneer Selvam, Department of Pharmaceutical Chemistry, PES's Rajaram and TarabaiBandekar College of Pharmacy, Faramagudi, Ponda- 403 401, Goa, India. E-mail: tpsphc@gmail.com

derivatives are a class of heterocyclic compounds that have attracted significant interest in medicinal chemistry because they have a wide range of pharmaceutical and pharmacological applications including potential anti-tumor, antimycobacterial, and antiviral activities. Moreover, in recent years, it was reported that many fused pyrimidine analogues were reported to be inhibitors of tyrosine kinase and cyclin-dependent kinases, which are involved in mediating the transmission of mitogenic signals and numerous other cellular events (14-19), including, cell proliferation, migration, differentiation, metabolism, and immune responses. It was also found that many of these derivatives may block proliferation of various cancer cell lines (20).

Led by the above facts on pyrimidine chemistry, we have synthesized new 2-(substituted benzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2*H*-thiazolo[3,2-*a*]pyrimidin-3(7*H*)-one derivatives (**4a-4j**). The main objective of the present investigation is a modified experimental approach to evaluate position of substitutions in synthesized compounds and validation with reference drugs such as gefitinib. The introduction of the substituted benzylidene at the 2nd position of the thiazolopyrimidine scaffold led to significant anticancer activity.

2. Materials and Methods

2.1. Chemicals and reagents

The chemicals and reagents used were obtained from various chemical units including Aldrich Co. (Powai, Mumbai, India), E. Merck India Ltd. (Ponda, Goa, India), CDH (Daryaganj, New Delhi, India), and SD Fine Chem (Worli Road, Mumbai, India). These solvents used were of laboratory research (LR) grade and purified before their use. The silica gel G used for analytical chromatography (TLC) was obtained from E. Merck India Ltd. Melting points were measured in open capillary tubes on a Boetius apparatus (Carl Zeiss Jena) and are uncorrected. ¹H-NMR spectra were taken on a 300 MHz NMR spectrometer (model Ultra Shield, Bruker, Rheinstetten, Germany) in $(d_6$ -DMSO) using tetramethylsilane [(CH₃)₄Si] as internal standard. Chemical shifts (δ) are expressed in ppm. Mass spectra were obtained on an instrument (JEOL-SX-102, Japan) using electron impact ionization. IR spectra were recorded in KBr pellets on a Fourier-transform infrared spectrometer (FT-IR 410, Jasco Corporation, Tokyo, Japan). Elemental analyses were performed on an elemental analyzer (Model 240c, Perkin Elmer, Thane, Maharashtra, India) and were within $\pm 0.4\%$ of the theoretical values.

2.2. General procedure for the synthesis of title compounds (4a-4j)

2.2.1. Preparation of 3-(4-fluorophenyl)-1-(furan-2yl)prop-2-en-1-one (1) The key intermediates were synthesized by a previously reported method (21). 3-(4-Fluorophenyl)-1-(furan-2-yl) prop-2-en-1-one (1) prepared by the mixture of KOH (0.055 mol), water (20 mL), ethanol (15 mL), 2-acetyl furan (0.043 mol), and p-fluorobenzaldehyde (0.043 mol) was stirred at 30-40°C for 2 h and kept overnight. It was then filtered, washed with water and with ethanol, dried and refluxed with glacial acetic acid (10 mL) for 2 h. The crystals separated after cooling were filtered and washed with water, dried and used in further reactions. Yield 79%, Mp 212°C; IR (KBr) cm⁻¹: 2,991 (Ar-CH_{str}), 1,733 (C=O), 1,631 (C=C), 1,030 (cyclic C-O-C_{str}), 823 (C-F); ¹H-NMR (300 MHz, DMSO-*d*₆, бррт): 7.51 (d, J = 8.2 Hz, 2H, ArH), 7.23 (dd, J_1 = 7.7 Hz, J_2 = 1.83 Hz, 2H, ArH), 6.53-7.21 (m, 3H, -CH-furan), 6.10-7.14 (d, 2H, =CH); MS (EI) m/z 216 $[M]^+$; Anal. Calcd. for C₁₃H₉FO₂: C, 72.22; H, 4.20; Found: C, 72.23; H, 4.22.

2.2.2. 4-(4-Fluorophenyl)-6-(furan-2-yl)-3,4-dihydropyrimidin-2(1H)-thione (2)

A mixture of 3-(4-fluorophenyl)-1-(furan-2-yl)prop-2-en-1-one (1) (0.039 mol) thiourea (0.03 mol) and potassium hydroxide (2.5 g) in 95% ethanol (100 mL) was heated under reflux for 3 h. The reaction mixture was concentrated to half of its volume, diluted with water, then acidified with dilute acetic acid and kept overnight. The solid thus obtained, was filtered, washed with water and recrystallized from ethanol to give 4-(4-fluorophenyl)-6-(furan-2-yl)-3,4-dihydropyrimidine-2(1H)-thione (2). Yield 72%, Mp 231°C; IR (KBr) cm⁻¹: 3,361 (NH_{str}), 3,021 (Ar-CH_{str}), 1,531 (C=C), 1,034 (cyclic C-O-C_{str}), 843 (C-F); ¹H-NMR (300 MHz, DMSO- d_{62} δ ppm): 7.39 (dd, $J_{1} = 6.4$ Hz, J₂ = 1.8 Hz, 2H, ArH), 7.41 (d, J = 8.2 Hz, 2H, ArH), 6.61-6.81 (m, 3H, –CH-furan), 6.14 (dd, $J_1 = 8.4$ Hz, $J_2 = 2.0$ Hz, 1H, pyrimidine H), 4.70 (dd, $J_1 = 12.0$ Hz, $J_2 = 2.0$ Hz, 1H, pyrimidine H), 3.27 (s, 2H, -NH); MS (EI) m/z 274 $[M]^+$; Anal. Calcd. for $C_{14}H_{11}FN_2OS$: C, 61.30; H, 4.04; N, 10.21; Found: C, 61.33; H, 4.06; N, 10.24.

2.2.3. 7-(4-Fluorophenyl)-5-(furan-2-yl)-2Hthiazolo[3,2-a]pyrimidin-3(7H)-one (3)

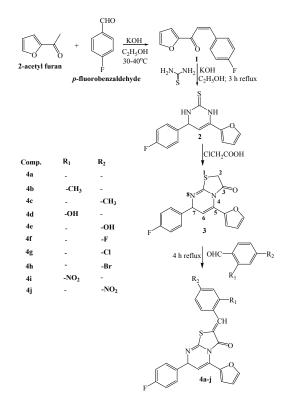
The chloroacetic acid (0.096 mol) was melted on a water bath and (2) (0.009 mol) added to it portion wise to maintain its homogeneity. The homogeneous mixture was further heated on a water bath for 30 min and kept overnight. The solid thus obtained was washed with water and recrystallized from ethanol to give 7-(4-fluorophenyl)-5-(furan-2-yl)-2*H*-thiazolo[3,2-*a*]pyrimidin-3(7*H*)-one (3). Yield 69%, Mp 227°C; IR (KBr) cm⁻¹: 3,354 (NH_{str}), 3,047 (Ar-CH_{str}), 1,721 (C=O), 1,512 (C=C), 1,021 (cyclic C–O– C_{str}), 822 (C-Cl); ¹H-NMR (300 MHz, DMSO-*d*₆, δ ppm): 7.31 (dd, *J*₁ = 7.1 Hz, *J*₂ = 2.2 Hz, 2H, ArH), 7.74 (d, *J* = 7.9 Hz, 2H, ArH), 6.71-7.12 (m, 3H, –CH-furan), 5.32 (dd, *J*₁ = 7.5 Hz, *J*₂ = 1.1 Hz, 1H, thiazolopyrimidine H), 4.13 (dd, *J*₁ 2H, $-CH_2$ thiazole); MS (EI) m/z 314 [M]⁺; Anal. Calcd. for $C_{16}H_{11}FN_2O_2S$: C, 61.14; H, 3.53; N, 8.91; Found: C, 61.12; H, 3.51; N, 8.94.

2.2.4. General procedure for the synthesis of 2-(substitutedbenzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2H-thiazolo[3,2-a]pyrimidin-3(7H)-one (4a-4j)

A mixture of (3) (0.002 mol), substituted benzaldehyde (0.002 mol), and anhydrous sodium acetate (0.002 mol) in 100% glacial acetic acid (10 mL) was heated under reflux for 4 h. The reaction mixture was kept overnight and the solid, thus separated, was filtered, washed with water and recrystallized from ethanol to furnish 2-(substituted benzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2*H*-thiazolo[3,2-*a*]pyrimidin-3(7*H*)-one (**4a-4j**) (Scheme 1).

2.3. Cell proliferation assay

The U937 human histocytic lymphoma cell line was obtained from cell line bank of National Center for Cellular Sciences (NCCS), Pune, India. These cells were maintained in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% fetal bovine serum (FBS) at 37° C, in a CO₂ incubator in the presence or absence of test compounds. The anticancer property of the compounds was measured by MTT assay (22). The cells were plated in a 96-well plate at a density of 5,000 cells/well. After 24 h, cell culture media was replaced with DMEM containing 10% FBS and the cells were treated with different concentrations of the compounds



Scheme 1. Synthetic protocols of target compounds (4a-4j).

(0.01-50 mM). The cells were later incubated for 72 h. Cytotoxicity was measured by adding 5 mg/mL of MTT to each well and incubating for another 3 h. The purple formazan crystals were dissolved by adding 100 μ L of DMSO to each well. The absorbance was read at 570 nm in a spectrophotometer. Cell death was calculated as follows: Cell death percentage = 100 – [test absorbance/ control absorbance] × 100. The test result is expressed as the concentration of a test compound which inhibits the cell growth by 50% (IC₅₀).

3. Results and Discussion

3.1. Chemistry

The chemical structures of the synthesized compounds were confirmed by infrared spectroscopy, proton nuclear magnetic resonance spectroscopy, mass spectrometry, and elemental analysis. The presence of the carbonyl and olefinic group in compound (1) is characterized by the presence of two strong bands in its IR spectrum at 1,733 and 1,631 cm⁻¹. The formations of compound (2) were confirmed by NH stretching, bending peaks in the range of 3,382, 1,621 cm⁻¹, and appearance of a singlet peak 3.27 for two protons in its ¹H-NMR spectra which might be assigned to NH group connecting the pyrimidine. The conversion of thiazolo[3,2-a]pyrimidin-3(7H)-one (3) can be recognized by a strong absorption peak at 1,721cm⁻¹ in IR due to the carbonyl group in the thiazole ring. The title compounds (4a-4j) showed a singlet at δ 7.12, 7.35, 7.32, 7.31, 7.14, 7.22, 7.25, 7.23, 7.19, and 7.32 ppm due to the benzylidine ring proton in ¹H-NMR confirms the formation of (4a-4i) respectively. Further mass spectra confirmed their purity and molecular weight.

3.2. Biological activity

All the selected compounds 2-(substituted benzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2H-thiazolo[3,2-a] pyrimidin-3(7H)-one (4a-4j) were evaluated for cytotoxic properties on the U937 human histocytic lymphoma cell line with gefitinib as a standard positive control. Inhibition of cell-proliferation was measured by MTT assay. The inhibitory potency (IC₅₀) of compounds (4a-4j) are given in Table 1. The fact that a majority of clinically active anticancer drugs possess a nitrogen hetero atomic system with one or two phenyl rings, at least one carbonyl group in their structure and the presence of hydrogen donor/ acceptor unit is noted. In all of the pioneering experiments important core fragments (23) are defined by the presence of a hydrogen donor/acceptor unit (HAD), a hydrophobic domain (A) (aryl ring substituted/unsubstituted) and an electron donor atom (D). These common features were found in the structures of well-established anticancer drugs such as gefitinib, erlotinib, lapatinib, and dasatinib as well as synthesized compounds (Figure 1). In general,

| Table 1. Anticancer study of synthesized compounds (4a-4j) |
|--|
| on U937 human histocytic lymphoma cell line |

| Compounds | R_1 | R_2 | $IC_{50}\left(\mu M\right)\pm SEM^{a}$ |
|-----------|------------------|------------------|--|
| 4a | - | _ | 10.04 ± 0.52 |
| 4b | -CH ₃ | - | 12.07 ± 0.31 |
| 4c | - | -CH ₃ | 5.37 ± 0.12 |
| 4d | -OH | - | 15.12 ± 0.22 |
| 4e | - | -OH | 5.81 ± 0.17 |
| 4f | - | -F | 3.04 ± 0.26 |
| 4g | - | -Cl | 4.04 ± 0.32 |
| 4h | - | -Br | 3.51 ± 0.43 |
| 4i | $-NO_2$ | - | 20.12 ± 0.26 |
| 4j | - | -NO ₂ | 6.42 ± 0.21 |
| Gefitinib | - | - | 1.00 ± 1.00 |

^a Mean of three independent experiments \pm mean standard error.

lipophilicity is one of the most important parameters because it is mainly involved in pharmacokinetic processes such as absorption, distribution, metabolism, excretion, and toxicity (ADMET) and in ligand-target interactions (24). Lipophilicity is the molecular parameter of choice in numerous quantitative structure-activity relationships (QSAR) of different classes of compounds (25). The promising activity of the compounds may be attributed to the substitutions on the hydrophobic domain. These compounds contain methyl, hydroxy, nitro, and halogens at the *para* position of the benzylidene aryl ring. Moreover, observed data showed that the para substituted derivatives exhibited better activity than other ortho and unsubstituted derivatives. Compounds 4f and 4h were found to be most potent and showed IC₅₀ values of 3.04 and 3.51 mM, respectively. When the ortho group was on the benzylidene aryl ring of compounds, we found a decrease in inhibitory activity. For example, compounds 4d and 4i were 3-fold less potent than 4e and 4j. Among the ten compounds synthesized, we found that compounds 4f and 4h showed comparable activity to that of gefitinib. Moreover, p-CH₃, p-OH, and p-NO₂ substituted compounds (4c, 4e, and 4j) had slighter lower activity than (4f-4h). However, the unsubstituted and ortho substituted (o-methyl, o-hydroxyl, and o-nitro) compounds (4a, 4b, 4d, and 4i) exhibited lesser activity. The anticancer activity of test compounds with decreasing order is shown in Figure 2 and is tabulated in Table 1.

3.3. Structure activity relationships (SAR) study

SAR studies give insights into molecular properties causing receptor affinity and selectivity. The promising nature of the compounds may be attributed to the substitutions on the hydrophobic domain (benzylidene aryl ring). These compounds had electron withdrawing and donating groups at the *ortho* and *para* positions of the hydrophobic aryl ring. In general it was observed that the *para* substituted derivatives were more active than the other derivatives. This may be because of the fact that the *para* substituted derivatives fit better into the receptor site.

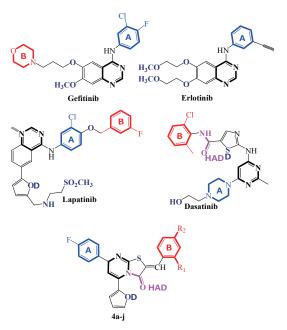


Figure 1. Vital core fragments of wellknown anticancer drugs and synthesized compounds with its important structural features: (HAD) hydrogen bond acceptor/donor domain. (A) hydrophobic aryl ring system, (B) distal aryl ring system, and (D) electron donor moiety.

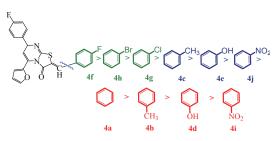


Figure 2. Decreasing order of anticancer activity of thiazolopyrimidine derivatives.

Based on these general concepts we planned to prepare different modifications in title compounds, namely, *ortho* and *para* substitution in the thiazolopyrimidine ring. These modifications showed insight into the dependency of the receptor binding efficacy. Furthermore the *para* substituted thiazolopyrimidine skeletal structure conserved the good receptor binding results. Finally *para* substituted compounds **4c**, **4e**, **4f**, **4g**, **4h**, and **4j** exhibited significant anticancer activity.

4. Conclusions

The literature survey revealed that *para* substitution on the phenyl ring appeared to greatly influence pharmacological activity. This research examined anticancer properties of a novel series of 2-(substituted benzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2H-thiazolo[3,2-*a*]pyrimidin-3(7*H*)-one (**4a-4j**) compounds. Results revealed that para substituted derivatives exhibited better anticancer activity.

Acknowledgements

The authors gratefully acknowledge the chemistry department at PES's Rajaram and Tarabai Bandekar College of Pharmacy for all facilities provided in terms of the use of the available chemicals and equipment. Also, we would like to thank the Central Instrumentation Facility, IIT Chennai, India for the spectral analysis of the compounds used in this study.

References

- Nehad AA, Amr AE, Alhusien AI. Synthesis, reactions, and pharmacological screening of heterocyclic derivatives using nicotinic acid as a natural synthon. Monatsh Chem. 2007; 138:559-567.
- Nehad AA, Nermien MS, Ashraf MM, Abdulla MM. Synthesis, analgesic, and antiparkinsonian profiles of some pyridine, pyrazoline, and thiopyrimidine derivatives. Monatsh Chem. 2007; 138:715-724.
- Ahmed FAS, Abdulla MM, Amr AE, Azza AH. Synthesis, reactions, and antiarrhythmic activity of substituted heterocyclic systems using 5-chloroanisic acid as starting material. Monatsh Chem. 2007; 138:1019-1027.
- Hammam AG, Fahmy AFM, Amr AE, Mohamed AM. Synthesis of novel tricyclic heterocyclic compounds as potential anticancer agents using chromanone and thiochromanoneas synthons. Ind J Chem. 2003; 42B:1985-1993.
- Amr AE, Abdulla MM. Synthesis and pharmacological screening of some new pyrimidine and cyclohexenone fused steroidal derivatives. Ind J Heterocycl Chem. 2002; 12:129-134.
- Amr AE, Ashraf MM, Salwa FM, Nagla AA, Hammam AG. Anticancer activities of some newly synthesized pyridine, pyrane, and pyrimidine derivatives. Bioorg Med Chem. 2006; 14:5481-5488.
- Amr AE, Hegab MI, Ibrahim AA, Abdalah MM. Synthesis and reactions of some fused oxazinone, pyrimidinone, thiopyrimidinone, and triazinone derivatives with a thiophene ring as analgesic, anticonvulsant, and antiparkinsonian agents. Monatsh Chem. 2003; 134:1395-1409.
- Amr AE, Nermien MS, Abdulla MM. Synthesis, reactions, and anti-inflammatory activity of heterocyclic systems fused to a thiophene moiety using citrazinic acid as synthon. Monatsh Chem. 2007; 138:699-707.
- Bridges AJ. Chemical inhibitors of protein kinases. Chem Rev. 2001; 101:2541-2572.
- Wang JD, Miller K, Boschelli DH, Ye F, Wu B, Floyd MB, Powell DW, Wissner A, Weber JM, Boschelli F. Inhibitors of Src tyrosine kinase: The preparation and structureactivity relationship of 4-anilino-3-cyanoquinolines and 4-anilinoquinazolines. Bioorg Med Chem Lett. 2000; 10:2477-2480.
- 11. Cai SX. Small molecule vascular disrupting agents: Potential new drugs for cancer treatment. Recent Pat Anticancer Drug Discov. 2007; 2:79-101.
- Shi LM, Fan Y, Lee JK, Waltham M, Andrews DT, Scherf U, Paull KD, Weinstein JN. Mining and visualizing large anticancer drug discovery databases. J Chem Inf Comput Sci. 2000; 40:367-379.
- 13. Monks A, Scudiero D, Skehan P, Shoemaker R, Paull

K, Vistica D, Hose C. Langley J, Cronise P, Wolff VA, Goodrich MG, Campbell H, Mayo J, Boyd MJ. Feasibility of a high-flux anticancer drug screen using a diverse panel of cultured human tumor cell lines. Natl Cancer Inst.1991; 83:757-766.

- Kim DC, Lee YR, Yang B, Shin KJ, Kim DJ, Chung BY, Yoo KH. Synthesis and cyclin-dependent kinase 2 inhibitors. Eur J Med Chem. 2003; 38:525-532.
- 15. Shenone S, Bruno O, Ranise A, Bondavalli F, Fossa P, Mosti L, Menozzi G, Carraro F, Naldini A, Bernini C, Manetti F, Botta M. New pyrazolo[3,4-d]pyrimidines endowed with A431 antiproliferative activity and inhibitory properties of Src phosphorylation. Bioorg Med Chem Lett. 2004; 14:2511-2517.
- Morisi R, Celano M, Tosi E, Schenone S, Navarra M, Ferretti E, Costante G, Durante C, Botta G, D'Agostino M, Brullo C, Filetti S, Botta M, Russo D. Growth inhibition of medullary thyroid carcinoma cells by pyrazolopyrimidine derivates. J Endocrinol Invest. 2007; 30:RC31-RC34.
- Angelucci A, Schenone S, Gravina GL, Muzi P, Festuccia C, Vicentini C, Botta M, Bologna M. Pyrazolo[3,4d]pyrimidines c-Src inhibitors reduce epidermal growth factor-induced migration in prostate cancer cells. Eur J Cancer. 2006; 42:2838-2845.
- Farley ME, Hoffman WF, Rubino RS, Hambaugh SR, Arrington KL, Hungate RW, Bilodeau MT, Tebben AJ, Rutledge RZ, Kendall RL, McFall RC, Huckle WR, Coll KE, Thomas KA. Synthesis and initial SAR studies of 3,6-disubstituted pyrazolo[1,5-a]pyrimidines: A new class of KDR kinase inhibitors. Bioorg Med Chem Lett. 2002; 12:2767-2770.
- Farley ME, Rubino RS, Hoffman WF, Hambaugh SR, Arrington KL, Hungate RW, Bilodeau MT, Tebben AJ, Rutledge RZ, Kendall RL, McFall RC, Huckle WR, Coll KE, Thomas KA. Optimization of a pyrazolo[1,5a]pyrimidine class of KDR kinase inhibitors: Improvements in physical properties enhance cellular activity and pharmacokinetics. Bioorg Med Chem Lett. 2002; 12:3537-3541.
- Krystof V, Moravcova D, Paprskarova M, Barbier P, Peyrot V, Hlobikova A, Havlicek L, Stramd M. Synthesis and biological activity of 8-azapurine and pyrazolo[4,3d]pyrimidine analogues of myoseverin. Eur J Med Chem. 2006; 41:1405-1411.
- AL-Hazimi HMA, Al-Alshaikh MA. Microwave assisted synthesis of substituted furan-2-carboxaldehydes and their reactions. Journal of Saudi Chemical Society. 2010; 14:373-382.
- Mossman T. Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. J Immunol Methods. 1983; 65:55-63.
- Zhang JM, Yang PL, Gray NS. Targeting cancer with small molecule kinase inhibitors. Nature. 2009; 9:28-39.
- Efremov RG, Chugunov AO, Pyrkov TV, Priestle JP, Arseniev AS, Jacoby E. Molecular lipophilicity in protein modeling and drug design. Curr Med Chem. 2007; 14:393-415.
- Bajda M, Boryczka S, Malawsk, B. Investigation of lipophilicity of anticancer-active thioquinoline derivatives. Biomed Chromatogr. 2007; 21:123-131.

(Received June 13, 2012; Revised July 21, 2012; Re-revised August 2, 2012; Accepted August 3, 2012)

Appendix

Spectral data of the synthesized compounds

2-Benzylidene-7-(4-fluorophenyl)-5-(furan-2-yl)-2Hthiazolo[3,2-a]pyrimidin-3(7H)-one (4a)

Yield 73%, Mp 284°C; IR (KBr) cm⁻¹: 3,061 (Ar-CH_{str}), 1,642 (C=O), 1,514 (C=C benzylidine), 1,032 (cyclic C–O–C_{str}), 811 (C-F); ¹H-NMR (300 MHz, DMSO- d_6 , δ ppm): 7.41 (dd, J_1 = 7.3 Hz, J_2 = 1.8 Hz 2H, ArH), 7.53 (d, J = 6.2 Hz, 2H, ArH), 7.56-7.87 (m, 5H, Ar-H), 7.12 (s, 1H, –CH benzylidine), 6.51-6.89 (m, 3H, –CH-furan), 5.16 (dd, J_1 = 8.1 Hz, J_2 = 2.2 Hz 1H, thiazolopyrimidine H), 3.21 (dd, J_1 = 17.6 Hz, J_2 = 1.8 Hz 1H, thiazolopyrimidine H); MS (EI) m/z 402 [M]⁺; Anal. Calcd. for C₂₃H₁₅FN₂O₂S: C, 68.64; H, 3.76; N, 6.96; Found: C, 68.69; H, 3.73; N, 6.92.

2-(2-Methylbenzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2H-thiazolo[3,2-a]pyrimidin-3(7H)-one (**4b**)

Yield 76%, Mp 291°C; IR (KBr) cm⁻¹: 3,047 (Ar-CH_{str}), 1,623 (C=O), 1,521 (C=C), 1,031 (cyclic C–O–C_{str}), 837 (C-F); ¹H-NMR (300 MHz, DMSO- d_6 , δ ppm): 7.24 (dd, $J_1 = 6.4$ Hz, $J_2 = 1.4$ Hz 2H, ArH), 7.24 (d, J = 8.2 Hz, 2H, ArH), 7.31-7.54 (m, 4H, Ar-H), 7.35 (s, 1H, –CH benzylidine), 6.14-6.66 (m, 3H, –CH-furan), 5.43 (dd, $J_1 = 8.6$ Hz, $J_2 = 2.9$ Hz 1H, thiazolopyrimidine H), 3.24 (dd, $J_1 = 11.6$ Hz, $J_2 = 1.6$ Hz 1H, thiazolopyrimidine H), 2.71 (m, 3H, –CH₃); MS (EI) m/z 416 [M]⁺; Anal. Calcd. for C₂₄H₁₇FN₂O₂S: C, 69.21; H, 4.11; N, 6.73; Found: C, 69.24; H, 4.13; N, 6.71.

2-(4-Methylbenzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2H-thiazolo[3,2-a]pyrimidin-3(7H)-one (**4c**)

Yield 82%, Mp 297°C; IR (KBr) cm⁻¹: 3,057 (Ar-CH_{str}), 1,632 (C=O), 1,543 (C=C), 1,031 (cyclic C–O–C_{str}), 811 (C-F); ¹H-NMR (300 MHz, DMSO- d_6 , δ ppm): 7.33 (dd, $J_1 = 7.2$ Hz, $J_2 = 1.8$ Hz, 2H, ArH), 7.54 (d, J = 7.7 Hz, 2H, ArH), 7.67-7.71 (m, 4H, Ar-H), 7.32 (s, 1H, –CH benzylidine), 6.23-6.74 (m, 3H, –CH-furan), 5.43 (dd, $J_1 = 8.1$ Hz, $J_2 = 1.7$ Hz, 1H, thiazolopyrimidine H), 3.14 (dd, $J_1 = 13.2$ Hz, $J_2 = 2.2$ Hz, 1H, thiazolopyrimidine H), 2.25 (m, 3H, –CH₃); MS (EI) m/z 416 [M]⁺; Anal. Calcd. for C₂₄H₁₇FN₂O₂S: C, 69.21; H, 4.11; N, 6.73; Found: C, 69.18; H, 4.14; N, 6.75.

2-(2-Hydroxybenzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2H-thiazolo[3,2-a]pyrimidin-3(7H)-one (**4d**)

Yield 82%, Mp 276°C; IR (KBr) cm⁻¹: 3,412 (phenolic OH), 3,043 (Ar-CH_{str}), 1,643 (C=O), 1,512 (C=C), 1,021 (cyclic C–O–C_{str}), 819 (C-F); ¹H-NMR (300 MHz, DMSO- d_6 , δ ppm): 9.93 (s, 1H; Ar-OH), 7.36 (dd, J_1 = 7.6 Hz, J_2 = 2.0 Hz, 2H, ArH), 7.51 (d, J = 7.5 Hz, 2H, ArH),

7.63-7.74 (m, 4H, Ar-H), 7.31 (s, 1H, –CH benzylidine) 6.23-6.65 (m, 3H, –CH-furan), 5.41 (dd, $J_1 = 8.7$ Hz, $J_2 = 1.8$ Hz, 1H, thiazolopyrimidine H), 3.13 (dd, $J_1 = 12.4$ Hz, $J_2 = 1.9$ Hz, 1H, thiazolopyrimidine H); MS (EI) m/z 418 [M]⁺; Anal. Calcd. for C₂₃H₁₅FN₂O₃S: C, 66.02; H, 3.61; N, 6.69; Found: C, 66.06; H, 3.64; N, 6.67.

2-(4-Hydroxybenzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2H-thiazolo[3,2-a]pyrimidin-3(7H)-one (**4e**)

Yield 79%, Mp 294°C; IR (KBr) cm⁻¹: 3,471 (phenolic OH), 3,037 (Ar-CH_{str}), 1,624 (C=O), 1,511 (C=C), 1,037 (cyclic C–O–C_{str}), 817 (C-F); ¹H-NMR (300 MHz, DMSO- d_6 , δ ppm): 9.71 (s, 1H, Ar-OH), 7.16 (dd, J_1 = 7.2 Hz, J_2 = 2.2 Hz, 2H, ArH), 7.26 (d, J = 7.1 Hz, 2H, ArH), 7.35-7.48 (m, 4H, Ar-H), 7.14 (s, 1H, –CH benzylidine), 6.26-6.59 (m, 3H, –CH-furan), 5.36 (dd, J_1 = 8.2 Hz, J_2 = 2.0 Hz, 1H, thiazolopyrimidine H), 3.17 (dd, J_1 = 11.8 Hz, J_2 = 2.0 Hz, 1H, thiazolopyrimidine H); MS (EI) m/z 418 [M]⁺; Anal. Calcd. for C₂₃H₁₅FN₂O₃S: C, 66.02; H, 3.61; N, 6.69; Found: C, 66.06; H, 3.64; N, 6.71.

2-(4-Fluorobenzylidene)-7-(4-fluorophenyl)-5-(furan-2yl)-2H-thiazolo[3,2-a]pyrimidin-3(7H)-one (4**f**)

Yield 73%, Mp 289°C; IR (KBr) cm⁻¹: 3,055 (Ar-CH_{str}), 1,636 (C=O), 1,536 (C=C), 1,035 (cyclic C– O–C_{str}), 822 (C-F), 912 (C-F); ¹H-NMR (300 MHz, DMSO-*d*₆, δ ppm): 7.47 (dd, *J*₁ = 6.8 Hz, *J*₂ = 1.4 Hz 2H, ArH), 7.53 (d, *J* = 7.0 Hz, 2H, ArH), 7.62-7.96 (m, 4H, Ar-H), 7.22 (s, 1H, –CH benzylidine), 6.21-6.76 (m, 3H, –CH-furan), 5.42 (dd, *J*₁ = 7.6 Hz, *J*₂ = 1.2 Hz, 1H, thiazolopyrimidine H), 3.19 (dd, *J*₁ = 12.5 Hz, *J*₂ = 2.4 Hz, 1H, thiazolopyrimidine H); MS (EI) m/z 420 [M]⁺; Anal. Calcd. for C₂₃H₁₄F₂N₂O₂S: C, 65.71; H, 3.36; N, 6.66; Found: C, 65.74; H, 3.37; N, 6.68.

2-(4-Chlorobenzylidene)-7-(4-fluorophenyl)-5-(furan-2-yl)-2H-thiazolo[3,2-a]pyrimidin-3(7H)-one (**4g**)

Yield 75%, Mp 291°C; IR (KBr) cm⁻¹: 3,053 (Ar-CH_{str}), 1,634 (C=O), 1,532 (C=C), 1,031 (cyclic C–O–C_{str}), 834 (C-F); ¹H-NMR (300 MHz, DMSO- d_6 , δ ppm): 7.41 (dd, $J_1 = 7.1$ Hz, $J_2 = 1.8$ Hz, 2H, ArH), 7.56 (d, J = 6.8 Hz, 2H, ArH), 7.61-7.96 (m, 4H, Ar-H), 7.25 (s, 1H, –CH benzylidine), 6.22-6.77 (m, 3H, –CH-furan), 5.42 (dd, J_1 = 7.9 Hz, $J_2 = 1.8$ Hz, 1H, thiazolopyrimidine H), 3.11 (dd, $J_1 = 14.2$ Hz, $J_2 = 2.0$ Hz, 1H, thiazolopyrimidine H); MS (EI) m/z 438 [M+2]; Anal. Calcd. for C₂₃H₁₄ClFN₂O₂S: C, 63.23; H, 3.23; N, 6.41; Found: C, 63.26; H, 3.22; N, 6.44.

2-(4-Bromobenzylidene)-7-(4-fluorophenyl)-5-(furan-2yl)-2H-thiazolo[3,2-a]pyrimidin-3(7H)-one (**4h**)

Yield 72%, Mp 286°C; IR (KBr) cm⁻¹: 3,051 (Ar-CH_{st}), 1,643 (C=O), 1,544 (C=C), 1,041 (cyclic C- O–C_{str}), 851 (C-F), 621 (C-Br); ¹H-NMR (300 MHz, DMSO- d_6 , δ ppm): 7.41 (dd, J_1 = 7.9 Hz, J_2 = 2.0 Hz, 2H, ArH), 7.53 (d, J = 6.3 Hz, 2H, ArH), 7.61-7.94 (m, 4H, Ar-H), 7.23 (s, 1H, –CH benzylidine), 6.26-6.75 (m, 3H, –CH-furan), 5.43 (dd, J_1 = 8.2 Hz, J_2 = 2.0 Hz, 1H, thiazolopyrimidine H), 3.12 (dd, J_1 = 16.1 Hz, J_2 = 2.2 Hz, 1H, thiazolopyrimidine H); MS (EI) m/z 483 [M+2]; Anal. Calcd. for C₂₃H₁₄BrFN₂O₂S: C, 57.39; H, 2.93; N, 5.82; Found: C, 57.35; H, 2.91; N, 5.86.

2-(2-Nitrobenzylidene)-7-(4-fluorophenyl)-5-(furan-2yl)-2H-thiazolo[3,2-a]pyrimidin-3(7H)-one (**4i**)

Yield 79%, Mp 293°C; IR (KBr) cm⁻¹: 3,054 (Ar-CH_{str}), 1,647 (C=O), 1,549 (C=C), 1,047 (cyclic C–O–C_{str}), 842 (C-F); ¹H-NMR (300 MHz, DMSO- d_6 , δ ppm): 7.86 (dd, $J_1 = 8.4$ Hz, $J_2 = 1.8$ Hz, 2H, ArH), 7.91 (d, J = 6.9 Hz, 2H, ArH), 8.02-8.35 (m, 4H, Ar-H), 7.19 (s, 1H, –CH benzylidine), 6.32-6.59 (m, 3H, –CH-furan), 5.36 (dd, J_1 = 8.0 Hz, J_2 = 1.8 Hz, 1H, thiazolopyrimidine H), 3.12 (dd, J_1 = 14.2 Hz, J_2 = 2.0 Hz, 1H, thiazolopyrimidine H); MS (EI) m/z 447 [M]⁺; Anal. Calcd. for C₂₃H₁₄FN₃O₄S: C, 61.74; H, 3.15; N, 9.39; Found: C, 61.77; H, 3.11; N, 9.37.

2-(4-Nitrobenzylidene)-7-(4-fluorophenyl)-5-(furan-2yl)-2H-thiazolo[3,2-a]pyrimidin-3(7H)-one (**4j**)

Yield 81%, Mp 286°C; IR (KBr) cm⁻¹: 3,051 (Ar-CH_{str}), 1,643 (C=O), 1,547 (C=C), 1,041 (cyclic C– O–C_{str}), 887 (C-F); ¹H-NMR (300 MHz, DMSO-*d₆*, δ ppm): 7.81 (dd, *J₁* = 8.0 Hz, *J₂* = 2.0 Hz, 2H, ArH), 7.94 (d, *J* = 6.3 Hz, 2H, ArH), 8.12-8.37 (m, 4H, Ar-H), 7.32 (s, 1H, –CH benzylidine), 6.19-6.52 (m, 3H, –CH-furan), 5.47 (dd, *J₁* = 8.2 Hz, *J₂* = 2.2 Hz, 1H, thiazolopyrimidine H), 3.14 (dd, *J₁* = 12.4 Hz, *J₂* = 1.8 Hz, 1H, thiazolopyrimidine H); MS (EI) m/z 447 [M]⁺; Anal. Calcd. for C₂₃H₁₄FN₃O₄S: C, 61.74; H, 3.15; N, 9.39; Found: C, 61.71; H, 3.17; N, 9.35.

Original Article

Antioxidant and hepatoprotective activity of an ethanol extract of *Syzygium jambos* (L.) leaves

Md. Rafikul Islam, Mst. Shahnaj Parvin, Md. Ekramul Islam^{*}

Department of Pharmacy, University of Rajshahi, Rajshahi, Bangladesh.

ABSTRACT: Herbal medicines have traditionally been used worldwide for the prevention and treatment of liver disease with fewer adverse effects. The leaves of the Syzygium jambos (SJL) plant were chosen and studied for their antioxidant activity in vitro and hepatoprotective activity in vivo. The antioxidant activity of the ethanol extract was examined in vitro using a 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical scavenging assay, reducing capacity, total phenol, total flavonoid content, and total antioxidant capacity. The extract had significant dose-dependent antioxidant activity in all in vitro experiments. IC₅₀ values of SJL and ascorbic acid (standard) were found to be 14.10 and 4.87 µg/mL, respectively, according to a DPPH radical scavenging assay. Hepatoprotective activity of the plant extract was evaluated in a rat model of carbon tetrachloride (CCl₄)-induced liver damage. CCl₄ significantly altered serum marker enzymes, total bilirubin, total protein, and liver weight. The extract caused these values to return to normal in rats with CCl₄-induced liver damage that were given SJL. This indicated the hepatoprotective potential of SJL and was comparable to use of the standard drug silymarin. Thus, the present study revealed that SJL may have antioxidant and hepatoprotective activity.

Keywords: Syzygium jambos, antioxidant, hepatoprotective activity

1. Introduction

Reactive oxygen species (ROS) and reactive nitrogen species (RNS) are produced by normal physiologic processes and fulfill important functions in the body at minute or moderate concentrations. ROS and RNS are necessary for the maturation process of cellular structures and can act as weapons for the host's defense system.

*Address correspondence to:

The importance of ROS production by the immune system is clearly evident in patients with granulomatous disease. These patients have a defective membrane-bound nicotinamide adenine dinucleotide phosphate (NADPH) oxidase system that precludes them from producing the superoxide anion radical (O_2^{-}) , thereby resulting in multiple and persistent infections (1,2). Oxygen radicals have crucial action such as signal transduction, gene transcription, and regulation of synthesis of cyclic guanosine monophosphate (cGMP) in cells (3, 4). Nitric oxide (NO) is a common signaling molecule and participates in virtually every one of the body's cellular and organ functions (5). Optimum amounts of NO produced by endothelial cells are essential for regulating the relaxation and proliferation of vascular smooth muscle cells, leukocyte adhesion, platelet aggregation, angiogenesis, thrombosis, vascular tone, and hemodynamics (5). In addition, NO produced by neurons serves as a neurotransmitter (6).

However, the generation of even slightly larger amounts of these essential compounds during metabolism or in response to environmental pollutants, radiation, chemicals, toxins, deep fried and spicy foods, and physical stress can cause massive physiologic problems by oxidation of bio-molecules (protein, amino acids, lipid, and DNA). Free radicals presumably play a major role in all pathologies. Free radicals are believed to be responsible for more than one hundred conditions like cancer, diabetes, atherosclerosis, arthritis, neuropathy, nephropathy, retinopathy, aging, compromised immunity, and cardiovascular diseases (7-10).

The liver has a central role in transforming and clearing chemicals and is closely related to the gastrointestinal tract, which makes it susceptible to drug toxicity, xenobiotics, and oxidative stress. Dysfunction of this organ results in impairment of energy metabolism and intracellular oxidant stress with excessive formation of ROS. CYP2E1 is a cytochrome P450 isoenzyme produced by the liver that also facilitates oxidative stress and cell injury (11,12). Although Kupffer cells and recruited neutrophils in the liver are part of the host-defense system, these inflammatory cells initiate additional liver injury under certain circumstances, such as when excess free radicals are present (13-16).

Dr. Md. Ekramul Islam, Department of Pharmacy, University of Rajshahi, Rajshahi-6205, Bangladesh. E-mail: ekrams74@yahoo.com.au

Although the body has mechanisms to defend against the damaging properties of ROS (17,18), its capacity to control ROS can be overwhelmed, causing irreversible oxidative damage and various conditions that threaten the liver and other parts of the body. Several synthetic and semi-synthetic chemicals are often used to protect the liver from the detrimental effects of free radicals but they are not sufficiently effective and cause adverse reactions. Modern medicine includes many important bioactive molecules with antioxidant and hepatoprotective properties that were derived from plants. Curcumin and lycopene are used as antioxidants and are respectively obtained from the plants Erythroxylum coca and Lycopersicon esculentum. Andrographis paniculata and Silybum marianum are two important plant sources of andrographolide and silymarin, respectively, that are widely used as hepatoprotective agents (19). Hence, recent research has hastened to identify notable hepatoprotective agents from plant products that will reduce the harmful effects of and problems associated with free radicals while causing minimal adverse reactions. As part of ongoing research, the current study examined the ethanol extract of Syzygium jambos leaves for its use as a hepatoprotective agent to reduce the damage caused by ROS and RNS.

Syzygium jambos (L.), commonly known as rose apple, belongs to the family Myrtaceae and may merely be a shrub but is generally a tree found all over parts of Bangladesh, Pakistan, and India (20,21). It has been used in traditional medicine to treat various ailments. The fruit is regarded as a tonic for the brain and liver. The seeds are used to curb diarrhea, dysentery, and catarrh and help treat diabetes. A decoction of the leaves is applied to sore eyes, it serves as a diuretic and expectorant, and it is used to treat rheumatism. The bark contains 7-12.4% tannin. The leaf extract of Syzygium jambos reportedly has antinociceptive and antimicrobial activity (22,23). No reports have described the plant's hepatoprotective activity. Thus, the present study sought to investigate the leaves of the S. jambos (SJL) for its hepatoprotective activity using a rat model of CCl₄-induced liver damage and different in vitro antioxidation experiments.

2. Materials and Methods

2.1. Chemicals

1,1-diphenyl-2-picrylhydrazyl (DPPH), trichloroacetic acid (TCA), ferric chloride, gallic acid, and quercetin were obtained from Sigma Chemical Co., USA. Ascorbic acid and aluminium chloride were obtained from SD Fine Chem. Ltd., Biosar, India. Ammonium molybdate, methanol, sodium phosphate, concentrated H₂SO₄, Folin-Ciocalteu reagent, sodium carbonate, potassium acetate, mono-sodium phosphate, bi-sodium phosphate, potassium ferricyanide, and trichloro acetic acid were purchased from Merck, Germany.

2.2. Preparation of extract

An ethanolic extract of leaves was used in the present study. Mature leaves were collected in July 2009 from Rajshahi, Bangladesh. The leaves were dried in the shade and pulverized in a mechanical grinder. The powder was extracted with ethanol. The extracted solution was filtered using a clean cloth and then filter paper. The extract was concentrated first in a rotary vacuum evaporator and then in a water bath. The extracted residue was weighed and the percent yield of leaves of *S. jambos* was 9.21% w/w. The extract was then frozen prior to examination of its potential antioxidant and hepatoprotective properties.

2.3. DPPH radical scavenging activity assay

The free radical scavenging capacity of the extract was determined using the stable free radical DPPH (24). The leaf extract was mixed with 95% ethanol to prepare a stock solution (5 mg/mL). DPPH solution (0.004%, w/v) was prepared in 95% ethanol. A freshly prepared DPPH solution (0.004%, w/v) was placed in test tubes and SJL was added followed by serial dilution (1 µg to 500 µg) in every test tube so that the final volume was 3 mL. After 10 min, the absorbance was read at 515 nm using a spectrophotometer (HACH 4000 DR UV-visible spectrophotometer, USA). Ascorbic acid was used as a reference standard and dissolved in distilled water to prepare a stock solution with the same concentration (5 mg/mL). A control sample of the same volume was prepared without any extract and reference ascorbic acid. A solution of 95% ethanol served as a blank. The % scavenging of the DPPH free radical was measured using the following equation:

% Scavenging activity = Absorbance of the control – Absorbance of the test sample/Absorbance of the control \times 100

The inhibition curve was plotted for experiments in triplicate and expressed as the % of mean inhibition \pm standard deviation. IC₅₀ values were obtained by Probit analysis (25). The IC₅₀ value is the concentration of the sample required to inhibit 50% of the radical.

2.4. Total antioxidant capacity assay

The antioxidant activity of the extract was evaluated by the phosphomolybdenum method in accordance with the procedure described by Prieto *et al.* (26). The assay is based on the reduction of Mo(VI) to Mo(V) by the extract/sample and subsequent formation of a green phosphate/Mo(V) complex at an acidic pH. A 0.3 mL extract was combined with 3 mL of reagent solution (0.6 M sulfuric acid, 28 mM sodium phosphate, and 4 mM ammonium molybdate). Test tubes containing the reaction solution were incubated at 95°C for 90 min. The absorbance of the solution was then measured against a blank at 695 nm using a spectrophotometer (HACH 4000 DU UV-visible spectrophotometer) after the solution cooled to room temperature. Ethanol (0.3 mL) in the place of extract/sample served as the blank. Antioxidant activity was expressed as the number of gram equivalents of ascorbic acid.

2.5. Reducing capacity assay

Reducing power of the extract was evaluated using the Oyaizu method (27). Different concentrations of SJL extract (125, 250, 500, and 1,000 µg/mL) in 1 mL of distilled water were mixed with phosphate buffer (2.5 mL, 0.2 M, pH 6.6) and potassium ferricyanide $[K_3Fe(CN)_6]$ (2.5 mL, 1% w/v). The mixture was incubated at 50°C for 20 min. After incubation, 2.5 mL of 10% trichloroacetic acid solution was added to each test tube and the mixture was centrifuged at 3,000 rpm for 10 min. Five mL of the upper layer solution was mixed with 5 mL of distilled water and 1 mL of ferric chloride solution (0.1%, w/v) and the absorbance was measured at 700 nm. The reducing power of the extract was linearly proportional to the concentration of the sample. Ascorbic acid served as the reference standard. Phosphate buffer (pH 6.6) served as the blank solution.

2.6. Determination of total phenol content

Total phenol content in the extract was determined with Folin-Ciocalteu reagent. Extract (200 μ g/mL) was mixed with 400 μ L of the Folin-Ciocalteu reagent and 1.5 mL of 20% sodium carbonate. The mixture was shaken thoroughly and brought to 10 mL using distilled water. The mixture was allowed to stand for 2 h. The absorbance at 765 nm was then determined. The total phenol content in SJL was then determined as the mg of gallic acid equivalent using equations that were obtained from a standard gallic acid graph (28).

2.7. Determination of total flavonoid content

The total flavonoid content was determined using a method previously described by Kumaran and Karunakaran (29). One mL of plant extract in ethanol (200 μ g/mL) was mixed with 1 mL aluminium trichloride in ethanol (20 mg/mL) and a drop of acetic acid and then diluted with ethanol to 25 mL. The absorption at 415 nm was read after 40 min. Blank samples were prepared from 1 mL of plant extract and a drop of acetic acid and then diluted to 25 mL with ethanol. The total flavonoid content was determined using a standard curve for quercetin (12.5-100 μ g/mL) and was expressed as mg of quercetin equivalent (QE/g of extract).

2.8. Hepatoprotective activity

The hepatoprotective activity of SJL extract was

determined using a rat model of carbon tetrachlorideinduced hepatotoxicity. After seven days of acclimatization, rats were divided into four groups consisting of three rats each. Treatment lasted for 8 days. Group I served as the normal control and received only normal saline (1 mL/kg, *i.p.*) for eight days. Group II served as the toxic control and was administered carbon tetrachloride in liquid paraffin (CCl₄:liquid paraffin 1:2; 1 mL/kg, *i.p.*) once every 72 h. Group III served as the positive control and was administered silymarin (25 mg/kg/day, p.o.) along with carbon tetrachloride in liquid paraffin (CCl₄:liquid paraffin 1:2; 1 mL/kg, i.p.) once every 72 h. Group IV was administered SJL (300 mg/kg, p.o.) daily and carbon tetrachloride in liquid paraffin (CCl₄:liquid paraffin 1:2; 1 mL/kg, i.p.) once every 72 h. Twenty-four h after the last dose, blood was taken from the retro-orbital plexus under sodium phenobarbital anesthesia and rats were dissected to remove the liver. Before blood was collected, the syringe was ringed with heparin to prevent hemolysis/clotting. The blood samples were then centrifuged at 2,500 rpm at 37°C to separate serum and were used to estimate the biochemical markers of liver damage, i.e. SGOT, SGPT (30,31), ALP (32), bilirubin (33), and total protein levels (34).

2.9. Statistical analysis

Linear regression analysis was used to calculate IC₅₀ values wherever needed. All results are expressed as average \pm SEM. Data were statistically evaluated in InStat software using one-way analysis of variance (ANOVA) followed by a post hoc Dunnett's test. *p* < 0.05 was considered statistically significant.

3. Results

3.1. DPPH radical scavenging activity

A DPPH assay is one of the most widely used methods of screening for the antioxidant activity of plant extracts. DPPH is a stable, nitrogen-centered free radical that produces a violet color in ethanol solution. When DPPH encounters proton donors such as antioxidants, it is reduced to a yellow product, diphenyl picryl hydrazine, and then absorbance decreases. The antioxidant activity of an ethanol extract of SJL was evaluated by measuring its scavenging capacity for the DPPH free radical; this capacity was expressed using the IC₅₀ value. A lower IC₅₀ value indicates an extract with greater scavenging activity. The extract had dose-dependent activity, i.e. DPPH scavenging activity increased proportionate to the increase in concentration of the extract. Results are shown in Figure 1. The IC₅₀ value of the extract was 14.10 µg/mL while that for reference ascorbic acid was $4.87 \mu g/mL$. These results indicate that the extract had definite free radical scavenging activity in comparison to ascorbic acid.

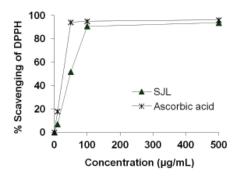


Figure 1. DPPH radical scavenging activity of the ethanolic extract of *S. jambos* leaves. Values are the average of experiments in triplicate and are expressed as mean \pm standard deviation.

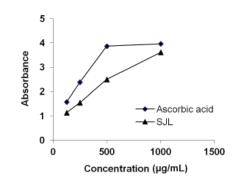


Figure 2. Reducing power of *S. jambos* leaves and ascorbic acid.

Table 1. Effect of ethanol extract of *S. jambos* leaves and silymarin on serum biochemical parameters in CCl₄-induced liver damage in rats

| Tractment group | | Serum biochemical parameters | | | | | |
|------------------|-----------------|------------------------------|------------------|-------------------|-----------------------|------------------|--|
| Treatment group | SGPT (U/L) | SGOT (U/L) | ALP (KA) | Bilirubin (mg/dL) | Total protein (mg/dL) | Liver weight (g) | |
| Normal (control) | 22.1 ± 0.33 | 38.8 ± 0.39 | 19.26 ± 0.04 | 1.06 ± 0.016 | 12.14 ± 0.18 | 5.96 ± 0.18 | |
| CCl ₄ | 74.3 ± 0.51 | 85.4 ± 0.39 | 68.12 ± 0.12 | 5.68 ± 0.027 | 7.67 ± 0.064 | 9.57 ± 0.31 | |
| SJL | 41.7 ± 0.60 | 66.9 ± 0.34 | 49.29 ± 0.09 | 4.59 ± 0.019 | 8.12 ± 0.06 | 7.33 ± 0.27 | |
| Silymarin | 26.8 ± 0.33 | 46.0 ± 0.34 | 24.47 ± 0.13 | 2.33 ± 0.036 | 11.08 ± 0.08 | 6.98 ± 0.42 | |

Values are mean \pm STD and each group contained three rats. Drug treatment lasted for 8 days, "p < 0.001 for the CCl₄-treated group compared to the normal control group; "p < 0.05 for experimental groups compared to the control group. Significance was tested using one-way ANOVA followed by a post hoc Dunnett's test.

3.2. Total antioxidant capacity

The total antioxidant capacity of SJL was determined using a calibration curve for ascorbic acid (y = 0.0043x + 0.1503) and was expressed as the ascorbic acid equivalent (AE). Total antioxidant capacity was 335.70 ± 65.77 mg AE/gm of extract.

3.3. Reducing capacity

The reducing capacity of a compound may serve as a significant indicator of its potential antioxidant activity (35). Reducing properties are generally associated with the presence of reductones, which have been shown to have antioxidant action by breaking the free radical chain reaction by donating a hydrogen atom (36,37). They can react with free radicals to convert them into more stable products. Reductones are also reported to react with certain precursors of peroxide, thus preventing peroxide formation.

The reducing capacity of SJL was investigated using $Fe^{3+}-Fe^{2+}$ transformation. The presence of reductones causes the reduction of $Fe^{3+}/ferricyanide$ complex to the Fe^{2+} form. This Fe^{2+} can be monitored by measuring the formation of Perl's Prussian blue at 700 nm. The extract had significant reducing activity compared to ascorbic acid and this activity increased proportionate to the increase in concentration of the extract, as shown in Figure 2. An increase in the reaction mixture's absorbance indicates an increase in the reducing capacity of the sample.

3.4. Total phenol content and flavonoid content

The Folin-Ciocalteu reagent was used to estimate total phenols present in the extract and this value was expressed as gallic acid equivalents (GAE). The total phenolic content of the sample, calculated on the basis of the standard curve for gallic acid, was found to be 161.78 ± 11.78 mg GAE/gm of SJL extract. The total flavonoid content of SJL was $1,033.37 \pm 62.05$ mg of quercetin equivalent per gram of extract.

3.5. Hepatoprotective activity

The hepatoprotective activity of the crude ethanol extract at a dose of 300 mg/kg body weight in rats with carbon tetrachloride-induced damage is shown in Table 1. For comparison, the table also shows the untreated group (control), carbon tetrachloride-treated group (induction control), and the group treated with the drug (silymarin). The control group (I) had serum SGPT of 22.1 \pm 0.33 U/L, SGOT of 38.8 \pm 0.39 U/L, ALP of 19.26 \pm 0.04 mg/dL, total bilirubin of 1.06 \pm

0.016 mg/dL, total protein of 12.14 ± 0.18 KA, and liver weight of 5.96 ± 0.18 g. In the group with CCl₄induced liver damage (II), serum SGPT increased to 74.3 ± 0.51 U/L, SGOT increased to 85.4 ± 0.39 U/L, ALP increased to 68.12 ± 0.12 mg/dL, total bilirubin increased to 5.68 ± 0.027 mg/dL, total protein increased to 7.67 ± 0.064 KA, and liver weight increased to 9.57 ± 0.31 g. Administration of SJL extract in rats with CCl₄-induced liver damage resulted in gradual normalization of SGPT, SGOT, ALP, total bilirubin, total protein, and liver weight (p < 0.001 compared to the CCl₄-treated group).

4. Discussion

There is growing evidence that the altered production and spatiotemporal distribution of ROS/RNS causes deleterious oxidative/nitrosative stress (38). This condition leads to the interaction of ROS/RNS and bio-molecules (protein, amino acids, lipid, and DNA) and interferes with the expression of a number of genes and signal transduction pathways. Thus, ROS/ RNS play a key role in damage to cell structures as well as various diseases and aging (39). The liver is one of the most important organs in the body. It plays a vital role in regulating, synthesizing, storing, secreting, transforming, and breaking down many different substances in the body. The liver plays a central role in transforming and clearing chemicals and hence it is most susceptible to the free radicals from these agents (40). The body has several mechanisms to counteract oxidative/nitrosative stress with antioxidants, either naturally generating them in situ (endogenous antioxidants) or obtaining them externally through foods (exogenous antioxidants). The roles of antioxidants are to neutralize the excess of free radicals, to protect the cells against their toxic effects, and to help prevent disease.

Results of the current study indicated that the ethanol extract of SJL has significant antioxidant capacity, obvious reducing capacity, and definite DPPH radical scavenging activity. These pharmacological effects of the extract may at least in part be due to phenols and flavonoids components that were found in the SJL ethanol extract. Phenols are ubiquitous secondary metabolites in plants and have a wide range of therapeutic uses because of their antioxidant, antimutagenic, anticarcinogenic, and free radical scavenging activity (41). The antioxidant activity of phenolic compounds is mainly due to their redox properties, which can play an important role in absorbing and neutralizing free radicals, quenching single and triplet oxygen, or decomposing peroxides (42). Flavonoids are a group of polyphenolic compounds with known properties, which include free radical scavenging and inhibition of hydrolytic and oxidative enzymes (43). Furthermore, these

compounds have a strong affinity for iron ions (which are known to catalyze many processes and lead to the appearance of free radicals), so their antiperoxidative activity could also be attributed to a concomitant ability to chelate iron. Therefore, the phenol and flavonoid components in the SJL ethanol extract may have contributed directly to antioxidant action noted in this study.

In most developing countries, there is a high incidence of viral hepatitis. Identification of an efficient hepatoprotective drug derived from natural sources is an urgent necessity. The changes associated with CCl₄-induced liver damage are similar to those of acute viral hepatitis (44). CCl_4 is therefore a useful tool for inducing hepatic damage in experimental animals. The hepatotoxicity of CCl₄ is the result of its reductive dehalogenation. It is catalyzed by cytochromic P450 to produce the highly reactive metabolite trichloromethyl (CCl_3) free radical. This then readily interacts with molecular oxygen to form the trichloromethyl peroxy radical (CCl₃OO·). These free radicals bind covalently to cellular proteins or lipids or extract a hydrogen atom from an unsaturated lipid, thereby initiating lipid peroxidation and consequently leading to liver damage. A substantial increase in the level of serum marker enzymes (SGOT, SGPT, and ALP) and total bilirubin was noted in the CCl₄ control group. The return of elevated levels of serum enzymes to near normal values in groups treated with the leaf extract or standard silymarin is an indication of the stabilization of the plasma membrane and the repair of hepatic tissue damage caused by CCl₄. A decrease in the level of total proteins in rats with CCl₄-induced liver damage is attributed to damage primarily in the endoplasmic reticulum. This results in a loss of P450 and subsequent decrease in protein synthesis. The rise in protein levels in treated groups suggests the stabilization of the endoplasmic reticulum and subsequent protein synthesis. In addition, CCl₄ led to a significant increase in liver weight because it blocks the secretion of hepatic triglycerides in plasma (45). Silymarin and the extract were found to prevent an increase in liver weight in rats. These results suggest that the ethanol extract of SJL offers hepatoprotection by reducing damage or by preserving normal hepatic physiological mechanisms that have been disturbed by a hepatotoxin such as CCl₄. This finding indicates that SJL has protective action in vivo.

Results of the present study indicate that an ethanol extract of SJL has potential antioxidant activity *in vitro* and hepatoprotective activity *in vivo*. However, further studies are needed to examine underlying mechanisms of antioxidant activity *in vitro* and hepatoprotective activity *in vivo*. Studies also need to isolate the active compound(s) responsible for this pharmacological activity.

References

- Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol. 2007; 39:44-84.
- 2. Dröge W. Free radicals in the physiological control of cell function. Physiol Rev. 2002; 82:47-95.
- 3. Zheng M, Storz G. Redox sensing by prokaryotic transcription factors. Biochem Pharmacol. 2000; 59:1-6.
- Lander HM. An essential role for free radicals and derived species in signal transduction. FASEB J. 1997; 11:118-124.
- Ignarro LJ, Cirino G, Casini A, Napoli C. Nitric oxide as a signaling molecule in the vascular system: An overview. J Cardiovasc Pharmacol. 1999; 34:879-886.
- Freidovich I. Fundamental aspects of reactive oxygen species, or what's the matter with oxygen? Ann N Y Acad Sci. 1999; 893:13-18.
- Kumpulainen JT, Salonen JT. Natural antioxidants and Anticarcinogens in Nutrition, Health and Disease. The Royal Society of Chemistry, UK, 1990; pp. 178-187.
- Cook NC, Samman S. Flavonoids-chemistry, metabolism, cardio protective effects, and dietary sources. Nut Biochem. 1996; 7:66-76.
- Liao K, Yin M. Individual and combined antioxidant effects of seven phenolic agents in human erythrocyte membrane ghosts and phosphatidylcholine liposome systems: Importance of the partition coefficient. J Agric Food Chem. 2000; 48:2266-2270.
- Beckman KB, Ames BN. The free radical theory of aging matures. Physiological Rev. 1998; 78:547-581.
- Guengerich FP, Kim DH, Iwasaki M. Role of human cytochrome P450 IIE1 in the oxidation of many low molecular weight cancer suspects. Chem Res Toxicol. 1991; 4:168-179.
- 12. Koop DR. Oxidative and reductive metabolism by cytochrome P450 2E1. FASEB J. 1992; 6:724-730.
- Jaeschke H. Reactive oxygen and mechanisms of inflammatory liver injury. J Gastroenterol Hepatol. 2000; 15:718-724.
- Jaeschke H, Smith CW, Clemens MG, Ganey PE, Roth RA. Mechanisms of inflammatory liver injury: Adhesion molecules and cytotoxicity of neutrophils. Toxicol Appl Pharmacol. 1996; 139:213-226.
- Jaeschke H, Smith CW. Mechanisms of neutrophilinduced parenchymal cell injury. J Leukoc Biol. 1997; 61:647-653.
- Laskin DL, Laskin JD. Role of macrophages and inflammatory mediators in chemically induced toxicity. Toxicology. 2001; 160:111-118.
- Halliwell B, Aeschbach R, Löliger J, Aruoma OI. The characterization of antioxidants. Food Chem Toxic. 1995; 33:601-617.
- Sies H. Strategies of antioxidant defense. Euro J Biochem. 1993; 215:213-219.
- Tantry MA. Plant natural products and drugs: A comprehensive study. Asian Journal of Traditional Medicines. 2009; 4:241-249.
- Abdul G. Medicinal Plants of Bangladesh with Chemical Constituents and Uses. 2nd ed., the Asiatic Society of Bangladesh, Dhaka, 2003; p. 398.
- Morton, Julia F, Miami FL. In: Fruits of warm climates, Rose Apple. 1987; pp. 383-386.
- 22. Avila-Peña D, Peña N, Quintero L, Suárez-Roca H.

Antinociceptive activity of *Syzygium jambos* leaves extract on rats. J Ethnopharmacol. 2007; 112:380-385.

- Murugan S, Uma DP, Kannika PN, Mani KR. Antimicrobial activity of *Syzygium jambos* against selected human pathogens. Int J Pharm Pharm Sci. 2011; 3:44-47
- Braca A, De Tommasi N, Di Bari L, Pizza C, Politi M, Morelli I. Antioxidant principles from *Bauhinia terapotensis*. J Nat Prod. 2001; 64:892-895.
- Viturro C, Molina A, Schmeda-Hirschmann G. Free radical scavengers from *Mutisia friesiana* (Asteraceae) and *Sanicula graveolens* (Apiaceae). Phytother Res. 1999; 13:422-424.
- Prieto P, Pineda M, Aguilar M. Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: Specific application to the determination of vitamin E. Anal Biochem. 1999; 269:337-341.
- Oyaizu M. Studies on product of browning reaction prepared from glucose amine. Japanese Journal of Nutrition. 1986; 44:307-315.
- Chia CC, Ming HY, Hwei MW, Jing CC. Estimation of total phenolic content by two complementary colorimetric methods. Journal of Food and Drug Analysis. 2002; 10:163-173.
- Kumaran, Karunakaran. Estimation of total flavonoid content in propolis by two complementary colorimetric methods. Journal of Food and Drug Analysis. 2007; 10:178-182.
- 30. Bergmeyer HU, Hørder M, Rej R. International Federation of Clinical Chemistry (IFCC) Scientific Committee, Analytical Section: Approved recommendation (1985) on IFCC methods for the measurement of catalytic concentration of enzymes. Part 2. IFCC method for aspartate aminotransferase (L-aspartate: 2-oxoglutarate aminotransferase, EC 2.6.1.1). J Clin Chem Clin Biochem. 1986; 24:497-510.
- 31. Schumann G, Bonora R, Ceriotti F, et al. IFCC primary reference procedures for the measurement of catalytic activity concentrations of enzymes at 37 degrees C. International Federation of Clinical Chemistry and Laboratory Medicine. Part 4. Reference procedure for the measurement of catalytic concentration of alanine aminotransferase. Clin Chem Lab Med. 2002; 40:718-724.
- Kind RN, King EJ. Urinary excretion of acid phosphatase. J Clin Path. 1954; 7:322.
- Jendrassik L, Grof P. Vereinfachte photometrische methoden zur bestimmung des blutbilirubins. Biochem Z. 1938; 81:297.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. J Bio Chem. 1951; 193:265-275.
- Valko M, Leibfritz DJ, Moncol MT, Cronin M, Mazur M, Telser T. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol. 2007; 39:44-84.
- Shahidi F, Janitha PK, Wanasundara PD. Phenolic antioxidants. Food Science and Nutrition. 1992; 32:67-103.
- Osawa T. Novel natiral antioxidants for utilization in food and biological systems. In: Post harvest biochemistry of plant food-materials in tropics (Uritani I, Garcia VV, Mendoza EM, eds.). Japan Scientific Societies Press, Japan, 1994; pp. 241-251.
- Tanaka M, Kuie CW, Nagashima Y, Taguchi T. Applications of antioxidative Maillard reaction products

from histidine and glucose to sardine products. Nippon Suisan Gakkaishi. 1988; 54:1409-1414.

- Duh PD, Tu YY, Yen GC. Antioxidant activity of the aqueous extract of harn jyur (Chrysanthemum morifolium. Ramat). Lebensmittel-Wissenschaft and Technologie. 1999; 32:269-277.
- Gordon MH. The mechanism of antioxidant action *in vitro*. In: Food Antioxidants (Hudson BJF, ed.). Elsevier Applied Science, London, 1990; pp.1-18.
- 41. Hare JM, Stamler JS. NO/redox disequilibrium in the failing heart and cardiovascular system. J Clin Invest. 2005; 115:509-517.
- Friedman, Scott E, Grendell, James H, McQuaid, Kenneth R. In: Current diagnosis & treatment in gastroenterology. Lang Medical Books/McGraw-Hill, New York, 2003; pp.

664-679.

- 43. Yen GC, Duh PD, Tsai CL. Relationship between antioxidant activity and maturity of peanut hulls. J Agric Food Chem. 1993; 41:67-70.
- Rubinstein D. Epinephrine release and liver glycogen levels after carbon tetrachloride administration. Am J Physiol. 1962; 203:1033-1037.
- 45. Aniya Y, Koyama T, Miyagi C, Miyahira M, Inomata C, Kinoshita S, Ichiba T. Free radical scavenging and hepatoprotective actions of the medicinal herb Crassocephalum crepietioides from Okinawa islands. Biol Pharm Bul. 2005; 28:19-23.

(Received July 11, 2012; Revised August 7, 2012; Accepted August 10, 2012)

Original Article

DOI: 10.5582/ddt.2012.v6.4.212

Antimicrobial screening of some Egyptian plants and active flavones from *Lagerstroemia indica* leaves

Yasser Diab^{1,*}, Khaled Atalla², Khaled Elbanna²

¹ Department of Biochemistry, Faculty of Agriculture, Fayoum University, Fayoum, Egypt;

² Department of Agricultural Microbiology, Faculty of Agriculture, Fayoum University, Fayoum, Egypt.

ABSTRACT: One hundred and twenty four plant extracts were evaluated for their antimicrobial activity against four pathogenic bacteria (Staphylococcus aureus (ATCC 8095), Salmonella enteritides (ATCC 13076), Escherichia coli (ATCC 25922), and Listeria monocytogenes (ATCC 15313)) and Candida albicans yeast (ATCC 10231) using the disk diffusion and broth microdilution methods. Of the plant extracts, fourteen exhibited antimicrobial activity against two or more of the five microorganisms tested. Only the methanol extract of Lagerstroemia indica leaves exhibited antimicrobial activity against all pathogenic bacteria and C. albicans yeast that were tested. Purification of the methanol extract of L. indica leaves using antimicrobial assay-guided isolation yielded one pure active compound. The chemical structure of the isolated active compound was found to be '4-methoxy apigenin-8-C- β -D-glucopyranoside; cytisoside according to detailed spectroscopic analysis of its nuclear magnetic resonance and mass spectrometry data. The compound exhibited antimicrobial activity against C. albicans (minimum lethal concentration (MLC): 32 µg/mL), S. aureus (MLC: 16 µg/mL), S. enteritides (MLC: 16 µg/mL), E. coli (MLC: 16 µg/mL), and L. monocytogenes (MLC: 16 µg/mL). The present study found that the methanol extract of L. indica leaves holds great promise as a potential source of beneficial antimicrobial components for different applications.

Keywords: Pathogenic bacteria, *Candida albicans*, *Lagerstroemia indica*, flavones, antimicrobials

*Address correspondence to:

1. Introduction

Secondary metabolites produced by plants constitute a major source of bioactive substances. Over the past two decades, the scientific interest in these plant metabolites has increased as part of the search for new therapeutic agents, partly due to the increasing resistance of microorganisms to most antimicrobial drugs currently used in medicine and agriculture (1). In addition, the need for safer agrochemicals with less environmental and mammalian toxicity is a major concern. In an attempt to discover new chemical classes of antimicrobial drugs that could resolve these problems, a wide range of plant extracts has been examined for antimicrobial properties (2,3).

Food products are susceptible to contamination and spoilage by bacteria. More than 90% of the cases of food poisoning or food-related infection each year are caused by Gram-positive bacteria such as Staphylococcus aureus and Listeria monocytogenes or Gram-negative bacteria such as Escherichia coli and Salmonella enteritidis (4-7). In recent years, the effects of plant extracts and phytochemicals on food pathogenic bacteria have been studied (8-11). Candidiasis is an infection caused by Candida yeast, especially Candida albicans, and is associated with skin infection and other health problems. Treatment of candidiasis can be problematic due to the limited number of effective antifungal drugs, toxicity of the available antifungal drugs, resistance of Candida to commonly-used antifungal drugs, relapse of Candida infections, and the high cost of antifungal drugs (12). The investigation of natural products for activity against Candida species has therefore increased in the last 10 years, with approximately 258 plant species from 94 families having been investigated (13).

The present study evaluated the chloroform and methanol extracts of 124 plant species grown in Egypt (belonging to 56 plant families) for their antimicrobial activity against *C. albicans* yeast and four food pathogenic bacteria (*S. aureus*, *S. enteritidis*, *E. coli*, and *L. monocytogenes*). As the methanol extract of the leaves of *Lagerstroemia indica* exhibited significant antimicrobial activity against all five microorganisms, bioactivity-guided separation of the methanol extract of *L. indica* leaves was used to isolate and identify the active compound.

Dr. Yasser Mohamed Diab, Biochemistry Department, Faculty of Agriculture, Fayoum University, Fayoum, 63514, Egypt.

E-mail: ydiab@hotmail.com; ymd00@fayoum.edu.eg

2. Materials and Methods

2.1. Chemicals and plant materials

All chemicals and reagents used in this study were of analytical grade and were purchased from Sigma Chemical Co. (St. Louis, MO, USA), BDH (Dorset, England) or Fluka Chemie Co. (Buchs, Switzerland).

Leaf samples from a total of 124 plant species belonging to 56 families were collected during April and May 2008 from El-Shrouk farm on the Cairo-Alexandria desert road, 72 km north of Cairo. The botanical identification of the collected specimens was confirmed by Dr. T. Labeb, Herbarium of Orman Garden, Horticulture Research Institute, Giza, Egypt. A voucher specimen of each plant was deposited in the herbarium of the Biochemistry Department, Faculty of Agriculture, Fayoum University. The leaves were cleaned, air-dried in the shade, and then powdered to pass through 24 mesh using a laboratory mill. Powdered material was stored in an air-tight container at room temperature (28 $\pm 2^{\circ}$ C) and protected from light until use.

2.2. Detection and identification methods

Analytical thin-layer chromatography (TLC) was carried out on Merck precoated silica gel plates (F245, 0.25 mm thickness). Analytes were visualized under ultraviolet (UV) light (254 and 365 nm) by spraying 30% H_2SO_4 in methanol followed by heating at 105°C for 5 min, by spraying AlCl₃ reagent to detect flavonoids, or by spraying naphthoresorcinol/phosphoric acid followed by heating at 105°C for 10 min to detect sugars. The purified compounds were preliminary characterized phytochemically for the presence of saponins, flavonoids, alkaloids, tannins, and glycosides using TLC according to methods described previously (*16*).

¹H, ¹³C nuclear magnetic resonance (NMR) and heteronuclear multiple bond correlation (HMBC) spectra were recorded in deutero-methanol (CD₃OD) on a Bruker DRx 400 MHz spectrometer (Varian Inc., Palo Alto, CA, USA) at 400 MHz for ¹H and 100 MHz for ¹³C. UV spectra were recorded on a Cecil 3000 series spectrophotometer (Cecil Instruments Ltd., Cambridge, UK). Mass spectra were recorded on a GC/MS Qp 100 Ex Shimadzu Mass spectrometer (Shimadzu, Kyoto, Japan) at 70 eV.

2.3. Preparation of plant extracts

A known amount of air-dried powdered leaves from each plant was extracted with chloroform at room temperature ($28 \pm 2^{\circ}$ C). This procedure was repeated at least five times and until the organic solvent remained colorless. The same chloroform-extracted samples were then extracted with methanol at least five times or until the solvent remained colorless. The extracts obtained were filtered using Whatman No. 1 filter paper and the combined extract (filtrate) was evaporated to dryness in a rotary evaporator at 45°C. The antimicrobial activity of the dry residue was then assayed.

2.4. Isolation of bioactive compound(s) from L. indica leaves

The bioactive methanol extract of L. indica leaves was subjected to chromatography to isolate antimicrobial component(s) as follows. Thirteen grams of extract residue in methanol were loaded onto a chromatographic column (5 cm × 100 cm) packed with silica gel (230-400 mesh, 700 g, Merck & Co. Inc., Whitehouse Station, NJ, USA) and eluted with a gradient of chloroform:methanol (85:15, 70:30, 50:50, and 30:70, v/v; 2 L for each eluent). Twenty fractions (100 mL) of each eluent were collected and assayed with TLC. Based on their similarities in TLC, the collected fractions were combined into 11 fractions that were further tested for antimicrobial activity. Two fractions (8 and 9) exhibited strong antimicrobial activity. These fractions (8, 1.5 g and 9, 2.9 g) eluted between 400-900 mL and 1,000-2,000 mL of chloroform:methanol (50:50), respectively. Fractions 8 and 9 were subjected to solid phase extraction using a C₁₈ cartridge (Waters, Milford, MA, USA) and subsequently chromatographed on a Sephadex LH20 (GE Healthcare-Bio-Sciences, Piscataway, NJ, USA) column (2.7 cm inner diameter × 60 cm, 50 g resin) using methanol as the mobile phase. The eluates were combined into eight fractions (A-H) on the basis of similar TLC profiles. Fraction E (960 mg) contained the most antimicrobial activity and was further purified on a silica gel column (1.5 cm inner diameter \times 60 cm; 50 g) with chloroform:methanol:water (60:40:5, v/v) followed by purification on a Sephadex LH20 column (1.6 cm inner diameter \times 40 cm; 20 g) with methanol as the eluent to yield 543 mg of pure active compound.

2.5. Test organisms and the agar disk diffusion method

The four food pathogenic bacteria (*S. aureus* (ATCC 8095), *S. enteritidis* (ATCC 13076), *E. coli* (ATCC 25922), *L. monocytogenes* (ATCC 15313)) and *C. albicans* (ATCC 10231) were obtained from the Department of Agricultural Microbiology, Faculty of Agriculture, Fayoum University, El-Fayoum, Egypt. Stock cultures of bacteria were maintained on nutrient agar slants at 4°C, and Candida yeast was maintained on potato dextrose agar slants at 4°C. Bacterial and yeast cultures were subcultured in Petri dishes prior to testing.

The disk diffusion method (14) was used to assay the antimicrobial activity of plant extracts. Three sterilized filter paper discs (6 mm) were soaked with each plant extract (1 g/10 mL) and dried at 40°C. The dried disks were transferred to the surface of the inoculated plates in triplicate. Plates with pathogenic bacteria were then incubated at 37°C for 24-48 h and plates with *Candida* yeast were incubated at 30°C for 48-72 h. Afterwards,

bioactivity was determined by measuring the diameter of inhibition zones (DIZ) around each disk in mm. Solvent without test compounds was used as a control.

2.6. Determination of minimum lethal concentrations (MLC)

The MLC of leaf extracts with potential antimicrobial activity and their isolated active constituent(s) were determined using the broth micro-dilution method (*15*). The lowest concentration of a tested extract or compound resulted in a viable count of less than 0.1% of the original inoculum (1×108 colony-forming units per milliliter, cfu/mL, as compared to the turbidity of the McFarland 0.5 standard). Ampicillin and fluconazole served as standards for comparison in antibacterial and antifungal tests, respectively.

3. Results

3.1. Antimicrobial activity of plant leaf extracts

The chloroform and methanol extracts of a number of Egyptian plants (124 plant species) belonging to 56 plant families were evaluated for their antimicrobial activity against five pathogenic organisms (*C. albicans* yeast and four food pathogenic bacteria: *S. aureus*, *S. enteritidis*, *E. coli*, and *L. monocytogenes*) using a disk diffusion assay. The methanolic extracts of fourteen plant species exhibited significant antimicrobial activity against two or more of the five pathogenic microorganisms that were tested. The diameter of inhibition zones (DIZ) and the MLC of the promising plant extracts are shown in Table 1. The data indicated that the methanol extracts of these plants exhibited

varying levels of antimicrobial activity against the test microorganisms.

The results in Table 1 show that among the fourteen promising plant extracts tested, extracts of *Pelargonium* odoratissimum, *Pelargonium zonale*, and *Rosa spp* were active against *E. coli*, *S. aureus*, *S. enteritidis*, and *L. monocytogenes*. Seven extracts were found to be only active against *E. coli*, *S. enteritidis*, and *L. monocytogenes*. This variation in activity may be due to the intrinsic tolerance of microorganisms and the nature and combination of phytocompounds present in these extracts (17). Only a leaf extract of *L. indica* was active against all five of the pathogenic microorganisms tested. Of the bacterial strains tested, *E. coli* was found to be the most sensitive and was inhibited by all plant extracts that exhibited antimicrobial activity.

3.2. Purification of an antimicrobial compound from methanolic L. indica leaf extract

Methanolic *L. indica* leaf extract had both bactericidal and fungicidal activity against the five microorganisms tested, with an MLC ranging between 60 and 120 µg/mL. This is similar to the MLC of standard antibiotics (fluconazole and ampicillin ranging from 24 to 128 µg/mL). The results clearly showed that the methanol extract of *L. indica* was the most effective plant extract. It inhibited the growth of both Gram-negative and Gram-positive bacteria that were tested and also of the yeast *C. albicans*, which has a sophisticated mechanism of resistance to many drugs (*18*). Because the methanolic extract of *L. indica* had broad-spectrum antimicrobial action against all five pathogenic organisms tested, this extract was selected for phytochemical analysis and isolation of its bioactive constituent(s).

| Table 1. Evaluation of the antimicrobial potential of promising plant extracts and the active compound from <i>L. indica</i> (cytisoside) |
|---|
| indicated by the diameter of inhibition zones (DIZ, mm) and the minimum lethal concentration (MLC, µg/mL) against |
| pathogenic organisms: C. albicans yeast and four bacteria, i.e. L. monocytogenes, S. aureus, S. enteritidis, and E. coli |

| Plant species | Plant family | L. monocytogenes | | S. enteritidis | | S. aureus | | E. coli | | C. albicans | |
|---------------------------|---------------|------------------|-------|----------------|-----|-----------|-------|---------|-------|-------------|-------|
| | | DIZ | MLC | DIZ | MLC | DIZ | MLC | DIZ | MLC | DIZ | MLC |
| Yucca desmettiana | Agavaceae | _ | _ | _ | _ | _ | _ | 24 | > 120 | 20 | > 120 |
| Dracaena marginata | Agavaceae | _ | _ | _ | _ | _ | _ | 21 | 120 | 23 | > 120 |
| Schinus terebinthifolius | Anacardiaceae | 23 | > 120 | 31 | 120 | _ | _ | 27 | > 120 | _ | _ |
| Euonymus japonica | Celastraceae | 15 | 120 | 15 | 120 | _ | _ | 20 | 120 | _ | _ |
| Conocarpus erectus | Combretaceae | 20 | 120 | 24 | 60 | _ | _ | 34 | 120 | _ | _ |
| Acalypha marginata | Euphorbiaceae | 17 | 120 | 25 | 30 | _ | _ | 30 | 120 | _ | _ |
| Pelargonium odoratissimum | Geraniaceae | 18 | 120 | 20 | 120 | 35 | 120 | 30 | 120 | _ | _ |
| Pelargonium zonale | Geraniaceae | 23 | 120 | 32 | 30 | 32 | > 120 | 30 | 30 | _ | _ |
| Asparagus plumosus | Liliaceae | _ | _ | _ | _ | _ | _ | 15 | > 120 | 18 | > 120 |
| Lagerstroemia indica | Lythraceae | 23 | 120 | 30 | 60 | 30 | 60 | 26 | 120 | 20 | 120 |
| Myrtus communis | Myrtaceae | 23 | > 120 | 31 | 120 | _ | _ | 20 | 120 | _ | _ |
| Punica granatum | Punicaceae | 17 | 120 | 21 | 30 | _ | _ | 28 | 120 | _ | _ |
| Rosa spp | Rosaceae | 20 | 120 | 29 | 120 | 34 | 120 | 32 | 120 | _ | _ |
| Cestrum diurnum | Solanaceae | 12 | > 120 | _ | _ | _ | _ | 16 | > 120 | 21 | 120 |
| Cytisoside (Control) | | _ | 16 | _ | 16 | _ | 16 | _ | 16 | _ | 32 |
| Fluconazole (Control) | | _ | _ | - | - | - | _ | _ | - | - | > 128 |
| Ampicillin (Control) | | - | 24 | - | 24 | - | 24 | _ | 24 | _ | - |

Phytochemical examination of the extract indicated the presence of phenolic compounds, flavonoids, triterpenoids, alkaloids, and glycoside compounds. One or more of these secondary metabolites may be responsible for antimicrobial activity (19).

Bioactivity-guided separation of the methanol extract of the dried leaves of *L. indica* resulted in the isolation of one chromatographically pure compound. The isolated compound had antimicrobial activity against the five human pathogenic microorganisms tested (Table 1). The MLC ranged between 16 and 32 µg/mL, indicating greater activity than that of the standard antibiotics used as positive controls in this experiment. Therefore, this compound was at least in part responsible for the antimicrobial activity of leaves of *L. indica*.

3.3. Characterization and identification of the active compound

The purified compound was obtained as a yellow amorphous solid that resulted in a positive color reaction with AlCl₃ reagent on TLC, suggesting a flavonoid. Electron ionization mass spectrometry resulted in a molecular ion peak [M⁺] at 434 m/z. This finding, together with ¹H- and ¹³C-NMR spectroscopic data, suggested a molecular formula of C₂₁H₂₂O₁₀. The UV spectrum had absorption maxima at λ 270 (band II) and 333 nm (band I), which are characteristic absorption bands of the flavone skeleton (20). No shift in band I of this compound was observed after the addition of AlCl₃/HCl, suggesting the formation of a hydroxyl-keto complex at 5-OH and the absence of an O-dihydroxyl group in the β -ring (21). The ¹H-NMR spectrum of this compound displayed signals characteristic of a methoxy group ($\delta_{\rm H}$ 3.5 (3H)) and six aromatic hydrogens ($\delta_{\rm H}$ 6.4 (H), 6.5 (H), 6.83 (2H), and 7.75 (2H)) in the low field region. The appearance of four aromatic proton signals at δ 6.83 (2H, d, J = 8.8 Hz, H-'3, H-'5) and 7.75 (2H, d, J = 8.8 Hz, H-'2, H-'6) along with a characteristic pattern for an A2B2 system confirmed the substitution of the β -ring at the '4 position. The proton signals at $\delta_{\rm H}$ 6.4 (1H, s) and 6.5 (1H, s) were attributed to H-3 based on HMBC and compared to reported data (20, 22). The presence of a free 7-hydroxyl group was noted based on the appearance of a bathochromic shift in the UV spectrum upon the addition of sodium acetate. The position of the methoxy group at the '4-O-position of the flavone moiety was confirmed by the heteronuclear correlation according to the HMBC spectrum and the UV spectrum data in the presence of NaOMe. Therefore, the flavone moiety of this compound was determined to be '4-methoxy apigenin (acacetin). The presence of β -D-glucopyranose as the sole sugar moiety was noted based on NMR spectra (Table 2) given the appearance of only one anomeric proton signal at δ 4.07 (d, J = 7.6 Hz), five glucosyl protons ($\delta_{\rm H}$ 3.08-3.8m) in the ¹H-NMR spectrum, and the appearance of one anomeric carbon atom at δ 104.68 ppm and five glucosyl

carbons (75.01, 79.76, 71.56, and 62.15 ppm; C-2 to C-6) in the ¹³C-NMR spectrum. The position of the C-glucose moiety at the C-8-position was noted based on the HMBC spectrum and by comparison to reported data (20). Thus, the structure of this compound (Figure 1) was found to be '4-methoxy apigenin (acacetin)-8-C- β -D-glucopyranose, which was previously reported as cytisoside (20).

4. Discussion

The present study identified '4-methoxy apigenin (acacetin)-8-C- β -D-glucopyranose (cytisoside) as the active antimicrobial component of methanolic *L. indica* leaf extract. This component had both antibacterial and antifungal activity. To the extent known, no previous studies isolated this active compound from the leaves of *L. indica*. Thus, this is the first study to isolate and identify an active flavone glycoside from *L. indica*,

Table 2. NMR data for the active isolated compound in CD_3OD

| CD ₃ OD | | | | | | |
|--------------------|------------------|-----------------|----------------------|--|--|--|
| Atom No. | $\delta_{\rm C}$ | ¹³ C | $^{1}\mathrm{H}$ | | | |
| 2 | С | 162.80 | _ | | | |
| 3 | CH | 103.50 | 6.50 (s) | | | |
| 4 | CO | 184.06 | - | | | |
| 5 | С | 164.88 | - | | | |
| 6 | CH | 95.24 | 6.40 (s) | | | |
| 7 | С | 165.04 | - | | | |
| 8 | С | 109.20 | - | | | |
| 9 | С | 158.73 | - | | | |
| 10 | С | 105.20 | - | | | |
| '1 | С | 123.13 | - | | | |
| '2 | CH | 129.46 | 7.75 (d, J = 8.8) | | | |
| '3 | CH | 117.04 | 6.83 (d, J = 8.8) | | | |
| '4 | С | 166.20 | - | | | |
| '5 | CH | 117.04 | 6.83 (d, J = 8.8) | | | |
| '6 | CH | 129.46 | 7.75 (d, $J = 8.8$) | | | |
| | OCH ₃ | 57.29 | 3.5 (s) | | | |
| "1 | CH | 104.68 | 4.07 (d, J = 7.6) | | | |
| "2 | CH | 75.01 | 3.26 | | | |
| "3 | CH | 79.76 | 3.40 | | | |
| "4 | СН | 71.32 | 3.08 | | | |
| "5 | CH | 77.56 | 3.21 | | | |
| "6 | CH_2 | 62.15 | 3.68, 3.80 | | | |

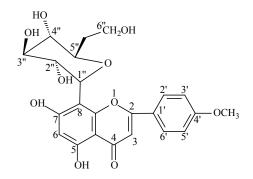


Figure 1. Structural formula of the active isolated compound ('4-methoxy apigenin-8-C- β -D-glucopyranoside; cytisoside).

i.e. cytisoside, with antimicrobial activity against the five pathogenic microorganisms (*C. albicans* yeast and four bacteria: *S. aureus*, *S. enteritidis*, *E. coli*, and *L. monocytogenes*) tested. Comparison revealed that the bioactive compound had greater antimicrobial activity than both the crude extract of *L. indica* and standard antibiotics (fluconazole and ampicillin).

Flavonoids are ubiquitous in photosynthesizing cells and therefore occur widely in the plant kingdom (23). Various flavonoids have been reported to possess a wide range of biological action, such as antimicrobial or antioxidant activity (19). Moreover, several groups of researchers have noted a synergy between biologically active flavonoids and existing chemotherapeutics (24). Flavonoids isolated from plant extracts are reported to have antimicrobial activity against food pathogenic bacteria and against C. albicans; these include compounds such as apigenin (25,26), quercetin and quercetin glycosides (27,28), luteolin (29), kaempferol (30), and flavan derivatives. Their remarkable activity is attributed to the inhibition of nucleic acid synthesis (31), inhibition of energy metabolism (32), and inhibition of cytoplasmic membrane function (33). Flavonoids are likely to have minimal toxicity because flavonoids are widely distributed in edible plants and beverages and have previously been used in traditional medicine.

The present study found that the methanol extract of *L. indica* leaves holds great promise as a potential source of beneficial antimicrobial components for different applications (foods and pharmaceuticals) and could resolve the problems of drug resistance and the harmful effects of synthetic compounds. However, further *in vivo* studies are needed to investigate the pharmacological and toxicological properties of *L. indica* extract before it can be considered as a new antimicrobial ingredient for the nutraceutical or functional food market.

Acknowledgements

The authors wish to thank Prof. A. M. Emam (Fayoum University, Egypt). Thanks are also extended to Prof. Dr. V. Roussis (Department of Pharmacognosy & Chemistry of Natural Products, School of Pharmacy, University of Athens, Greece). The authors also thank Mr. A. S. Mahmoud (Fayoum University, Egypt) for his help with preliminary screening and Dr. Martin Krehenbrink (Oxford University, UK) for help with revisions.

References

- WHO publication. WHO strategy for containment of antimicrobial resistance. Available at: www.who.int/ drugresistance/WHO_Global_Strategy_English.pdf. 2001 (accessed March 2012).
- Cragg GM, Newman DJ, Snader KM. Natural products in drug discovery and development. J Nat Prod. 1997; 60:52-60.

- De Valk H, Delarocque-Astagneau E, Colomb G, Ple S, Godard E, Vaillant V, Haeghebaet S, Bouvet Ph, Grimont F, Grimont P, Desenclos J-C. A community-wide outbreak of *Salmonella enteritidis* serotype Typhimurium infection associated with eating a raw milk soft cheese in France. Epidemiol Infect. 2000; 124:1-7.
- Idris S, Ndukwe GI, Gimba CE. Preliminary phytochemical screening and antimicrobial activity of seed extracts of *Persea americana* (Avocado pear). Bayero Journal of Pure and Applied Sciences. 2009; 2: 173-176.
- Rudol M, Sherer S. High incidence of *Listeria* monocytogenes in Europea red smear cheese. Int J Food Microbiol. 2001; 63:91-98.
- Araujo VS, Pagliares VA, Queiroz ML, Freitas-Almeida AC. Occurrence of Staphylococcus and enteropathogens in soft cheese commercialized in the city of Rio de Janeiro, Brazil. J Appl Microbiol. 2002; 92:1172-1177.
- Conedera G, Dalvit P, Martini M, Galiero G, Gramaglia M, Goffredo E, Loffredo G, Morabito S, Ottaviani D, Paterlini F, Pezzotti G, Pisanu M, Semprini P, Caprioli A. Verocytotoxin-producing *Escherichia coli* 0157 in minced beef and dairy products in Italy. Int J Food Microbiol. 2004; 96:67-73.
- Annan K, Houghton PJ. Antibacterial, antioxidant and fibroblast growth stimulation of aqueous extracts of *Ficus asperifolia* Miq. and *Gossypium arboreum* L., wound-healing plants of Ghana. J Ethnopharmacol. 2008; 119:141-144.
- Akorum S, bendjeddou D, Satta D, Lalaoui K. Antibacterial activity and acute toxicity effect of flavonoids extracted from *Mentha longifolia*. Am-Euras J Sci Res. 2009; 4:93-99.
- Mbosso EJT, Ngouela S, Nguedia JCA, Beng VP, Rohmer M, Tsamo E. *In vitro* antimicrobial activity of extracts and compounds of some selected medicinal plants from Cameroon. J Ethnopharmacol. 2010; 128: 476-481.
- 11. Sakunpak A, Panichayupakaranant P. Antibacterial activity of Thai edible plants against gastrointestinal pathogenic bacteria and isolation of a new broad spectrum antibacterial polyisoprenylated benzophenone, chamuangone. Food Chem. 2012; 130:826-831.
- Runyoro DK, Matee MI, Ngassapa OD, Joseph CC, Mbwambo ZH. Screening of Tanzanian medicinal plants for anti-Candida activity. BMC Complement Altern Med. 2006; 6:11.
- Naeini A, Khosravi AR, Chitsaz M, Shokri H, Kamlnejad M. Anti-Candida albicans activity of some Iranian plants used in traditional medicine. J Mycol Med. 2009; 19: 168-172.
- Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. Am J Clin Pathol. 1966; 45:493-496.
- NCCLS. Performance standards for antimicrobial susceptibility testing; ninth informational supplement; NCCLS document M100-S9 2008; (pp. 120-126). Wayne: National Committee for Clinical Laboratory Standard.
- 16. Farnsworth NR. Biological and phytochemical screening of plants. J Pharm Sci. 1966; 55:225-276.
- Pathak K, Pathak A, Singl A. Flavonoids as medicinal agents – recent advances. Fitoterapia. LXII, 1991; 5:371-389.
- 18. Casalinuovo IA, Francesco PDI, Garaci E. Fluconazole

resistance in *Candida albicans*: A review of mechanisms. Eur Rev Med Pharmacol Sci. 2004; 8:69-77.

- 19. Harborne JB, Baxter H. The handbook of natural flavonoids. John Wiley and Sons, Chichester, UK, 1999.
- Mabry TJ, Markham KR, Thomas MB. The Systematic Identification of Flavonoids. Springer Verlag, Berlin, Heidelberg, New York, 1970.
- Markham KR. Flavones, flavonols and their glycosides. In: Methods in plant Biochemistry (Harborne J, ed.). London, UK, 1989; pp. 197-232.
- Markham KR, Geiger H. ¹H nuclear magnetic resonance spectroscopy of flavonoids and their glycosides in hexadeuterodimethylsulphoxide. In: The Flavonoids. Advances in Research since 1986 (Harborne JB, ed.). Chapman and Hall, London, UK, 1993; pp. 441-497.
- Havsteen B. Flavonoids, a class of natural products of high pharmacological potency. Biochem Pharmacol. 1983; 32:1141-1148.
- Cushnie TP, Lamb AJ. Antimicrobial activity of flavonoids. Int J Antimicrob Agents. 2005; 26:343-356.
- 25. Sato Y, Suzaki S, Nishikawa T, Kihara M, Shibata H, Higuti T. Phytochemical flavones isolated from *Scutellaria barbata* and antibacterial activity against methicillin-resistant *Staphylococcus aureus*. J Ethnopharmacol. 2000; 72:483-488.
- Narayana KR, Reddy SR, Chaluvadi MR, Krishna DR. Bioflavonoids classification, pharmacological, biochemical effects and therapeutic potential. Indian J

Pharmacol. 2001; 33:2-16.

- Ramaswamy AS, Jayaraman S, Sirsi M, Rao KH. Antibacterial action of some naturally occurring citrus bioflavonoids. Indian J Exp Biol. 1972; 10:72-73.
- Taleb-Contini SH, Salvador MJ, Watanabe E, Ito I, de Oleveira DCR. Antimicrobial activity of flavonoids and steroids isolated from two *Chromolaena* species. Rev Bras Cienc Farm. 2003; 39:403-408.
- Miski M, Ulubelen A, Johansson C, Mabry TJ. Antibacterial activity studies of flavonoids from *Salvia palaestina*. J Nat Prod. 1983; 46:874-875.
- Nishino C, Enoki N, Tawata S, Mori A, Kobayashi K, Fukushima M. Antibacterial activity of flavonoids against *Staphylococcus epidermidis*, a skin bacterium. Agric Biol Chem. 1987; 51:139-143.
- Ohemeng KA, Schwender CF, Fu KP, Barrett JF. DNA gyrase inhibitory and antibacterial activity of some flavones (1). Bioorg Med Chem Lett. 1993; 3:225-230.
- Haraguchi H, Tanimoto K, Tamura Y, Mizutani K, Kinoshita T. Mode of antibacterial action of retrochalcones from Glycyrrhiza inflata. Phytochemistry. 1998; 48:125-129.
- Tsuchiya H, Iinuma M. Reduction of membrane fluidity by antibacterial sophoraflavanone G isolated from *Sophora exigua*. Phytomedicine. 2000; 7:161-165.

(Received June 13, 2012; Revised August 14, 2012; Accepted August 15, 2012)

Original Article

DOI: 10.5582/ddt.2012.v6.4.218

Optimization of cell-wall skeleton derived from *Mycobacterium bovis* BCG Tokyo 172 (SMP-105) emulsion in delayed-type hypersensitivity and antitumor models

Masanori Miyauchi^{1,*}, Masashi Murata¹, Akihisa Fukushima¹, Toshiyuki Sato¹, Masae Nakagawa¹, Tetsuya Fujii¹, Norimasa Koseki², Nobuyuki Chiba¹, Yasuo Kashiwazaki¹

¹ Pharmacology Research Laboratories, Dainippon Sumitomo Pharma Co., Ltd., Osaka-shi, Osaka, Japan;

² Formulation Research & Development Laboratories, Dainippon Sumitomo Pharma Co., Ltd., Ibaraki-shi, Osaka, Japan.

ABSTRACT: Cell-wall skeleton prepared from Mycobacterium bovis BCG (BCG-CWS) is known as a potent adjuvant and has been shown to possess antitumor activity in many non-clinical and clinical studies. As there are no approved BCG-CWS formulations for cancer therapy, we investigated the potential for cancer immunotherapy of SMP-105, our originally produced BCG-CWS. For optimizing SMP-105 emulsion, we compared the effects of drakeoland squalane-based SMP-105 emulsions on IFN-y production in rats and evaluated their ability to induce skin reaction in guinea pigs. Both emulsions had the same activity in both experiments. We selected squalane as base material and produced two types of squalane-based formulations (vialed emulsion and pumped emulsion) that can easily be prepared as oilin-water emulsions. Although the vialed emulsion showed the same pattern of distribution as a usual homogenized emulsion, the pumped emulsion showed more uniform distribution than the other two emulsions. Whereas both emulsions enhanced strong delayed type hypersensitivity (DTH) reaction in a mouse model, the pumped emulsion induced slightly smaller edema. Data on oil droplet size distribution suggest that few micrometer oil droplet size might be appropriate for oil-in-water microemulsion of SMP-105. The antitumor potency of SMP-105 emulsion was stronger than that of some of the launched tolllike receptor (TLR) agonists (Aldara cream, Picibanil, and Immunobladder). Aldara and Picibanil showed limited antitumor effectiveness, while Immunobladder had almost the same effect as SMP-105 at the highest dose, but needed about 10 times the amount of SMP-105. These findings first indicate that SMP-105 has great potential in cancer immunotherapy.

Keywords: Oil-in-water emulsion, oil droplet size distribution, BCG-CWS, SMP-105

1. Introduction

With the recent approval of sipuleucel-T (Provenge[®]) and ipilimumab (Yervoy[®]) by the FDA (1), cancer immunotherapy seems to be well underway, and is increasingly attracting a great deal of attention. Cellwall skeleton prepared from Mycobacterium bovis BCG (BCG-CWS) is known as an activator of innate immunity (2) and has been studied in many clinical studies (3-6). We have previously reported that SMP-105, a highly pure cell-wall skeleton prepared from Mycobacterium bovis BCG Tokyo 172 strain, exhibits potent immunostimulatory activity and strong antitumor effect in animal models (7,8). SMP-105 is an insoluble toll-like receptor 2 (TLR2) ligand that elicits immune reactions, including induction of interferon- γ (IFN- γ) producing cells and cytotoxic T lymphocytes (CTL), and prevents tumor growth through TLR2 (9). SMP-105 requires phagocytosis by macrophages or dendritic cells (DCs) for immune activation and shows different in vitro and in vivo effects from those of Pam3CSK4, a soluble TLR2 ligand (10). Intradermal injection of an oil droplet of emulsified SMP-105 shows that this antigen is readily engulfed by phagocytes at the draining lymph nodes (11).

Freund's Complete Adjuvant (FCA), an immunopotentiator composed of inactivated and dried mycobacteria in mineral oil, is known to stimulate cell-mediated immunity, but may cause severe side effects, which effectively excludes it from clinical use. BCG-CWS on the other hand is used as oil-in-water emulsion, because the immunostimulatory effect of this adjuvant depends on its emulsion form (12). Antitumor activity of a drakeol-base oil-in-water emulsion in clinical research has already been reported (13). On the other hand, there are reports showing that BCG-CWS emulsified with squalane induces safer antitumor immunity (14,15). Squalane-based emulsions

^{*}Address correspondence to:

Dr. Masanori Miyauchi, Drug Research Division, Dainippon Sumitomo Pharma Co., Ltd., 3-1-98, Kasugadenaka, Konohana-ku, Osaka-shi, Osaka, 554-0022, Japan. E-mail: masanori-miyauchi@ds-pharma.co.jp

and their hydrogenated derivatives have been shown as potent and safe vaccine adjuvant in preclinical and clinical studies (*16-19*). In fact, MF59TM, an adjuvant emulsion based on squalane, has already been approved for human use (*20*).

Studies that investigated BCG-CWS adjuvant particle size and their phagocytosis have indicated that particles with a size ranging from 0.5-2 µm are readily engulfed by macrophages (21, 22). Labeled squalane oil-based formulations have been shown to be taken up by macrophages and DCs at the site of injection (23). Ideal particles size has also been investigated for drug delivery systems and as a contributing factor to generation of immune response (24-26). Particles made from poly lactic-co-glycolic acid (PLGA) can be used as a delivery system that provides adjuvant activity (27,28). These polymeric particulate delivery systems are able to present antigens and activate both humoral and cellular responses (29,30). A 5 µm PLGA particle containing hepatitis B virus surface antigen elicited higher immune response than a larger particle of about 12 μ m (31). Although we have previously shown that SMP-105 requires phagocytosis by immune cells for enhancing delayed type hypersensitivity (DTH) reaction in vivo (10), there are no studies on the correlation between oil droplet size of BCG-CWS emulsion and immune activation.

In this report, we prepared several SMP-105 emulsions and compared their DTH reaction and effects in a lymph node metastasis model. We also characterized oil droplet size distribution in each emulsion. Finally, we investigated antitumor effect of originally optimized SMP-105 emulsion and compared it to that of launched TLR agonists in clinical use.

2. Materials and Methods

2.1. Preparation of SMP-105

SMP-105 was prepared as previously described (4,32). Contamination with endotoxin was less than 0.005 endotoxin units/mg. The oil-in-water emulsion of SMP-105 (homogenized emulsion) with squalane or drakeol was prepared by homogenization with a Potter-type homogenizer as previously described (8). The first original SMP-105 formulation, an oil-in-water emulsion of SMP-105 (vialed emulsion), was prepared and lyophilized on the thousand-vial scale (7,11). The vialed emulsion can be prepared by adding only water and vortexing for several seconds. Vehicle preparation used the same formulation as the vialed emulsion, except for SMP-105. The second original SMP-105 formulation, a pumped emulsified oil-in-water emulsion of SMP-105 (pumped emulsion), was prepared by pumping with an SPG pump connector (SPG techno, Miyazaki, Japan). Oil droplet size of the uniformly-sized emulsion was modulated by changing the frequency of pumping (once

to 10 times). The pumped emulsion and vialed emulsion had identical content.

2.2. Materials

Aldara cream (5% imiquimod) was purchased from 3M Pharmaceuticals (3M Pharmaceuticals, Minnesota, USA), and Immunobladder was purchased from Alfresa Corporation (Japan BCG Laboratory, Tokyo, Japan). Picibanil (OK-432) 5KE was purchased from KSK Co., Ltd. (Chugai Pharmaceutical Co., Ltd., Tokyo, Japan).

2.3. Oil droplet distribution

Oil droplet distribution was analyzed by laser diffraction using a SALD-3000J (Shimadzu Corporation, Kyoto, Japan). Oil droplet size is reported as mean diameter.

2.4. Cells

Lewis lung carcinoma 3LL cell line was obtained from the Cancer Institute of the Japanese Foundation for Cancer Research (Tokyo, Japan). The 3LL tumor cells were maintained in RPMI-1640 medium supplemented with 10% FCS, 50 µg/mL streptomycin, and 50 U/mL penicillin. To prepare inactivated 3LL cells, the cells were incubated for 20 min at 37°C in culture medium containing 200 µg/mL mitomycin C (Kyowa Hakko Kogyo, Tokyo, Japan), followed by repeated washing with sufficient culture medium. Line 10 hepatocellular carcinoma cells were implanted intraperitoneally, and a range of cell stocks were prepared and stored in liquid nitrogen. In each experiment, the cells were freshly thawed before intradermal inoculation.

2.5. Animals

LEW/Crj male rats and C57BL/6J female mice were purchased from Charles River Japan (Kanagawa, Japan). Strain 2 male guinea pigs were obtained from Japan SLC Inc. (Shizuoka, Japan) and used at 6 weeks of age. All animals were maintained under specific pathogen free conditions, and all animal experiments were conducted according to the guidelines of the Animal Care and Use Committee of Dainippon Sumitomo Pharma.

2.6. Rat IFN-y production

The oil-in-water emulsion of SMP-105 (60 μ g/0.1 mL) with squalane or drakeol was administrated into the back paws of LEW/Crj rats 3 times daily. Blood was withdrawn 6 h after the third administration, and the concentration of rat IFN- γ in the serum was determined by enzyme-linked immunosorbent assay (genzyme TECHNE, Invitrogen Japan, Tokyo, Japan).

2.7. Skin reaction

The oil-in-water emulsion of SMP-105 (30 μ g/0.1 mL) with squalane or drakeol was intradermally administrated once into the backs of Strain 2 guinea pigs. The size of skin reactions was measured at day 7, 13, 21, 27, 34, and 39 after administration.

2.8. DTH reaction

DTH reaction was evaluated as previously described (8,10). In brief, a mixture of inactivated 3LL cells (3 $\times 10^4$ cells) and the vehicle; SMP-105 (12.5 µg), was intradermally administered into the left flank region of C57BL/6J mice twice with a 7-day interval. Seven days after the second administration, inactivated 3LL cells were inoculated at 10⁵ cells in 50 µL HBSS into the left footpads of the mice. Just before, and 24 h after inoculation, the thickness of the left footpad was measured using a dial gauge. Percentage footpad swelling was calculated according to the following equation: Footpad swelling (%) = (thickness of post-injected footpad (mm)) – thickness of pre-injected footpad (mm))/ (thickness of pre-injected footpad (mm)) × 100.

2.9. Antitumor effect in guinea pigs

SMP-105 antitumor effect was evaluated as described previously (7,11). In brief, line 10 hepatoma cells were inoculated intradermally at 1×10^6 cells in 0.1 mL into the right thoracic flank region of guinea pigs. SMP-105 and Immunobladder or vehicle was injected intradermally into sites distal to the site of tumor inoculation on days 0, 7, and 14. Picivanil was injected intradermally into sites distal to the site of tumor inoculation on days 0, 3, 7, 10, and 14. As in clinical use, Aldara cream was applied onto the inoculation sites. Animals were sacrificed by anesthesia with a high concentration of carbon dioxide and the axillary lymph nodes were collected and weighed. For pathological study, metastasis was scored from 0 to 4 based on the area occupied by the tumor cells. Metastasis rate (%) was calculated according to the following equation: Metastasis rate (%) = the number of animals with score 1 to 4 /the total number of animals in the group \times 100. Metastasis Score: 0, no tumor cells observed; 1, tumor cells found as small clusters; 2, clusters composed of a significant number of tumor cells; 3, clusters composed of a large number of tumor cells, some of which are undergoing mitosis; 4, tumor clusters occupy more than half the area of the lymph node.

2.10. Statistical analysis

Results from all experiments are expressed as mean \pm standard deviation (SD). Significant differences in skin reaction, DTH reaction, and antitumor effects were assessed using Dunnett's multiple comparison. Only one data of DTH (Figure 3A) was assessed using T-test.

Statistical analysis was performed using the SAS system for Windows (SAS Institute Inc., Cary, NC, USA).

3. Results

3.1. Immunostimulatory effect of drakeol-based and squalane-based SMP-105 emulsions

Drakeol and squalane are commonly used to emulsify BCG-CWS for clinical and research use. To determine the immunostimulatory effect of drakeol-based and squalane-based SMP-105 emulsions, we measured IFN- γ concentration in serum from rats administered each of the SMP-105 emulsions. As expected, vehicle and SMP-105 dispersed in saline had no effect on IFN- γ production. On the other hand, both SMP-105 emulsions induced equally potent IFN- γ production (Figure 1A).

Because BCG-CWS has the ability to trigger skin reaction, we measured the size of skin reactions (ulcer, edema, and induration) formed after administration of each SMP-105 emulsion. Both SMP-105 emulsions dosedependently and significantly induced skin reactions (Figure 1B). The size of the skin reactions was about the same for both emulsions. These results suggest that drakeol and squalane have similar properties as base material for SMP-105 emulsion.

3.2. Oil droplet size distribution of oil-in-water SMP-105 emulsions

To characterize the prepared SMP-105 emulsions, we

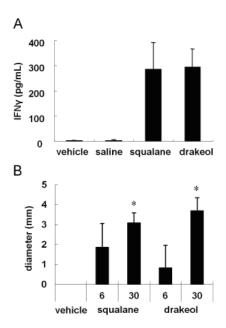


Figure 1. Immunostimulatory effect of drakeol-based and squalane-based SMP-105 emulsions. (A) IFN- γ concentration in serum from rat intradermally administrated each oil based emulsion dispersed in saline. Results are given as means \pm SD of 4-5 mice. (B) Size of skin reactions induced by intradermal administration of each emulsion to guinea pigs. Results are given as means \pm SD of maximum skin reaction size in 5 guinea pigs. * p < 0.05 when compared to vehicle.

measured oil droplet size distribution in each emulsion (Figure 2). Oil droplet size of SMP-105 homogenized oil-in-water emulsion with squalane and drakeol was distributed from sub-micrometer to several tens micrometers with the median about 2-3 μ m (Figure 2A). Both SMP-105 emulsions (squalane-based and drakeol-based) showed the same oil droplet size distribution. The vialed emulsion had an oil droplet size close to that of the homogenized emulsion that was distributed from sub-micrometer to several tens micrometers with the median about 2-3 μ m. On the other hands, the pumped emulsion showed uniform oil droplet size as compared to the homogenized and vialed emulsions (Figure 2B). Oil droplet size in the pumped emulsion differed depending on the pumping frequency (once to 10 times).

3.3. SMP-105 emulsions enhancement of DTH in mouse model

To investigate the immunostimulatory effect of each SMP-105 emulsion, we evaluated DTH reaction with each emulsion injection elicited by 3LL cell in mice. Mice were immunized with inactivated 3LL cell suspension admixed with each emulsion, before being injected with inactivated 3LL alone in the hind

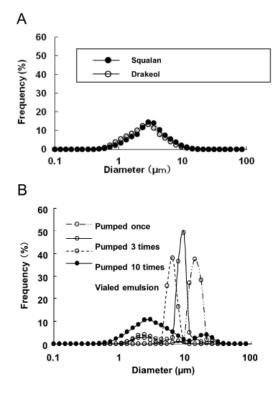


Figure 2. Analysis of oil droplet size distribution in SMP-105 emulsion. Oil droplet size distribution in SMP-105 emulsion was analyzed by laser diffraction particle size analyzer. (A) Squalane-based emulsion (black circle) and drakeol-based emulsion (white circle). (B) Vialed emulsion (black circle) and pumped emulsion (white circle) in diameter by laser diffraction particle size analyzer.

footpad, and edema was measured. As expected, the homogenized emulsion strongly enhanced swelling at the high dose (12.5 μ g) of SMP-105 (Figure 3A). Although both original emulsions evoked strong footpad swelling around the high dose (12.5 μ g), the edema produced by the 10 times pumped emulsion tended to be slightly weaker than that evoked by the vialed emulsion (Figures 3B and 3C). Because the pumped emulsion was composed of approximately the same material as the vialed emulsion, adequate oil droplet size seems to be required for acquired immunity against DTH reactions.

To further investigate oil droplet size, we prepared a vialed emulsion with pumping ten times that showed similar oil droplet size distribution to pumped emulsion and evaluated its induction of DTH. The pumped vialed emulsion enhanced footpad swelling induction but the swelling at highest dose was slightly weaker than that enhanced by the vialed emulsion (Figure 3D). These findings suggest that SMP-105 emulsion oil droplet size affects its immunostimulatory effect.

3.4. Antitumor effect of SMP-105 emulsions in guinea pig metastasis model

Guinea pigs were inoculated with line 10 hepatoma cells and each SMP-105 emulsion: homogenized emulsion, vialed emulsion, (10 times) pumped emulsion or vehicle. Inoculations were carried out at sites different from tumor inoculation site in order to avoid damage to tumor cells by local inflammation.

All emulsions (homogenized, vialed, and pumped) demonstrated prominent antitumor activity at 60 μ g as indicated by in lymph node metastasis rate (Figures 4A and 4C) and metastasis score (Figures 4B and 4D). Growth of primary implanted tumor decreased in some animals about 2 weeks after the first dose of SMP-105.

3.5. Antitumor effect of several TLR agonists in guinea pig model

SMP-105 emulsions enhanced potent DTH reaction and showed antitumor activity. To confirm the antitumor activity of SMP-105, we evaluated the adequacy of the prepared SMP-105 vialed emulsion by comparing its antitumor effect on guinea pig lymph node metastasis to that of marketed TLR agonists (Aldara; TLR7 agonist, OK-432; TLR4 agonist and Immunobladder; live BCG). SMP-105 showed potent antitumor effect with a rate of lymph node metastasis about 20% (Figures 5A and 5B). Aldara inhibited lymph node metastasis by only 14.3%, but reduced the score significantly. Picivanil showed no antitumor effect on the rate of lymph node metastasis (Figures 5C and 5D). On the other hand, Immunobladder showed a dose-dependent potent antitumor effect (Figures 5E and 5F). The maximum effect of Immunobladder was almost equal to that of SMP-105, but required ten times the amount.

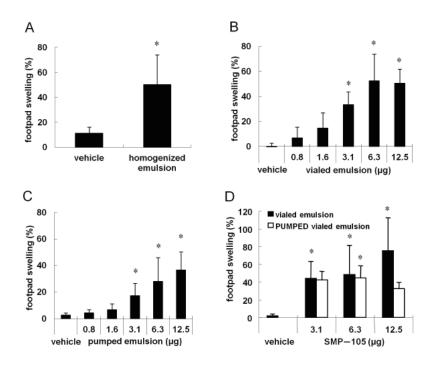


Figure 3. DTH reaction experiment with SMP-105 emulsions. A mixture of inactivated 3LL cells (3 × 10⁴ cells) and each SMP-105 emulsion (0.8-12.5 µg) was intradermally administered into the left flank region of C57BL/6J mice twice at a 7-day interval. (A) Homogenized emulsion of 12.5 µg, (B) vialed emulsion, (C) pumped emulsion at each dose, (D) vialed emulsion and pumped vialed emulsion. Seven days after the second administration, inactivated 3LL cells were inoculated into the left footpads and swelling was monitored by measuring footpad thickness 24 h after inoculation. Relative swelling is calculated as means \pm SD of 6 mice. * p < 0.05 when compared to vehicle.

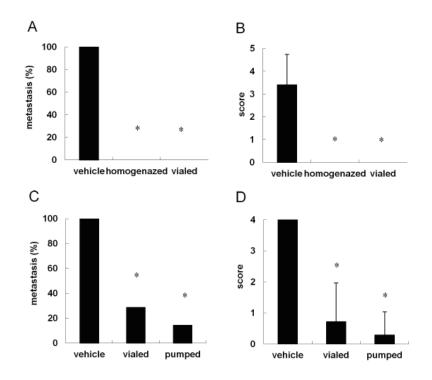


Figure 4. Antitumor effect of SMP-105 emulsions. After inoculation with line 10 hepatoma cells, SMP-105 emulsion or vehicle was injected into the same side as the tumor on days 0, 7, and 14. (A) Rate (%) of lymph node metastasis in the case of homogenized emulsion (60 µg) or vialed emulsion (60 µg). (B) Score (mean \pm SD) of lymph node metastasis in the case of homogenized emulsion (60 µg) or vialed emulsion (60 µg). (C) Rate (%) of lymph node metastasis in the case of vialed emulsion (60 µg) or pumped emulsion (60 µg). (D) Score (mean \pm SD) of lymph node metastasis in the case of vialed emulsion (60 µg) or pumped emulsion (60 µg). (D) Score (mean \pm SD) of lymph node metastasis in the case of vialed emulsion (60 µg) or pumped emulsion (60 µg). Lymph node metastasis rates are given as mean of 4-8 guinea pigs. * p < 0.05 when compared to vehicle.

www.ddtjournal.com

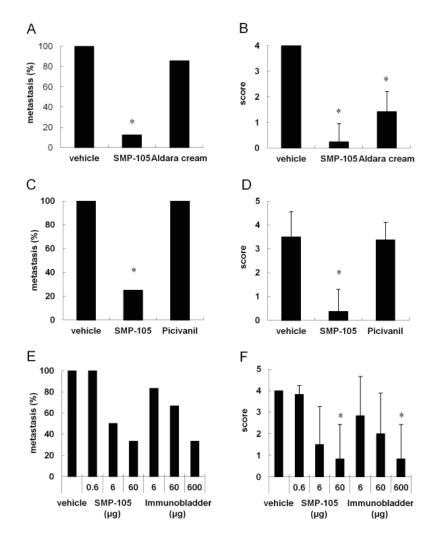


Figure 5. Antitumor effect of SMP-105 emulsion and TLR agonists on line 10 hepatoma in guinea pigs. After inoculation of line 10 hepatoma cells, SMP-105 emulsion, TLR agonist, or vehicle was injected into the same side as the tumor on days 0, 7, and 14. (A) Rate (%) of lymph node metastasis in the case of vialed emulsion (60 µg) or Aldara cream (100 µg). (B) Score (mean \pm SD) of lymph node metastasis in the case of vialed emulsion (60 µg) or Aldara cream (100 µg). (C) Rate (%) of lymph node metastasis in the case of vialed emulsion (60 µg) or Aldara cream (100 µg). (C) Rate (%) of lymph node metastasis in the case of vialed emulsion (60 µg) or Aldara cream (100 µg). (C) Rate (%) of lymph node metastasis in the case of vialed emulsion (6 µg) or Picibanil (2KE). (D) Score (mean \pm SD) of lymph node metastasis in the case of vialed emulsion (6, 6, 60 µg) or Immunobladder (6, 60, 600 µg). (F) Score (mean \pm SD) of lymph node metastasis in the case of vialed emulsion (0.6, 6, 60 µg) or Immunobladder (6, 60, 600 µg). Lymph node metastasis rates are given as mean of 6-8 guinea pigs. * p < 0.05 when compared to vehicle.

4. Discussion

In this study, we prepared original BCS-CWS (SMP-105) emulsions with drakeol and squalane and showed their immunostimulatory effect and antitumor activity in animal models. We successfully generated two types of original SMP-105 formulations that can easily be made into oil-in-water emulsions. With these results, we adopted squalane as base material for SMP-105 emulsion, because of its immunostimulatory activity and its suitable properties for clinical use (Figure 1). Our original squalane-based emulsions, can easily be prepared at use, and successfully enhanced both potent DTH reaction in mouse model and antitumor activity with guinea pig lymph node metastasis model.

In this study, SMP-105 pumped emulsion induced potent footpad swelling but tended to be weaker than

the vialed emulsion in enhancing DTH reaction. Interestingly, when the vialed emulsion was pumped with a connector, edema decreased compared to that observed with the vialed emulsion. We also confirmed roughly emulsified SMP-105 by voltex that showed about the same oil droplet size to once pumped emulsion enhanced DTH reaction with comparable swelling to 10 times pumped emulsion (data not shown). All emulsions were composed of almost the same material, but had different oil droplet size. The fact that both emulsions enhanced DTH reaction indicates that oil droplet size affects immunostimulatory activity. In particular, oil droplet size of the vialed emulsion seems to be an advantage for easy uptake by phagocytes. Oil droplet size distribution in the vialed emulsion ranged from sub-micrometer to over ten micrometers in diameter (Figure 2) with a mean size (about 2-3 µm) close to the adequate size for easy uptake by macrophages (21,22). As previously indicated, SMP-105 requires phagocytosis by macrophages or DCs for immune activation both in vitro and in vivo (10). This indicates that both emulsions prepared in this study, although having different oil droplet size, were engulfed by phagocytes, suggesting that small micro droplet (about 2-3 µm) is easily devoured by phagocytes compared to large micro droplet (10 µm or larger). This consideration is consistent with the findings of other studies that show that several micrometer particles can be taken by phagocytes, but induce significant immune response (21,22,31). Because it is difficult to evaluate SMP-105 emulsion in vitro, we could not show in this study oil droplet uptake quantitatively and directly. Further studies are required to elucidate the mechanism of oil droplet uptake.

Our experiment on DTH reaction showed slight difference in immunostimulatory effect between the prepared emulsions. On the other hand, evaluation of the antitumor effect of both emulsion showed similar effect (Figure 4). Because the difference in footpad swelling between the two emulsions was slight, we could not detect difference in the antitumor effect on lymph node metastasis in guinea pig model. These results indicate that DTH experiment is sensitive enough to evaluate BCG-CWS formulations immunostimulatory effect *in vivo*.

Our group previously reported that SMP-105 is a TLR2 ligand that prevents tumors growth through TLR2 (9). Accordingly, we compare in this study the antitumor effect of SMP-105 emulsion to that of three launched TLR agonists (Aldara, Picibanil, and Immunobladder) (Figure 5). Although Aldara and Picibanil were administrated in sufficient amount and adequate route as indicated in drug package insert, they showed limited antitumor effect (Figure 5). SMP-105 emulsion showed potent antitumor effect equivalent to that of Immunobladder at low doses. This finding indicates that SMP-105 has promising clinical antitumor use.

Overall, we showed that squalane can be appropriate base material for SMP-105 emulsion and produced two types of squalane-based formulations that can be easily made into oil-in-water emulsions at the time of administration. The results of the prepared emulsions oil droplet distribution and DTH experiment suggest that small micrometer droplet (about 2-3 µm) can induce more potent immune reaction than large micrometer droplet (10 µm or larger). It is believed that small micrometer droplet is easily engulfed by phagocytes. This study is first to show that oil droplet size of few micrometer is optimal for SMP-105 microemulsion with squalane. We also showed for the first time that SMP-105 has more potent antitumor effect than launched TLR agonists in guinea pig metastasis model. These findings indicate that SMP-105 is a promising candidate for clinical investigation.

Acknowledgements

We thank Dr. Takehiko Nomura for preparing SMP-105 and its emulsions.

References

- Sharma P, Wagner K, Wolchok JD, Allison JP. Novel cancer immunotherapy agents with survival benefit: Recent successes and next steps. Nat Rev Cancer. 11:805-812.
- Begum NA, Ishii K, Kurita-Taniguchi M, Tanabe M, Kobayashi M, Moriwaki Y, Matsumoto M, Fukumori Y, Azuma I, Toyoshima K, Seya T. Mycobacterium bovis BCG cell wall-specific differentially expressed genes identified by differential display and cDNA subtraction in human macrophages. Infect Immun. 2004; 72:937-948.
- Kodama K, Higashiyama M, Takami K, Oda K, Okami J, Maeda J, Akazawa T, Matsumoto M, Seya T, Wada M, Toyoshima K. Innate immune therapy with a Bacillus Calmette-Guerin cell wall skeleton after radical surgery for non-small cell lung cancer: A case-control study. Surg Today. 2009; 39:194-200.
- Azuma I, Ribi EE, Meyer TJ, Zbar B. Biologically active components from mycobacterial cell walls. I. Isolation and composition of cell wall skeleton and component P3. J Natl Cancer Inst. 1974; 52:95-101.
- Matsumoto M, Seya T, Kikkawa S, *et al.* Interferon gamma-producing ability in blood lymphocytes of patients with lung cancer through activation of the innate immune system by BCG cell wall skeleton. Int Immunopharmacol. 2001; 1:1559-1569.
- Hayashi D, Takii T, Fujiwara N, Fujita Y, Yano I, Yamamoto S, Kondo M, Yasuda E, Inagaki E, Kanai K, Fujiwara A, Kawarazaki A, Chiba T, Onozaki K. Comparable studies of immunostimulating activities *in vitro* among *Mycobacterium bovis* bacillus Calmette-Guerin (BCG) substrains. FEMS Immunol Med Microbiol. 2009; 56:116-128.
- Kashiwazaki Y, Murata M, Fujii T, Nakagawa M, Fukushima A, Chiba N, Azuma I, Yamaoka T. Immune response against cell-wall skeleton of *Mycobacterium bovis* BCG at the inoculation site and peripheral lymphoid organs. Drug Discov Ther. 2008; 2:178-187.
- Miyauchi M, Murata M, Shibuya K, Koga-Yamakawa E, Uenishi Y, Kusunose N, Sunagawa M, Yano I, Kashiwazaki Y. Arabino-mycolates derived from cell-wall skeleton of *Mycobacterium bovis* BCG as a prominent structure for recognition by host immunity. Drug Discov Ther. 2011; 5:130-135.
- Murata M. Activation of Toll-like receptor 2 by a novel preparation of cell wall skeleton from *Mycobacterium bovis* BCG Tokyo (SMP-105) sufficiently enhances immune responses against tumors. Cancer Sci. 2008; 99:1435-1440.
- Miyauchi M, Murata M, Shibuya K, Koga-Yamakawa E, Yanagawa Y, Azuma I, Kashiwazaki Y. Phagocytosis plays a dual role in activating dendritic cells; digestive production of active Toll-like receptor ligands and cooperation with Toll-like receptor signaling. Drug Discov Ther. 2010; 4:135-143.
- Kashiwazaki Y, Murata M, Sato T, Miyauchi M, Nakagawa M, Fukushima A, Chiba N, Azuma I, Yamaoka T. Injection of cell-wall skeleton of *Mycobacterium bovis*

BCG draining to a sentinel lymph node eliminates both lymph node metastases and the primary transplanted tumor. Drug Discov Ther. 2008; 2:168-177.

- Okuyama H, Takeda J, Onoe K, Morikawa K. Histological studies on adjuvanticity of BCG-Cell walls: Comparison of adjuvanticity between oil-in-water and water-in-oil forms. Bulletin of the Institute of Immunological Science, Hokkaido University. 1978; 38:13-23.
- Hayashi A. Interferon-gamma as a marker for the effective cancer immnotherapy with BCG-Cell wall skeleton. Proc Jpn Acad. 1994; 70:205-209.
- Yoo YC, Hata K, Lee KB, Azuma I. Inhibitory effect of BCG cell-wall skeletons (BCG-CWS) emulsified in squalane on tumor growth and metastasis in mice. Arch Pharm Res. 2002; 25:522-527.
- Yarkoni E, Rapp HJ. Influence of type of oil and surfactant concentration on the efficacy of emulsified *Mycobacterium bovis* BCG cell walls to induce tumor regression in guinea pigs. Infect Immun. 1980; 28:881-886.
- Kahn JO, Sinangil F, Baenziger J, Murcar N, Wynne D, Coleman RL, Stenimer KS, Dekker CL, Chernoff D. Clinical and immunologic responses to human immunodeficiency virus (HIV) type 1SF2 gp120 subunit vaccine combined with MF59 adjuvant with or without muramyl tripeptide dipalmitoyl phosphatidylethanolamine in non-HIV-infected human volunteers. J Infect Dis. 1994; 170:1288-1291.
- Langenberg AG, Burke RL, Adair SF, Sekulovich R, Tigges M, Dekker CL, Corey L. A recombinant glycoprotein vaccine for herpes simplex virus type 2: Safety and immunogenicity [corrected]. Ann Intern Med. 1995; 122:889-898.
- Pass RF, Duliege AM, Boppana S, Sekulovich R, Percell S, Britt W, Burke RL. A subunit cytomegalovirus vaccine based on recombinant envelope glycoprotein B and a new adjuvant. J Infect Dis. 1999; 180:970-975.
- Nitayaphan S, Khamboonruang C, Sirisophana N, *et al.* A phase I/II trial of HIV SF2 gp120/MF59 vaccine in seronegative thais. Vaccine. 2000; 18:1448-1455.
- Schultze V, D'Agosto V, Wack A, Novicki D, Zorn J, Hennig R. Safety of MF59[™] adjuvant. Vaccine. 2008; 26:3209-3222.
- 21. Tabata Y, Ikada Y. Effect of the size and surface charge of polymer microspheres on their phagocytosis by macrophage. Biomaterials. 1988; 9:356-362.
- 22. Ayhan H, Tuncel A, Bor N, Piskin E. Phagocytosis of monosize polystyrene-based microspheres having

different size and surface properties. J Biomater Sci Polym Ed. 1995; 7:329-342.

- Dupuis M, Murphy TJ, Higgins D, Ugozzoli M, van Nest G, Ott G, McDonald DM. Dendritic cells internalize vaccine adjuvant after intramuscular injection. Cell Immunol. 1998; 186:18-27.
- Tabata Y, Ikada Y. Macrophage phagocytosis of biodegradable microspheres composed of L-lactic acid/ glycolic acid homo- and copolymers. J Biomed Mater Res. 1988; 22:837-858.
- Eldridge JH, Staas JK, Meulbroek JA, McGhee JR, Tice TR, Gilley RM. Biodegradable microspheres as a vaccine delivery system. Mol Immunol. 1991; 28:287-294.
- 26. Thiele L, Rothen-Rutishauser B, Jilek S, Wunderli-Allenspach H, Merkle HP, Walter E. Evaluation of particle uptake in human blood monocyte-derived cells *in vitro*. Does phagocytosis activity of dendritic cells measure up with macrophages? J Control Release. 2001; 76:59-71.
- Jiang W, Gupta RK, Deshpande MC, Schwendeman SP. Biodegradable poly(lactic-co-glycolic acid) microparticles for injectable delivery of vaccine antigens. Adv Drug Deliv Rev. 2005; 57:391-410.
- O'Hagan DT, Singh M. Microparticles as vaccine adjuvants and delivery systems. Expert Rev Vaccines. 2003; 2:269-283.
- Men Y, Audran R, Thomasin C, Eberl G, Demotz S, Merkle HP, Gander B, Corradin G. MHC class Iand class II-restricted processing and presentation of microencapsulated antigens. Vaccine. 1999; 17:1047-1056.
- Carcaboso AM, Hernandez RM, Igartua M, Rosas JE, Patarroyo ME, Pedraz JL. Potent, long lasting systemic antibody levels and mixed Th1/Th2 immune response after nasal immunization with malaria antigen loaded PLGA microparticles. Vaccine. 2004; 22:1423-1432.
- Thomas C, Gupta V, Ahsan F. Particle size influences the immune response produced by hepatitis B vaccine formulated in inhalable particles. Pharm Res. 2010; 27:905-919.
- Uenishi Y, Okada T, Okabe S, Sunagawa M. Study on the cell wall skeleton derived from Mycobacterium bovis BCG Tokyo 172 (SMP-105): Establishment of preparation and analytical methods. Chem Pharm Bull (Tokyo). 2007; 55:843-852.

(Received June 19, 2012; Revised July 31, 2012; Accepted August 4, 2012)

Commentary

Animal welfare and use of silkworm as a model animal

Nobukazu Sekimizu^{1,*}, Atmika Paudel², Hiroshi Hamamoto²

¹ Genome Pharmaceuticals Institute Co., Ltd., Tokyo, Japan;

² Laboratory of Microbiology, Graduate School of Pharmaceutical Sciences, The University of Tokyo, Tokyo, Japan.

ABSTRACT: Sacrificing model animals is required for developing effective drugs before being used in human beings. In Japan today, at least 4,210,000 mice and other mammals are sacrificed to a total of 6,140,000 per year for the purpose of medical studies. All the animals treated in Japan, including test animals, are managed under control of "Act on Welfare and Management of Animals". Under the principle of this Act, no person shall kill, injure, or inflict cruelty on animals without due cause. "Animal" addressed in the Act can be defined as a "vertebrate animal". If we can make use of invertebrate animals in testing instead of vertebrate ones, that would be a remarkable solution for the issue of animal welfare. Furthermore, there are numerous advantages of using invertebrate animal models: less space and small equipment are enough for taking care of a large number of animals and thus are cost-effective, they can be easily handled, and many biological processes and genes are conserved between mammals and invertebrates. Today, many invertebrates have been used as animal models, but silkworms have many beneficial traits compared to mammals as well as other insects. In a Genome Pharmaceutical Institute's study, we were able to achieve a lot making use of silkworms as model animals. We would like to suggest that pharmaceutical companies and institutes consider the use of the silkworm as a model animal which is efficacious both for financial value by cost cutting and ethical aspects in animals' welfare.

Keywords: Ethical issue, alternatives, developing drugs, medical studies, 3R, test animals, cost

*Address correspondence to:

E-mail: nsekimizu@genome-pharm.jp

It is said to be unavoidable to sacrifice model animals for developing effective drugs (1) before being used in human beings. However, use of mammalian animals has not been convenient due to high costs, long breeding times, and large and sophisticated space requirements. Another major limiting factor is ethical issues associated with the use of mammalian models (2). Various alternatives are being developed such as replacing mammals with cultured cells, but the effect of those is still limited (3). Therefore in Japan today, at least 4,210,000 mice and other mammals such as rats, guinea pigs, rabbits, dogs, cats, and monkeys are sacrificed to a total of 6,140,000 per year for the purpose of Medical studies (4).

All the animals treated in Japan, including test animals, are managed under control of "Act on Welfare and Management of Animals" enacted in 1973. The fundamental principle of this law is provided as "In light of the fact that animals are living beings, no person shall kill, injure, or inflict cruelty on animals without due cause, and every person shall treat animals properly by taking into account their natural habits and giving consideration to the symbiosis between humans and animals." (Article 2). This principle indicates that inflicting animals more pain than for inevitable tests is forbidden, even in necessary animal testing (Article 44) (5). It is required to minimize both numbers of test animals and the pain given to them. This principle has been basic and traditional in Western culture, and was already proposed in 1959, as the "3R" principle by W. M. Russell and by R. L. Burch in the UK (6). The three Rs are: "Replacement" - replacing the way of testing without test animals, "Reduction" - reducing the number of them, and "Refinement" - minimizing the quality and the intensity of pain given to animals to the least amount. "Animal" addressed in the principle can be defined as a "vertebrate animal" (7). In Japan, this principle was reflected in amendment of "Act on Welfare and Management of Animals", Article 44 in 2005.

If we can make use of invertebrate animals in testing instead of vertebrate ones, that would be a remarkable solution for the issue of animal welfare. There are numerous advantages of using invertebrate animal models: less space and small equipment are enough for taking care of a large number of animals and thus are

Dr. Nobukazu Sekimizu, Genome Pharmaceuticals Institute Co., Ltd., Laboratory 401, The University of Tokyo Entrepreneur Plaza, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-0033, Japan.

cost-effective, they can be easily handled, there are less ethical issues surrounding their use, and many biological processes and genes are conserved between mammals and invertebrates. Many invertebrates have been used as animal models: fruit fly Drosophila melanogaster (8,9), grasshopper Romalea microptera (10), wax moth larva Galleria mellonella (11), honey bee Apis mellifera (12), and silkworm larva Bomboyx mori (13). Silkworms have many beneficial traits compared to mammals as well as other insects. Silkworms are not only easy for injection experiments but also they can be injected either through the intrahemolymph route that corresponds to intravenous in humans or the intramidgut route that corresponds to oral in humans. Figure 1 shows the easy ways of injecting silkworms through these routes. Unlike D. melanogaster, the silkworm has a large enough body size to carry out experiments for accurate dose administration and organ isolation. While injection into Drosophila requires special techniques and even though female flies are larger, they however have a hard outer surface which makes them difficult for injection. Unlike

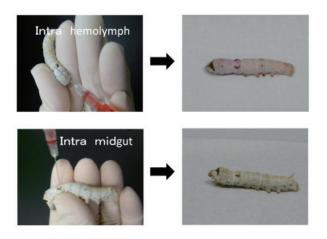


Figure 1. Injection into hemolymph and midgut. When injection angle is shallow, red ink is injected into blood (**Upper**). On the other hand, when injection angle is deep, it is injected into midgut and silkworm is not stained (**Lower**).

Table 1. Comparision of silkworm with other insects

R. microptera and G. mellonella, the silkworm has an established method of breeding as it has been used for silk for over five thousand years. Unlike honey-bees, silkworms do not bite and even the adult moth cannot fly. The larvae do not have any sharp hair or horns that sting, so they are not harmful and do not require special techniques and caution for use. Moreover, the locomotion of silkworm larvae is slow which ensures that it cannot escape away from the laboratory setting easily which minimizes the risk of biological hazards. They can be fed an artificial diet that is easily available and can be bred all year round so that there is no shortage of larvae for experiments. The established rearing method allows the same kind of larval breeds which is very important for reproducible results. Larvae molt four times, it is easy to separate each instar larvae and they are stable having little individual genetic differences that give uniformity in research results. The whole genome is now known which allows for genetic manipulations and there are already many ongoing research projects that utilize transgenic and genetically modified silkworms.

Different models of the silkworm have already been established: bacterial infection model (13, 14), baculoviral infection model (15), model to test innate immunity (16-18), diabetic model (19), bacterial virulence model (20-22), model to test pharmacokinetics (23), and model to test toxicity and metabolism (24). Most of these research projects have proven the correlation between results in mammals and silkworms. Table 1 distinguishes various features of the silkworm with other insects.

In the Genome Pharmaceutical Institute's study, we were able to achieve a lot making use of silkworms as model animals. Not only did we discover new effective chemicals including a new antibiotic "Kaikosin", but also we reduced the number of mice (small mammals in standards of test animals' size) and that helps towards making a solution for problems in animals' welfare, and cutting costs of testing. We are using silkworms instead of mice in the initial screening and testing in

| Items | Silkworm | Drosophila | Honey bee | Waxmoth | Grass hopper |
|--|---|---|------------------|---|--------------|
| Size | 40-60 mm | 1-3 mm | 15-17 mm | 30-40 mm | 60-80 mm |
| Breeding method | Well established (> 5000 years) | Well established | Well established | Established | Established |
| Locomotion | Larva: slow, Adult: cannot fly | Flies | Flies | Larva: faster than silkworm, Adult: flies | Jumps, flies |
| Special handling technique | Not required | Required | Required | Not required | Required |
| Chance of biohazard | Less | Higher | Higher | Higher than silkworm, less than others | Higher |
| Injection technique | Easier, anyone can learn within couple of hours | Difficult, requires skilled personnel | | Easier | |
| Isolation of organs | Easier | Difficult ,not always possible | Easier | Easier | Easier |
| Route of administration/ Accuracy of administrated dosage | Oral, injection to dorsal surface: intrahemolymph, intramidgut/ accurate in case of injection | Oral, injection to dorsal surface, not accurate | | Oral, topical, injection to ventral surface/ accurate in case of injection | - |
| Diseases models | Many | Many | | Few | Few |

| Experimental models | 10,000 chemical compounds | 15,000 natural compounds |
|--------------------------------------|--|--|
| Our testing method General method | 100 mice + 30,000 silkworms 30,000 mice | 150 mice + 45,000 silkworms 45,000 mice |
| 'Reduction' effectiveness of mice | $30,000 \rightarrow 100$ mice | $45,000 \rightarrow 150$ mice |

Table 2. Experimental models used in screening antimicrobial agents from 10,000 chemical compounds and 15,000 natural compounds

our company which is said to be a 'Replacement'. We used silkworms for screening chemical compounds as well as natural compounds. In screening for a curative effect with 10,000 chemical compounds' samples and 15,000 samples of products from soil bacteria (natural compounds) with antimicrobial activity, the number of mice used as test animals in our testing method is obviously less than that in the general method which here refers to tests in a mouse model directly after *in-vitro* analysis. We used silkworms after *in-vitro* analysis and before testing in the mouse model, and thus we could decrease the number of mice used for the tests. This decrement in the number of test mice is the 'Reduction' effectiveness (Table 2).

As shown in Table 2, we have gained a remarkable result in reduction of the number of mice in testing, by replacing them with silkworms in the initial testing. Having these results, we have been suggesting to pharmaceutical companies and institutes to consider the use of our silkworm related technologies, which is efficacious, with both a financial value by cost cutting and ethical aspects in animals' welfare. If we can replace 10% of tests run using mice as test animals in Japan, that means at least 400,000 mice can be saved from testing per year.

We should be aware of the fact that in Europe, more strict regulation, Cosmetics Directive 76/768/ EEC was enacted in 2003. Dr. Tsutomu Kurosawa, the president of the society refers to the directive as "One of the epochs of our activities was the 7th amendment of EU cosmetics Directives in 2003. This directive ordered the total abundance of animal experimentation for cosmetic development and trade." (25). Based on this regulation, animal testing is gradually banned for developing materials and products for cosmetics, and products, subject to regulation of this Directive are already banned for sales in the EU. Some of the whole tests are still run, since a substitute way of testing without use of animals cannot be found, and it will take some more years for total abolishment. However, it won't be long before the day cosmetics produced using animal testing will be banned from sales in the EU. This is the global trend, and in the near future, animal usage alternatives or use of creatures, not controlled by the concept of animal welfare will be more valuable. With these issues, the silkworm is thus an ideal living creature for testing and can be counted on as a reliable test animal having great potential without animal welfare regulations.

References

- Ohno Y. Alternatives to animal experiments in pharmacology. Nihon Yakurigaku Zasshi. 2011; 138:99-102.
- Baumans V. Use of animals in experimental research: An ethical dilemma? Gene Ther. 2004; 11(Suppl 1):S64-S66.
- Kagiyama N. Animal welfare and laboratory animal science. Bulletin of Gakushikai. 2008; 872:79-84.
- 4. Japanese association of laboratory animal resources, Data of sales amount of testing animals in 2010, 2011, p.4
- Act on Welfare and Management of Animals (Japan), Article 44, 1973.
- Russell WMS, Burch RL. The Principles of Humane Experimental Technique. http://altweb.jhsph.edu/pubs/ books/humane_exp/addendum (accessed May 26, 2012).
- Ohno Y. Importance of research on alternatives to animal experiments and its current issues: Animal experiments in The Japanese Pharmacological Society. Nihon Yakurigaku Zasshi. 2005; 125:325-329.
- Lemaitre B, Nicolas E, Michaut L, Reichhart JM, Hoffmann JA. The dorsoventral regulatory gene cassette *spätzle/Toll/cactus* controls the potent antifungal response in Drosophila adults. Cell. 1996; 86:973-983.
- Bernal A, Kimbrell DA. Drosophila Thor participates in host immune defense and connects a translational regulator with innate immunity. Proc Natl Acad Sci U S A. 2000; 97:6019-6024.
- Johny S, Lange CE, Solter LF, Merisko A, Whitman DW. New insect system for testing antibiotics. J Parasitol. 2007; 93:1505-1511.
- Champion OL, Cooper IAM, James SL, Ford D, Karlyshev A, Wren BW, Duffield M, Oyston PCF, Titball RW. Galleria mellonella as an alternative infection model for *Yersinia pseudotuberculosis*. Microbiology. 2009; 155:1516-1522.
- Dussaubat C, Brunet JL, Higes M, Colbourne JK, Lopez J, Choi JH, Martín-Hernández R, Botías C, Cousin M, McDonnell C, Bonnet M, Belzunces LP, Moritz RF, Le Conte Y, Alaux C. Gut pathology and responses to the microsporidium *Nosema ceranae* in the honey bee *Apis mellifera*. PLoS One. 2012; 7:e37017.
- Kaito C, Akimitsu N, Watanabe H, Sekimizu K. Silkworm larvae as an animal model of bacterial infection pathogenic to humans. Microb Pathog. 2002; 32:183-190.
- Hamamoto H, Kurokawa K, Kaito C, Kamura K, Razanajatovo IM, Kusuhara H, Santa T, Sekimizu K. Quantitative evaluation of the therapeutic effects of antibiotics using silkworms infected with human pathogenic microorganisms. Antimicrob Agents Chemother. 2004; 48:774-779.
- 15. Orihara Y, Hamamoto H, Kasuga H, Shimada T, Kawaguchi Y, Sekimizu K. A silkworm-baculovirus model for assessing the therapeutic effects of antiviral compounds: Characterization and application to the

isolation of antivirals from traditional medicines. J Gen Virol. 2008; 89:188-194.

- Fujiyuki T, Hamamoto H, Ishii K, Urai M, Kataoka K, Takeda T, Shibata S, Sekimizu K. Evaluation of innate immune stimulating activity of polysaccharides using a silkworm (*Bombyx mori*) muscle contraction assay. Drug Discov Ther. 2012; 6:88-93.
- Ishii K, Hamamoto H, Kamimura M, Sekimizu K. Activation of the silkworm cytokine by bacterial and fungal cell wall components *via* a reactive oxygen species-triggered mechanism. J Biol Chem. 2008; 283:2185-2191.
- Ishii K, Hamamoto H, Kamimura M, Nakamura Y, Noda H, Imamura K, Mita K, Sekimizu K. Insect cytokine paralytic peptide (PP) induces cellular and humoral immune responses in the silkworm *Bombyx mori*. J Biol Chem. 2010; 285:28635-28642.
- Matsumoto Y, Sumiya E, Sugita T, Sekimizu K. An invertebrate hyperglycemic model for the identification of anti-diabetic drugs. PLoS One. 2011; 30:e18292.
- 20. Kurokawa K, Kaito C, Sekimizu K. Two-component signaling in the virulence of *Staphylococcus aureus*: A

silkworm larvae-pathogenic agent infection model of virulence. Methods Enzymol. 2007; 422:233-244.

- Hanada Y, Sekimizu K, Kaito C. Silkworm apolipophorin protein inhibits *Staphylococcus aureus* virulence. J Biol Chem. 2011; 286:39360-39369.
- Miyazaki S, Matsumoto Y, Sekimizu K, Kaito C. 2012. Evaluation of *Staphylococcus aureus* virulence factors using a silkworm model. FEMS Microbiol Lett. 2012; 326:116-124.
- Asami Y, Horie R, Hamamoto H, Sekimizu K. Use of silkworms for identification of drug candidates having appropriate pharmacokinetics from plant sources. BMC Pharmacol. 2010; 10:7.
- Hamamoto H, Tonoike A, Narushima K, Horie R, Sekimizu K. Silkworm as a model animal to evaluate drug candidate toxicity and metabolism. Comp Biochem Physiol C Toxicol Pharmacol. 2009; 149:334-339.
- Japanese Society for Alternative to Animal Experiments. http://www.asas.or.jp/jsaae/kaichoaisatu.html (accessed May 26, 2012).

(Received August 11, 2012; Accepted August 13, 2012)



Guide for Authors

1. Scope of Articles

Drug Discoveries & Therapeutics welcomes contributions in all fields of pharmaceutical and therapeutic research such as medicinal chemistry, pharmacelogy, pharmaceutical analysis, pharmaceutics, pharmaceutical administration, and experimental and clinical studies of effects, mechanisms, or uses of various treatments. Studies in drugrelated fields such as biology, biochemistry, physiology, microbiology, and immunology are also within the scope of this journal.

2. Submission Types

Original Articles should be well-documented, novel, and significant to the field as a whole. An Original Article should be arranged into the following sections: Title page, Abstract, Introduction, Materials and Methods, Results, Discussion, Acknowledgments, and References. Original articles should not exceed 5,000 words in length (excluding references) and should be limited to a maximum of 50 references. Articles may contain a maximum of 10 figures and/or tables.

Brief Reports definitively documenting either experimental results or informative clinical observations will be considered for publication in this category. Brief Reports are not intended for publication of incomplete or preliminary findings. Brief Reports should not exceed 3,000 words in length (excluding references) and should be limited to a maximum of 4 figures and/or tables and 30 references. A Brief Report contains the same sections as an Original Article, but the Results and Discussion sections should be combined.

Reviews should present a full and up-to-date account of recent developments within an area of research. Normally, reviews should not exceed 8,000 words in length (excluding references) and should be limited to a maximum of 100 references. Mini reviews are also accepted.

Policy Forum articles discuss research and policy issues in areas related to life science such as public health, the medical care system, and social science and may address governmental issues at district, national, and international levels of discourse. Policy Forum articles should not exceed 2,000 words in length (excluding references).

Case Reports should be detailed reports of the symptoms, signs, diagnosis, treatment, and follow-up of an individual patient. Case reports may contain a demographic profile of the patient but usually describe an unusual or novel occurrence. Unreported or unusual side effects or adverse interactions involving medications will also be considered. Case Reports should not exceed 3,000 words in length (excluding references).

News articles should report the latest events in health sciences and medical research from around the world. News should not exceed 500 words in length.

Letters should present considered opinions in response to articles published in Drug Discoveries & Therapeutics in the last 6 months or issues of general interest. Letters should not exceed 800 words in length and may contain a maximum of 10 references.

3. Editorial Policies

Ethics: Drug Discoveries & Therapeutics requires that authors of reports of investigations in humans or animals indicate that those studies were formally approved by a relevant ethics committee or review board.

Conflict of Interest: All authors are required to disclose any actual or potential conflict of interest including financial interests or relationships with other people or organizations that might raise questions of bias in the work reported. If no conflict of interest exists for each author, please state "There is no conflict of interest to disclose".

Submission Declaration: When a manuscript is considered for submission to Drug Discoveries & Therapeutics, the authors should confirm that 1) no part of this manuscript is currently under consideration for publication elsewhere; 2) this manuscript does not contain the same information in whole or in part as manuscripts that have been published, accepted, or are under review elsewhere, except in the form of an abstract, a letter to the editor, or part of a published lecture or academic thesis; 3) authorization for publication has been obtained from the authors' employer or institution; and 4) all contributing authors have agreed to submit this manuscript.

Cover Letter: The manuscript must be accompanied by a cover letter signed by the corresponding author on behalf of all authors. The letter should indicate the basic findings of the work and their significance. The letter should also include a statement affirming that all authors concur with the submission and that the material submitted for publication has not been published previously or is not under consideration for publication elsewhere. The cover letter should be submitted in PDF format. For example of Cover Letter, please visit http://www.ddtjournal.com/downcentre.php (Download Centre).

Copyright: A signed JOURNAL PUBLISHING AGREEMENT (JPA) must be provided by post, fax, or as a scanned file before acceptance of the article. Only forms with a hand-written signature are accepted. This copyright will ensure the widest possible dissemination of information. A form facilitating transfer of copyright can be downloaded by clicking the appropriate link and can be returned to the e-mail address or fax number noted on the form (Please visit Download Centre). Please note that your manuscript will not proceed to the next step in publication until the JPA form is received. In addition, if excerpts from other copyrighted works are included, the author(s) must obtain written permission from the copyright owners and credit the source(s) in the article.

Suggested Reviewers: A list of up to 3 reviewers who are qualified to assess the scientific merit of the study is welcomed. Reviewer information including names, affiliations, addresses, and e-mail should be provided at the same time the manuscript is submitted online. Please do not suggest reviewers with known conflicts of interest, including participants or anyone with a stake in the proposed research; anyone from the same institution: former students, advisors, or research collaborators (within the last three years); or close personal contacts. Please note that the Editor-in-Chief may accept one or more of the proposed reviewers or may request a review by other qualified persons.

Language Editing: Manuscripts prepared by authors whose native language is not English should have their work proofread by a native English speaker before submission. If not, this might delay the publication of your manuscript in Drug Discoveries & Therapeutics.

The Editing Support Organization can provide English proofreading, Japanese-English translation, and Chinese-English translation services to authors who want to publish in Drug Discoveries & Therapeutics and need assistance before submitting a manuscript. Authors can visit this organization directly at http://www.iacmhr. com/iac-eso/support.php?lang=en. IAC-ESO was established to facilitate manuscript preparation by researchers whose native language is not English and to help edit works intended for international academic journals.

4. Manuscript Preparation

Manuscripts should be written in clear, grammatically correct English and submitted as a Microsoft Word file in a single-column format. Manuscripts must be paginated and typed in 12-point Times New Roman font with 24-point line spacing. Please do not embed figures in the text. Abbreviations should be used as little as possible and should be explained at first mention unless the term is a well-known abbreviation (*e.g.* DNA). Single words should not be abbreviated.

Title page: The title page must include 1) the title of the paper (Please note the title should be short, informative, and contain the major key words); 2) full name(s) and affiliation(s) of the author(s), 3) abbreviated names of the author(s), 4) full name, mailing address, telephone/fax numbers, and e-mail address of the corresponding author; and 5) conflicts of interest (if you have an actual or potential conflict of interest to disclose, it must be included as a footnote on the title page of the manuscript; if no conflict of interest exists for each author, please state "There is no conflict of interest to disclose"). Please visit Download Centre and refer to the title page of the manuscript sample.

Abstract: A one-paragraph abstract consisting of no more than 250 words must be included. The abstract should briefly state the purpose of the study, methods, main findings, and conclusions. Abbreviations must be kept to a minimum and non-standard abbreviations explained in brackets at first mention. References should be avoided in the abstract. Key words or phrases that do not occur in the title should be included in the Abstract page.

Introduction: The introduction should be a concise statement of the basis for the study and its scientific context.

Materials and Methods: The description should be brief but with sufficient detail to enable others to reproduce the experiments. Procedures that have been published previously should not be described in detail but appropriate references should simply be cited. Only new and significant modifications of previously published procedures require complete description. Names of products and manufacturers with their locations (city and state/country) should be given and sources of animals and cell lines should always be indicated. All clinical investigations must have been conducted in accordance with Declaration of Helsinki principles. All human and animal studies must have been approved by the appropriate institutional review board(s) and a specific declaration of approval must be made within this section.

Results: The description of the experimental results should be succinct but in sufficient detail to allow the experiments to be analyzed and interpreted by an independent reader. If necessary, subheadings may be used for an orderly presentation. All figures and tables must be referred to in the text.

Discussion: The data should be interpreted concisely without repeating material already presented in the Results section. Speculation is permissible, but it must be well-founded, and discussion of the wider implications of the findings is encouraged. Conclusions derived from the study should be included in this section.

Acknowledgments: All funding sources should be credited in the Acknowledgments section. In addition, people who contributed to the work but who do not meet the criteria for authors should be listed along with their contributions.

References: References should be numbered in the order in which they appear in the text. Citing of unpublished results, personal communications, conference abstracts, and theses in the reference list is not recommended but these sources may be mentioned in the text. In the reference list, cite the names of all authors when there are fifteen or fewer authors; if there are sixteen or more authors, list the first three followed by *et al.* Names of journals should be abbreviated in the style used in PubMed. Authors are responsible for the accuracy of the references. Examples are given below: *Example 1* (Sample journal reference): Nakata M, Tang W. Japan-China Joint Medical Workshop on Drug Discoveries and Therapeutics 2008: The need of Asian pharmaceutical researchers' cooperation. Drug Discov Ther. 2008; 2:262-263.

Example 2 (Sample journal reference with more than 15 authors):

Darby S, Hill D, Auvinen A, *et al.* Radon in homes and risk of lung cancer: Collaborative analysis of individual data from 13 European case-control studies. BMJ. 2005; 330:223.

Example 3 (Sample book reference): Shalev AY. Post-traumatic stress disorder: Diagnosis, history and life course. In: Post-traumatic Stress Disorder, Diagnosis, Management and Treatment (Nutt DJ, Davidson JR, Zohar J, eds.). Martin Dunitz, London, UK, 2000; pp. 1-15.

Example 4 (Sample web page reference): World Health Organization. The World Health Report 2008 – primary health care: Now more than ever. *http://www.who.int/whr/2008/ whr08_en.pdf* (accessed September 23, 2010).

Tables: All tables should be prepared in Microsoft Word or Excel and should be arranged at the end of the manuscript after the References section. Please note that tables should not in image format. All tables should have a concise title and should be numbered consecutively with Arabic numerals. If necessary, additional information should be given below the table.

Figure Legend: The figure legend should be typed on a separate page of the main manuscript and should include a short title and explanation. The legend should be concise but comprehensive and should be understood without referring to the text. Symbols used in figures must be explained.

Figure Preparation: All figures should be clear and cited in numerical order in the text. Figures must fit a one- or two-column format on the journal page: 8.3 cm (3.3 in.) wide for a single column, 17.3 cm (6.8 in.) wide for a double column; maximum height: 24.0 cm (9.5 in.). Please make sure that artwork files are in an acceptable format (TIFF or JPEG) at minimum resolution (600 dpi for illustrations, graphs, and annotated artwork, and 300 dpi for micrographs and photographs). Please provide all figures as separate files. Please note that low-resolution images are one of the leading causes of article resubmission and schedule delays. All color figures will be reproduced in full color in the online edition of the journal at no cost to authors

Units and Symbols: Units and symbols conforming to the International System of Units (SI) should be used for physicochemical quantities. Solidus notation (*e.g.* mg/kg, mg/mL, mol/mm²/min) should be used. Please refer to the SI Guide www.bipm.org/en/si/ for standard units.

Supplemental data: Supplemental data might be useful for supporting and enhancing your scientific research and

Drug Discoveries & Therapeutics accepts the submission of these materials which will be only published online alongside the electronic version of your article. Supplemental files (figures, tables, and other text materials) should be prepared according to the above guidelines, numbered in Arabic numerals (e.g., Figure S1, Figure S2, and Table S1, Table S2) and referred to in the text. All figures and tables should have titles and legends. All figure legends, tables and supplemental text materials should be placed at the end of the paper. Please note all of these supplemental data should be provided at the time of initial submission and note that the editors reserve the right to limit the size and length of Supplemental Data.

5. Submission Checklist

The Submission Checklist will be useful during the final checking of a manuscript prior to sending it to Drug Discoveries & Therapeutics for review. Please visit Download Centre and download the Submission Checklist file.

6. Online submission

Manuscripts should be submitted to Drug Discoveries & Therapeutics online at http:// www.ddtjournal.com. The manuscript file should be smaller than 5 MB in size. If for any reason you are unable to submit a file online, please contact the Editorial Office by e-mail at office@ddtjournal.com

7. Accepted manuscripts

Proofs: Galley proofs in PDF format will be sent to the corresponding author *via* e-mail. Corrections must be returned to the editor (proof-editing@ddtjournal.com) within 3 working days.

Offprints: Authors will be provided with electronic offprints of their article. Paper offprints can be ordered at prices quoted on the order form that accompanies the proofs.

Page Charge: A page charge of \$140 will be assessed for each printed page of an accepted manuscript. The charge for printing color figures is \$340 for each page. Under exceptional circumstances, the author(s) may apply to the editorial office for a waiver of the publication charges at the time of submission.

(Revised October 2011)

Editorial and Head Office:

Pearl City Koishikawa 603 2-4-5 Kasuga, Bunkyo-ku Tokyo 112-0003 Japan Tel: +81-3-5840-9697 Fax: +81-3-5840-9698 E-mail: office@ddtjournal.com





JOURNAL PUBLISHING AGREEMENT (JPA)

Manuscript No.:

Title:

Corresponding author:

The International Advancement Center for Medicine & Health Research Co., Ltd. (IACMHR Co., Ltd.) is pleased to accept the above article for publication in Drug Discoveries & Therapeutics. The International Research and Cooperation Association for Bio & Socio-Sciences Advancement (IRCA-BSSA) reserves all rights to the published article. Your written acceptance of this JOURNAL PUBLISHING AGREEMENT is required before the article can be published. Please read this form carefully and sign it if you agree to its terms. The signed JOURNAL PUBLISHING AGREEMENT should be sent to the Drug Discoveries & Therapeutics office (Pearl City Koishikawa 603, 2-4-5 Kasuga, Bunkyo-ku, Tokyo 112-0003, Japan; E-mail: office@ddtjournal.com; Tel: +81-3-5840-9697; Fax: +81-3-5840-9698).

1. Authorship Criteria

As the corresponding author, I certify on behalf of all of the authors that:

1) The article is an original work and does not involve fraud, fabrication, or plagiarism.

2) The article has not been published previously and is not currently under consideration for publication elsewhere. If accepted by Drug Discoveries & Therapeutics, the article will not be submitted for publication to any other journal.

3) The article contains no libelous or other unlawful statements and does not contain any materials that infringes upon individual privacy or proprietary rights or any statutory copyright.

4) I have obtained written permission from copyright owners for any excerpts from copyrighted works that are included and have credited the sources in my article.

5) All authors have made significant contributions to the study including the conception and design of this work, the analysis of the data, and the writing of the manuscript.

6) All authors have reviewed this manuscript and take responsibility for its content and approve its publication.

7) I have informed all of the authors of the terms of this publishing agreement and I am signing on their behalf as their agent.

2. Copyright Transfer Agreement

I hereby assign and transfer to IACMHR Co., Ltd. all exclusive rights of copyright ownership to the above work in the journal Drug Discoveries & Therapeutics, including but not limited to the right 1) to publish, republish, derivate, distribute, transmit, sell, and otherwise use the work and other related material worldwide, in whole or in part, in all languages, in electronic, printed, or any other forms of media now known or hereafter developed and the right 2) to authorize or license third parties to do any of the above.

I understand that these exclusive rights will become the property of IACMHR Co., Ltd., from the date the article is accepted for publication in the journal Drug Discoveries & Therapeutics. I also understand that IACMHR Co., Ltd. as a copyright owner has sole authority to license and permit reproductions of the article.

I understand that except for copyright, other proprietary rights related to the Work (*e.g.* patent or other rights to any process or procedure) shall be retained by the authors. To reproduce any text, figures, tables, or illustrations from this Work in future works of their own, the authors must obtain written permission from IACMHR Co., Ltd.; such permission cannot be unreasonably withheld by IACMHR Co., Ltd.

3. Conflict of Interest Disclosure

I confirm that all funding sources supporting the work and all institutions or people who contributed to the work but who do not meet the criteria for authors are acknowledged. I also confirm that all commercial affiliations, stock ownership, equity interests, or patent-licensing arrangements that could be considered to pose a financial conflict of interest in connection with the article have been disclosed.

Corresponding Author's Name (Signature):

Date:

Drug Discoveries & Therapeutics (www.ddtjournal.com)

Pearl City Koishikawa 603, 2-4-5 Kasuga, Bunkyo-ku, Tokyo 112-0003, Japan; E-mail: office@ddtjournal.com; Tel: +81-3-5840-9697; Fax: +81-3-5840-9698